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Stadthagen Gomez, Gustavo; Tehler, Disa Elisabet; Høyland-Kroghsbo, Nina Molin; Wen, Jiayu; Krogh, Anders; Jensen, Klaus Thorleif; Santoni Rugiu, Eric; Engelholm, Lars Henning; Lund, Anders H.

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Loss of miR-10a Activates *Lpo* and Collaborates with Activated Wnt Signaling in Inducing Intestinal Neoplasia in Female Mice

Gustavo Stadthagen1,*, Disa Tehler1, Nina Molin Høyland-Kroghsbo1, Jiayu Wen2, Anders Krogh2, Klaus T. Jensen1, Eric Santoni-Rugiu3, Lars H. Engelholm4, Anders H. Lund1*

1 Biotech Research and Innovation Centre (BRIC), University of Copenhagen, Copenhagen, Denmark, 2 Bioinformatics Centre Department of Biology, University of Copenhagen, Copenhagen, Denmark, 3 Department of Pathology, Diagnostic Centre, Rigshospitalet, Copenhagen, Denmark, 4 The Finsen Laboratory, Rigshospitalet, Faculty of Health Sciences, University of Copenhagen, Copenhagen, Denmark

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Abstract

miRNAs are small regulatory RNAs that, due to their considerable potential to target a wide range of mRNAs, are implicated in essentially all biological process, including cancer. miR-10a is particularly interesting considering its conserved location in the *Hox* cluster of developmental regulators. A role for this microRNA has been described in developmental regulation as well as for various cancers. However, previous miR-10a studies are exclusively based on transient knockdowns of this miRNA and to extensively study miR-10a loss we have generated a *miR-10a* knock out mouse. Here we show that, in the *Apc<sup>min</sup>* mouse model of intestinal neoplasia, female *miR-10a* deficient mice develop significantly more adenomas than *miR-10<sup>−/−</sup>* and male controls. We further found that *Lpo* is extensively upregulated in the intestinal epithelium of mice deprived of *miR-10a*. Using *in vitro* assays, we demonstrate that the primary miR-10a target *KLF4* can upregulate transcription of *Lpo*, whereas siRNA knockdown of *KLF4* reduces *LPO* levels in HCT-116 cells. Furthermore, *Klf4* is upregulated in the intestines of *miR-10a* knockout mice. *Lpo* has previously been shown to have the capacity to oxidize estrogens into potent depurinating mutagens, creating an instable genomic environment that can cause initiation of cancer. Therefore, we postulate that *Lpo* upregulation in the intestinal epithelium of *miR-10a* deficient mice together with the predominant abundance of estrogens in female animals mainly accounts for the sex-related cancer phenotype we observed. This suggests that *miR-10a* could be used as a potent diagnostic marker for discovering signatures of women that are at high risk of developing colorectal carcinoma, which today is one of the leading causes of cancer-related deaths.

Introduction

A growing number of studies show the importance of aberrant miRNA expression in cancer. Although miRNA profiling studies have proven useful in defining signatures of cancer-deregulated miRNAs with diagnostic and/or prognostic value [1,2], establishing casual relationships is not always possible. Altered miRNA expression in cancer can arise from genomic abnormalities but also by alteration of upstream regulators of miRNA expression and/or maturation, including epigenetic silencing [3].

The *miR-10* miRNA family members are encoded in evolutionarily conserved loci within the *Homeobox* (*Hox*) gene clusters of developmental regulators [4,5]. Co-expression of *miR-10* and *Hox* genes during development [6,7] and experimental evidence of *miR-10* targeting of *Hox* transcripts [8–10] has suggested a role for this miRNA family in development. Mammalian *miR-10a* and *miR-10b* are located upstream from *HoxB4* and *HoxD4* respectively and they present a very high degree of sequence conservation, differing at their eleventh nucleotide only (U and A respectively), which thermodynamically enables them to target a fully overlapping set of miRNAs [11,12]. Importantly, both up- and downregulation of *miR-10* has been reported in several cancers and although the number of studies where such deregulation was causally linked to the pathogenesis of cancer remains scarce (for a review, see [4]), some *miR-10* targets have been demonstrated to be mechanistically linked to metastasis, invasion and migration as well as cell proliferation [9,10,13–16].

Colorectal cancer is the second most commonly diagnosed cancer in women and it is one of the leading causes of cancer-related deaths in the world [17]. Colon cancer arises from the epithelial cells of the lumen of the colon where benign adenomatous polyps are established as an initial step. These further progresses into more advanced adenomas showing high-grade dysplasia and can ultimately evolve into invasive cancer. One of the most studied causes of colon cancer is aberrant signaling of the evolutionary conserved Wnt pathway, which is...
Posttranscriptional regulation by microRNA molecules constitutes an important mechanism for gene regulation and numerous studies have demonstrated a correlation between deregulated microRNA levels and diseases, such as cancer. However, genetics studies linking individual microRNAs to the etiology of cancer remain scarce. Here, we provide causal evidence for the involvement of the conserved microRNA miR-10a in the development of intestinal adenomas in the face of activated Wnt signaling. Interestingly, we find that loss of miR-10a mediates an increase in intestinal adenomas in female mice only and delineate the pathway to involve aberrant upregulation of the miR-10a target Klf4 and subsequent transcriptional activation of the Lpo gene encoding the antibacterial protein Lactoperoxidase. Lpo, in turn, has previously been demonstrated to oxidize estrogens into DNA-damaging mutagens.
difference in the level of this close member of the miR-10 family, suggesting no occurrence of dose-dependent compensation via trans-regulation of miR-10b in the absence of miR-10a (Figure S1D). HoxB4 is located 992 nucleotides downstream from the miR-10a gene and the transcription of both genes has been proposed to be co-regulated [6,7]. Deletion of miR-10a did not interfere with transcription of HoxB4 since similar levels of HoxB4 mRNA were detected by quantitative RT-PCR in intestinal samples (Figure S1E).

miR-10a deficiency enhances intestinal tumorigenesis in female ApcMin mice

Since miR-10a inactivation alone did not give rise to increased spontaneous tumor formation, we evaluated if the lack of this miRNA could modify tumor formation upon an additional oncogenic injury. Profiling of WT mouse tissues for miR-10a and miR-10b revealed that miR-10a was relatively highly expressed in the mouse intestinal tract (Figure S2). Furthermore, profiling studies have shown that miR-10a expression is deregulated in human colon cancer [9,24,25]. Therefore, the ApcMin mouse model [27] was chosen to evaluate the role of miR-10a in intestinal neoplasia development. ApcMin mice carry a mutation in the murine homolog of the human APC gene [28] and develop multiple intestinal tubular adenomas similar to those found in patients with the familial adenomatous polyposis syndrome. Furthermore, the ApcMin mouse model is frequently used to evaluate the significance of genetic modifiers [29-32].

To examine the impact of disrupting miR-10a in ApcMin mice, the miR-10a KO and ApcMin mouse strains (both in a C57BL/6 background) were intercrossed. Mice were sacrificed at around 140 days of age for tumor burden evaluation. Strikingly, the mean tumor multiplicity in small intestines of miR-10a KO mice (79.33, n = 15) was almost twice as high as in corresponding miR-10a+/ApcMin age matched controls (41.95, n = 22) (p = 0.0042) (Figure 2A). Tumor multiplicity (TM) and tumor incidence (TI) in the large intestine were also higher in the female mice lacking miR-10a (TM = 2.40, TI = 72.2%) compared to control (TM = 0.82, TI = 54.2%) but those differences were less significant or not significant, respectively (p = 0.14 for males and p = 0.38; Figure 2B). Noteworthy, the miR-10a genotype did not affect tumor multiplicity in ApcMin male mice irrespective of the anatomic location (p = 0.61 for small intestine, p = 0.16 for large intestine; Figures 2A and 2B). The incidence of polyps in the large intestine of male mice also remained unaffected (p = 0.30).

To examine the effect of miR-10a deficiency on tumor size, the flat adenomas of the small intestine were measured at their largest diameter, and this measure was used as indicator of tumor size. No significant difference was observed in mean tumor diameters between miR-10a KO and wild-type ApcMin control mice irrelevant of the gender (p = 0.614 for males and p = 0.071 for females; Figure 2C).

Entire intestinal tracts were paraffin-embedded as “Swiss rolls” and hematoxylin and eosin (H&E) stained sections were examined microscopically based on pathological criteria. Qualitatively, compared to miR-10a+/ApcMin mice intestines, samples from miR-10a−/− ApcMin mice presented more frequently adenomas with high-grade dysplasia and a higher incidence of tubulo-villous adenomas, these differences were more evident in female mice (Figure 2D). However, no invasive carcinomatous processes were observed in any of the analyzed samples. The increased tumor multiplicity and colonic epithelial dysplasia along with unaffected adenoma sizes in the absence of miR-10a, suggest that miR-10a is involved in the tumor initiation/promotion steps but not in enhancing cell proliferation in the ApcMin model of intestinal neoplasia [33,34]. However, due to ethical constraints, the mice are sacrificed at a relatively young age and we cannot formally rule out that an effect on tumor progression would be discernible in the end-stage tumors.

Identification of miR-10a targets in the intestines of female mice

miRNA exert their biological functions primarily by regulating the translation and stability of targeted mRNAs [35,36]. Microarray analysis of deregulated transcripts upon alteration of individual miRNAs in cells and tissues has been proven as a useful tool for identifying direct and indirect miRNA targets [37,38]. Therefore, colon mRNA expression was analyzed in miR-10a KO and WT female mice using Affymetrix microarrays. Although 452 transcripts were significantly deregulated in the miR-10a KO samples compared to WT (P ≤ 0.05), the levels of up- or downregulation were modest and only three protein coding genes had false discovery rates (FDR) lower than 15% (Table S2). In addition, we did not detect any enrichment for predicted miR-10a targets among the deregulated transcripts. Although adaptation to loss of miR-10a or functional redundancy by the remaining miR-10b could account for the invariable levels of miR-10 intestinal targets in the absence of miR-10a, low levels of mRNA deregulation upon miRNA alterations have been previously observed [39,40]. Furthermore, the variation inherent to tissue samples may shadow a high deregulation within a specific cell type of the tissue. With the exception of Lpo, qRT-PCR measurement of selected transcripts, using independent sample sets, did not show any consistent deregulation of genes identified by the microarray as variant between genotypes. Transcript abundance of selected oncogenes and tumor suppressors, relevant in intestinal tumorigenesis or previously predicted as miR-10a targets but not detected in the microarray, were also unchanged in miR-10a KO female mice compared to WT intestines (Figure S3).

Lpo is a secondary target of miR-10a in the intestines of female mice and in human cell lines

Interestingly, Lpo was identified as exceptionally highly upregulated in the intestines of miR-10a KO female mice, displaying a 9.44 fold increase in expression compared to WT (p = 1.1×10−6; adjusted for multiple testing). Lactoperoxidase normally plays a role in antimicrobial defense and removal of toxic hydrogen peroxide [41,42]. However, this enzyme has also been shown to catalyze the activation of endogenous and xenobiotic compounds, such as estrogens and arylamines, into potent depurinating mutagens [43-46]. By increasing genome instability, LPO has been proposed to exert a pro-oncogenic role in tissues like the mammary gland [43,47]. Given the importance of genome stability in the initiation and progression of intestinal tumorigenesis [48], a similar mechanism might be involved in the phenotype observed in miR-10a−/− ApcMin female mice, i.e. the upregulation of Lpo in miR-10a KO mice would enhance estrogen oncogenic activation, leading to a highly instable genomic environment. qRT-PCR confirmed Lpo overexpression by 29-fold in female miR-10a KO intestines compared to WT (Figure 3A). Similar degrees of Lpo upregulation were obtained in male miR-10a KO mice (data not shown). Accordingly, analysis of protein extracts from miR-10a KO and WT intestines equally revealed a strong induction of Lpo in miR-10a KO deficient samples (Figure 3B). In agreement with the analysis of Lpo mRNA and Lpo protein level, a clear difference in both intensity and distribution area of Lpo staining was observed between miR-10a KO and WT mice (p ≤ 0.006, Pearson chi-square test with exact probability). Consistently, all stained WT samples
Figure 2. Disruption of miR-10a leads to enhanced intestinal tumorigenesis in Apc<sup>Min</sup> mice. Tumor multiplicity in the small (A) and large intestines (B) of female and male miR-10a<sup>++/++</sup>;Apc<sup>Min</sup> (WT; n = 22 and n = 19 for each sex) and miR-10a<sup>−/−</sup>;Apc<sup>Min</sup> (KO; n = 15 and n = 16 for each sex) mice; each dot represents data for one mouse. Mean adenoma multiplicities per mouse for each group were: WT = 41.95 and KO = 79.33 for female.
had faint Lpo signal in a limited area of the intestine thus scoring low expression while the majority of miR-10a KO tissue samples had a significantly more intense Lpo signal and a more widespread Lpo expression pattern, thus scoring medium to high (Figure 3C and 3D). No bona fide miR-10a binding sites could be identified in the 3’ UTR of Lpo but cryptic sites in the 5’ UTR and coding sequence (CDS) carried significant complementarity to miR-10a (Figure S4A). To determine whether Lpo was a direct target of miR-10a, via the putative binding sites identified in the 5’ UTR and the CDS of the gene, luciferase reporters holding the 5’ UTR, the entire CDS or a fragment of the CDS containing the most potent binding site were constructed. However, none of the reporters

Figure 3. Lpo is transcriptionally upregulated in the intestines of miR-10a deficient female mice. (A) Lpo mRNA is ~29-fold upregulated in intestines of miR-10a KO compared to WT mice as shown by qRT-PCR. Lpo mRNA levels are normalized to Actb and values ± SD are shown relative to the first WT sample. (B) Western-blot from same tissue samples as in (A) confirming upregulation on protein level. Vinculin was used as loading control. As evident from Lpo and Vinculin control as well as ponceau staining (now shown), sample 4 did not contain any protein for unknown reason. (C) Representative immunohistochemistry staining of Lpo in miR-10a WT and KO intestine. Scale bar 100 μm. (D) Scoring of Lpo expression level estimated by distribution and staining intensity in Lpo stained intestines of WT and miR-10a KO mice. Scoring is divided into low, medium or high expression. Consistent with qRT-PCR and Western blotting analysis a significant difference (p<0.006, Pearson chi-square test with exact probability) in Lpo expression is observed between the different genotypes.

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were affected by co-transfection with a miR-10a duplex (Figure S4B), suggesting that the identified sites were not functional miR-10a targets in this set-up. Further qRT-PCR analysis of LPO transcripts in intestinal samples using primers in intronic and exonic sequence elements, revealed that the primary transcript of LPO was upregulated in miR-10a KO samples to similar levels as the LPO mRNA (Figure S4C and S4D), indicating that Lpo deregulation is transcriptional. Altogether, these results suggest that Lpo is not directly regulated by miR-10a via cognate interaction with target sites in the mRNA but instead that Lpo is regulated at the transcriptional level, probably by one or several primary targets of miR-10a.

The miR-10a target Klf4 is upregulated in miR-10a KO mice and induces transcription of LPO in vitro

We hypothesized that one or more transcription factors under miR-10a regulation could be responsible for enhancing LPO transcription. To identify such transcription factors, we scanned a 2 kb region upstream of the LPO transcription start site (TSS) using Consite and Transfac databases for transcription factor binding site motifs. From the obtained lists of transcription factors, we extracted those that were predicted as putative miR-10a targets by TargetScan [49] and pursued the analysis of one interesting candidate: Krüppel-like factor 4 (KLF4). KLF4, a zinc finger-type transcription factor primarily expressed in the gastrointestinal tract, is an important regulator of differentiation and cell growth arrest of the colonic epithelium and was previously shown to be regulated by miR-10a [50,51]. Upregulation of KLF4 has formerly been observed in early stages of colon carcinoma compared to normal mucosal levels [52]. Using an in vitro setup, in the epithelial-like colon carcinoma cell line HCT-116, we demonstrated a 50% reduction of KLF4 mRNA abundance upon transfection with miR-10a (Figure 4A). Importantly, miR-10a mediated repression of this target was also observed at the protein level (Figure 4B). These results confirm KLF4 as a target of miR-10a as previously described [50].

To investigate the link between KLF4 and LPO we cloned a 1 kb fragment upstream of the TSS of LPO, holding core promoter elements, in front of a luciferase reporter. Co-transfection of this reporter vector with a KLF4 overexpression plasmid in HCT-116 cells resulted in a robust upregulation of luciferase activity, demonstrating the capacity of KLF4 to regulate LPO (Figure 4C and 4D). Using siRNA-mediated knockdown of KLF4 we further linked KLF4 to the regulation of LPO. Although these assays are complicated by a cell density-dependent expression of LPO, we found that knockdown of KLF4 in HCT-116 cells markedly reduced LPO mRNA levels (Figure 4E).

Primary miRNA targets commonly show a relatively low degree of regulation on mRNA level after depletion of the miRNA. It can particularly be difficult to detect these changes in tissue samples that already present a considerable variation in mRNA expression between individuals and within different cell types of the tissue. In these types of experiments it is therefore crucial to use a sufficiently large population to reach statistical significance. In our microarray

Figure 4. Transcription factor KLF4 is regulated by miR-10a and can regulate the LPO promoter in vitro. HCT-116 cells were transfected with a miR-10a duplex or control for 72 h. (A) Relative mRNA levels of KLF4 were measured by qRT-PCR and ACTB was used for normalization. Data are shown as mean ± S.D. of three replicates relative to the control and are representative of three independent experiments. * p < 0.05 using a two-tailed t-test. (B) Protein levels in miR-10a or control transfected cells were assessed by Western-blot using antibodies against KLF4. GAPDH was used as loading control. (C) Western Blot showing the over expression from the pcDNA3.1-KLF4 vector. GAPDH was used as loading control. (D) Luciferase reporter assay in HCT-116 cells (24 h) with pGL4-luc2 holding part of the LPO promoter (1 kb upstream TSS) or the pGL4-luc2 empty vector co-transfected with a vector over-expressing KLF4 or a control vector (pcDNA3.1+). Data are shown as mean ± S.D. of three replicates relative to the pcDNA3.1 transfected control and are representative of eleven independent experiments. **** p < 0.0001 using a two-tailed t-test. (E) HCT-116 cells were transfected with KLF4 siRNA for 48 h or 72 h. Relative mRNA levels of LPO were measured by qRT-PCR and ACTB was used for normalization. Data are shown as mean ± S.D. of three replicates relative to the control and are representative of five independent experiments.
experiment we used intestinal RNA from only 6 animals (*miR-10a KO; n = 3 and WT; n = 3) and we hypothesized that the limited population could account for the lack of detectable *Klf4* deregulation. To increase the statistical power we measured intestinal *Klf4* mRNA levels from 16 *miR-10a* KO and 13 WT mice by qRT-PCR and used four housekeeping genes for normalization. The results showed a significant increase of *Klf4* mRNA levels in the *miR-10a* KO mice compared to the WT (Figure 5A). Consistently, *Klf4* stainings demonstrated a marked increase of protein distribution and intensity in the mouse intestines of *miR-10a* KO compared to WT (Figure 5B and 5C). Specificity of *Klf4* staining was verified by an independent antibody (Abcam; ab131733), which gave a similar expression pattern but resulted in an overall weaker staining (data not shown). Hence, our *in silico*, *in vitro* and *in vivo* results support the existence of a regulatory network linking *miR-10a* to *LPO* via KLF4 and potentially other transcription factors.

**Discussion**

The enormous gene regulatory potential of miRNAs is well demonstrated by many studies showing that perturbed miRNA expression is capable of affecting diverse cellular functions and could ultimately cause disease, including cancer. However, it has been suggested that miRNAs are primarily important in fine-tuning miRNA expression and regulation executed by single miRNAs are in most cases not sufficient to account for pathological phenotypes [40,53]. In line with this, KO of individual miRNAs in other animal models have revealed that most are devoid of obvious phenotypes in the absence of additional lesions or stresses [54,55]. Particular interest in the miR-10 family members arises from their conserved genomic location in *Hox* clusters and the increasing amount of evidence for their implication in vertebrate biology and human disease [4]. Here we addressed the question of the direct causal effect of *miR-10a* in mammalian homeostasis, with a special focus on tumor development, by generating *miR-10a* null mice.

Despite the body of evidence suggesting a role for *miR-10* in *Hox* regulation, the *miR-10a*−/− mice showed an absence of major developmental defects in the posterior trunk. Nevertheless, these results are in agreement with the virtual lack of phenotypic differences upon inhibition or overexpression of *miR-10* during zebra fish development [7]. In the case of *miR-10a*−/− mice, the lack of appreciable phenotypes could be explained by redundancy and functional compensation by *miR-10b*, which levels remained unaffected in *miR-10a* KO mice. A double inactivation of *miR-10a* and *miR-10b* would allow disambiguation of the *miR-10* role in mammalian development.

Interestingly, one gene, Lactoperoxidase, consistently showed an exceptionally high degree of deregulation in the intestines of *miR-10a* deficient mice and to our knowledge this is the first report correlating this gene in to a specific miRNA deficiency. Lpo is mainly described as an antibacterial agent [41,56–58] exclusively found in mucosal surfaces, including colon epithelium [59] and exocrine secretions, like milk, tears, and saliva [60]. Importantly, apart from its antimicrobial and hydrogen peroxide detoxification activities, the role of LPO in carcinogenesis, particularly of the mammary gland, has been intensively studied [43–45,61,62]. In the reduction of peroxides, peroxidases can co-catalyze the oxidation of aromatic and heterocyclic amines into electrophilic metabolites with DNA binding capacity. In this way, Lactoperoxidase can catalyze the mutagenic activation of diverse endogenous and xenobiotic carcinogens, including natural [44,62] and synthetic estrogens [63] as well as other synthetic or environmental aryamines [43,45]. Lpo-mediated activation of such compounds to the derivative quinones and semiquinones, has been shown to induce the formation of the depurinating adducts N3Ade and N7Gua *in vitro* and *in vivo* [44,45,61–63]. Subsequent error-prone

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**Figure 5. Klf4 is upregulated in miR-10a KO intestines.** (A) mRNA levels of *Klf4* were measured by qRT-PCR, *Actb*, *Ubc*, *Hprt* and 36b4 were used for normalization. Data are shown as mean ± S.D. of *miR-10a* KO (n = 16) and WT (n = 13) samples relative to an average of the controls. *p* < 0.05 using a Mann-Whitney test. (B) Representative immunohistochemistry staining of *Klf4* in *miR-10a* WT (n = 5) and KO (n = 8) intestine. Scale bar 100 μm. (C) ViSiomorphDP software scoring of *Klf4* expression level estimated by distribution and staining intensity in *Klf4* stained intestines of WT and *miR-10a* KO mice. *p* = 0.019, student’s t-test.

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base excision repair mechanisms may lead to mutations that can be initiating events in breast, prostate and other types of cancer [47]. Furthermore, estradiol and its catechol metabolites have been shown to induce deletions and loss of heterozygosity in epithelial breast cells resulting in an oncogenic transformed phenotype [64].

Here we showed that Lpo is constitutively upregulated in the intestinal epithelium of miR-10a<sup>-/-</sup> mice and that when these mice were crossed with Apc<sup>Min</sup> females, females displayed a significantly increased tumor burden in their intestinal epithelium. We therefore propose that, in our set-up, upregulated Lpo induces significantly increased tumor burden in their intestinal epithelium. Consistently, this effect would be sex-dependent, since estrogens are intrinsically at higher concentrations in female relative to male mice. Similar to our observations, other genetic studies with Apc<sup>Min</sup> mice have shown that mutation of genes important for genome stability maintenance, generally lead to an increase in adenoma multiplicities [65–68].

Regarding the regulatory mechanism, the Lactoperoxidase mRNA does not contain a miR-10a target site in its 3′UTR. Functional interactions between microRNAs and target sites in other locations than the 3′UTR have been described before, including for miR-10a [69–74]. We therefore tested the functionality of putative miR-10a binding sites in the 5′UTR and coding region of LPO, however, our results led us to exclude the possibility of a direct posttranscriptional regulation of LPO by miR-10a. Instead we obtained evidence supporting a model where LPO expression is regulated by the primary miR-10a target KLF4, which is indeed over-expressed in the intestines of miR-10a KO mice. Putative binding sites for this transcription factor are present in the promoter of LPO, and transcriptional activation of LPO could be reproduced in vitro by over-expressing KLF4. Moreover, siRNA knockdown of KLF4 in HCT-116 cells resulted in downregulation of LPO expression. Of notice, although not tested experimentally, our bioinformatics approach identified other transcription factors representing primary miR-10a targets with putative binding sites in the LPO promoter, suggesting that additional factors may participate in the indirect regulation of LPO by miR-10a. This could, in part, explain the lack of detected variation in the expression levels of direct miR-10a targets (including Klf4) in our microarray experiments, since the occurrence of small degrees of deregulation of primary targets could have a measurable effect only upon convergence in a secondary node. Alternative explanations could be evoked due to the large variation inherent to tissue samples and the fact that the colon tissue comprises a variety of cell types, such as epithelial, luminal and muscular cells, which all have specific genetic programs, thus very different transcriptomes. A change in mRNA expression in one cell type could therefore be masked by the lack of change in another, which could be due to alternative mRNA-independent regulations or a potential rescue by miR-10b. Nevertheless, transcriptomic, bioinformaticcs and biochemical evidence allowed us to reveal a regulatory network where miR-10a can indirectly alter the levels of LPO through KLF4. Interestingly, intestinal miR-10a expression has previously been shown to be downregulated by microbiota in mice [75], and considering the antibacterial functions of Lpo in innate immunity, it is alluring to suggest that miR-10 could function as the sensor of immune stimuli in this environment where its downregulation would induce Lpo as an antibacterial mechanism in normal epithelium. However, having such a defense program constantly activated in the presence of estrogen, as would be the case in the miR-10a<sup>-/-</sup> female mice, would ultimately be damaging for the cells.

Our results are in contrast with reports of miR-10a upregulation in colon cancer samples [24,25]. Such upregulation could correspond to a consequence rather than a cause of oncogenic transformation. This is enforced by the observation by Monzo et. al. showing an upregulation of miR-10a in stage II but not in stage I colon cancer samples [24], though the pathogenic role of miR-10a upregulation in advanced colon cancer remains to be elucidated. Moreover, Klf4 has been described to inhibit cell growth and play a tumor suppressive rather than an oncogenic role in colorectal cancer [51]. However, as suggested in our study, oncogenic downregulation of miR-10a might be a very early event promoting cellular transformation by a similar mechanism to the one previously described for Lpo in mammary carcinoma [44]. Furthermore, and as mentioned above, not only Klf4 but likely other primary targets of miR-10a are also involved in the regulation of Lpo.

In summary, here we present evidence that miR-10a, through a complex regulatory network involving the transcription factor Klf4, can contribute to tumor formation in female mice. By the indirect upregulation of Lpo levels in the intestinal epithelium, miR-10a deficiency in these mice creates an environment where estrogen could be transformed into potent depurinating mutagens that can ultimately lead to the initiation of cancer and tumor formation. Therefore we suggest that miR-10a may serve as a potential diagnostic marker for identifying groups of women that are at high risk of developing colorectal cancer.

Materials and Methods

Generation of miR-10a neo/+ ES cells

The miR-10a targeting plasmid was constructed using standard recombinant techniques. R1-129 mouse ES cells were electroporated with the linearized targeting plasmid. G418 clones were selected and grown independently. Each clone was genotyped by southern blotting using the probes 5′/PacI and 3′/NsiI (Table S1) and P<sub>ac</sub>- or N<sub>si</sub>-digested DNA to verify 5′and 3′recombination respectively.

Ethics statement

All mice were handled according to good animal handling practices and the animal experiments approved by the Danish Animal Experiments Inspectorate.

Mice

miR-10a KO mice were generated as follows. ES cells from one of the three screened and correctly targeted clones were microinjected in C57BL/6 blastocyst and implanted in foster mothers. The resulting germline chimeras were bred with C57BL/6 (B6) to generate heterozygous mice for the targeted allele. F1 miR-10a<sup>−/−</sup> males were crossed to a ubiquitously expressing Cre mouse line to eliminate the neo resistance cassette by Cre-LoxP recombination. Mice carrying the miR-10a floxed allele (miR-10a<sup>−/−</sup>) were intercrossed with B6 mice for at least 7 generations before generating experimental cohorts of homozygous mutant and control mice. All mice were genotyped by PCR using tail-tip DNA and primers described in Figure 1A and Table S1, PCR conditions were 95°C for 5 min followed by 35 cycles of denaturation at 95°C for 30 sec, annealing at 56°C for 30 sec, extension at 72°C for 1 min and a final extension step at 72°C for 7 min. Apc<sup>Min</sup> mice were obtained from The Jackson Laboratories and are described elsewhere [27,28]. Only males were used for breeding with miR-10a<sup>−/−</sup> or miR-10a<sup>-/-</sup> mice. Genotyping of the
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*Min* allele was performed by PCR using the primers APC-fw, APC-rw and APC-Min under the same conditions described above. All primer sequences are shown in Table S1. Mice had free access to food and water.

**Cell culture**

HCT-116 cells were maintained in McCoy’s 5A medium (Gibco) with 10% fetal bovine serum (Thermo Scientific) and 1% penicillin/streptomycin (Invitrogen), incubated at 37°C in 5% CO2.

**Quantitative RT-PCR**

Total RNA was isolated from mice tissues with TRIzol (Invitrogen), as specified by the manufacturer, followed by DNase I treatment with the DNA-free kit (Applied Biosystems). HCT-116 cells were seeded in 6-well plates and reversively transected with 50 nM of Allstars negative control (Qiagen; cat:1027281), a miR-10a duplex (Ambion; AM17100, PM10787) or siRNAs against *KLF4* (pool of two siRNAs with sequences: CUCUACACAUGAA-GAAGGCAait[IT]a[IT], GUGGGAUAACAGGGUAUAAita[IT]a[IT], 25 nM each) using Lipofectamine2000 (Invitrogen). Cells were harvested 48 h or 72 h post-transfection for total RNA extraction using the mirNeasy Kit (Qiagen; 217004).

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**Microarray analysis**

Total RNA was extracted from colon samples of 4 months old, WT and *miR-10a*−/− female mice. Organs were collected as described in the previous section but after PBS washing, samples were frozen in liquid nitrogen and kept at −80°C until RNA was extracted. Four biological replicates of each genotype were analyzed on Affymetrix microarrays (GeneChip Mouse Gene 1.0 ST Array) at the Microarray Centre, Rigshospitalet, Copenhagen University Hospital as previously described [76]. Affymetrix probe set intensity of miR-10a KO and WT samples were preprocessed using the aroma package in BioConductor, including steps of background correction, normalization, and summarization by RMA (Robust Multichip Average) method.

We then applied a non-specific filtering step to exclude those genes showing low overall expression levels, as these genes were unlikely to show down- or upregulation after miRNA transfection. To do this, we required the interquartile range of probe set expression levels to be greater than the first quartile value of the interquartile range of expression levels for all probe sets. The duplicated probe sets mapped to the same genes and the probe sets without entrez gene annotation were also removed by this step. The remaining probe sets were subsequently mapped to gene symbols using the Affymetrix mogene10transcriptcluster.db annotation Package. Differentially expressed genes were identified using moderated t-test with P-value less than 0.05 and log FC > 2. Expression differences were calculated using the biomaRt package in BioConductor. We defined three datasets: upregulated set (296 transcripts) with P-values < 0.05 and log FC > 2, upregulated set (156 transcripts) with P-values < 0.05 and log FC < 0, and no change set containing 323 transcripts from the gene set with P-value near 1.

**Western blot analysis**

HCT-116 cells were seeded in 6-well plates and reverse transected with 50 nM Allstars negative control (Qiagen; Cat: SK-4800). Stained sections were scanned using a NanoZoomer-2.0HT (Hamamatsu, Denmark) using 40× magnification. Scanned sections were evaluated, blinded in respect to genotype of mice, by low, medium or high LPO expression. Comparison of LPO expression between *miR-10a*+/+ (n = 5) and *miR-10a*−/− (n = 10) mice was done by Pearson Chi-square test with exact probability and the analysis revealed a significant scoring difference (P =< 0.006) between the two genotypes.

For immunohistochemical detection of Klf4 a polyclonal goat-anti-mouse Klf4 antibody (R&D; AF3158) was used diluted 1:80 (2.5 μg/ml) where chromogen staining was achieved using the EnVision+ system (Dako, Glostrup, Denmark, K4003) in combination with NovaRED HRP substrate (VWR international, Herlev, Denmark, SK-4800), only 1/3 of normal hematoxylin stain was used to prevent a too strong blue nuclear signal that could bias the analysis. Stained sections were scanned using a NanoZoomer-2.0HT (Hamamatsu, Denmark) using 40× magnification. To quantify the expression level of Klf4, whole scanned sections were analyzed by the staining analysis software VisiophormDP, which is part of the Visiopharm software package (Visiopharm, Horsholm, Denmark). The formula Apositive/(Apositive+Anegative)*(255-Imean) was used as a measure of the Klf4 expression. Aabsolute = area of positive cells nuclei, Aabsolute = area of Klf4 negative cell nuclei and Imean = the mean intensity value (0–255, where the darkest colors have the lowest value) of the separating color band (ref). Comparison of Klf4 expression between miR-10a+/+ (n = 5) and miR-10a−/− (n = 8) mice was done by Students t-test and the analysis revealed a significant scoring difference (p = 0.019) between the two genotypes.

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1027281) or a miR-10a duplex (Ambion; AM17100, PM10787) using Lipofectamine2000 (Invitrogen). Cells were harvested 72 h post-transfection for protein extraction.

Cells or tissues were lysed and homogenized in RIPA buffer (150 mM NaCl, 0.5% DOC, 0.1% SDS, 1% Igepal, 50 mM Tris-HCl pH 8, 2 mM EDTA) containing 1 mM Pefabloc (Roche Applied Science) and 1× Complete Mini protease inhibitor mixture (Roche Applied Science), 20 μg of protein per lane from was separated on a 4–12% NuPAGE Bis-Tris gel (Invitrogen) for cell culture experiments and a 3–8% Tris-Acetate gel (Invitrogen) for mouse tissues, followed by transfer to a nitrocellulose membrane. Membranes were blocked in 5% milk for 40 min at room temperature and incubated over night with primary antibody at 4°C. Antibodies used were purchased from Santa Cruz: LPO (H-60) sc-134848), KLF4 (H-180; sc-20691).

Luciferase reporter assays and vectors
A 1 kb sequence upstream the transcription start site of the LPO gene was cloned into pGL4.1uc2 (Promega) using the following primer sequences (restriction sites XhoI and NotI are shown in lowercase letters): Fwd: 5'-CTCTGgATTTGCTGGATACTCAGC-3', Rev: 5'- TgCtgAgCTGCAGCAAGTTGTGCCCC-3'. The KLF4 over expressing vector was purchased from Addgene (pcDNA3.1-HA-KLF4-FL; plasmid #34593). HCT-116 cells were seeded in 96-well plates (15 000 cells/well) and transfected the next day with 50 ng of pGL4.1uc-2.LPO promoter or pGL4.1uc2 (empty) and 50 ng of pcDNA3.1-HA-KLF4-FL or pcDNA3.1+ (empty) using Lipofectamine2000 (Invitrogen). Luciferase expression was measured 24 h after transfection using the Dual-Glo luciferase assay (Promega, E2940). For investigating the potential miR-10a binding sites part of the 5’UTR of Lpo was cloned into psi-CHECK-2 (Promega) using primers (restriction sites NotI is shown in lowercase letters): Fwd-5’UTR: 5’-GACgtgAcATCATCCTGGACTCAGT-CCT-3’, Rev-5’UTR: 5’-CGTgtgACAAAGGgACACAT- CAGGTCGA-3’, and either the whole coding sequence (CDS) or a 609 bp sequence harboring the best miR-10a CDS binding site was cloned into a Firefly luciferase fusion vector pPK-CMV-F4 (PromoKine) using primer sequences and HindIII (restriction sites NheI and the 5’UTR and CDS). (B) Luciferase reporter assay in HCT-116 cells (24 h) with constructs holding either the Lpo coding sequence (CDS), part of Lpo CDS including the best miR-10a putative binding site or the LPO 5’UTR co-transfected with a Renilla expressing vector along with a miR-10a duplex or control. Data are shown as mean ± S.D. of three replicates relative to the control and are representative of three independent experiments. n.s. using a two-tailed t-test. (C) Lpo primary transcript levels, using primers in Lpo intron 9, and (D) Lpo mature mRNA levels, using primers in Lpo exon 7 and 8, was measured by qRT-PCR. Actb was used for normalization and values ± SD are shown relative to the first WT sample. (TIF)

Supporting Information

Figure S1 Basic phenotypic characterization of miR-10a deficient mice. (A) miR-10a−/− allele segregation in the offspring of miR-10a−/− mice. (B) Weight of miR-10a+/− (WT males n = 17, females n = 24) and miR-10a−/− (KO, males n = 21, females n = 27) mice at 8 weeks of age. Detection of mature miR-10a (C) and miR-10b (D) by qRT-PCR in intestines of miR-10a−/− (WT) and miR-10b−/− (KO) mice. (E) Relative expression levels of HoxB4 miRNA in intestines from mice of the indicated genotypes. Circles represent values from female mice and squares represent data from male mice. At least three different mice of each genotype were analyzed in three independent experiments. Results from one representative experiment are shown. (TIF)

Table S1 Probes and primers used for mouse genotyping. (PDF)

Table S2 Deregulated genes in the intestines of miR-10a−/− mice. 432 genes up- or downregulated with an FDR smaller than 0.05 before correction are shown. Genes are ordered by Fold change. (XLSX)

Table S3 Primers used in qRT-PCR. (PDF)

Author Contributions

Conceived and designed the experiments: GS DT NMHK AHL. Performed the experiments: GS DT NMHK KTJ LHE. Analyzed the data: GS DT NMHK JW AK ESR LHE AHL. Wrote the paper: GS DT. Performed the experiments: GS DT NMHK KTJ LHE. Analyzed the data: GS DT NMHK JW AK ESR LHE AHL. Wrote the paper: GS DT AHL.


