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A "grappling hook" interaction connects self-assembly and chaperone activity of Nucleophosmin 1

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Abstract

How the self-assembly of partially disordered proteins generates functional compartments in the cytoplasm and particularly in the nucleus is poorly understood. Nucleophosmin 1 (NPM1) is an abundant nucleolar protein that forms large oligomers and undergoes liquid–liquid phase separation by binding RNA or ribosomal proteins. It provides the scaffold for ribosome assembly but also prevents protein aggregation as part of the cellular stress response. Here, we use aggregation assays and native mass spectrometry (MS) to examine the relationship between the self-assembly and chaperone activity of NPM1. We find that oligomerization of full-length NPM1 modulates its ability to retard amyloid formation in vitro. Machine learning-based structure prediction and cryo-electron microscopy reveal fuzzy interactions between the acidic disordered region and the C-terminal nucleotide-binding domain, which cross-link NPM1 pentamers into partially disordered oligomers. The addition of basic peptides results in a tighter association within the oligomers, reducing their capacity to prevent amyloid formation. Together, our findings show that NPM1 uses a "grappling hook" mechanism to form a network-like structure that traps aggregation-prone proteins. Nucleolar proteins and RNAs simultaneously modulate the association strength and chaperone activity, suggesting a mechanism by which nucleolar composition regulates the chaperone activity of NPM1.

Keywords: native mass spectrometry, molecular chaperones, amyloid formation, membraneless organelles

Significance statement:

Membraneless organelles perform specific tasks inside the cell, yet it is unclear how assemblies of disordered proteins give rise to functional structures. A prominent example is the nucleolus, which prevents protein aggregation under stress and controls ribosome assembly. Here, we investigate the scaffold protein Nucleophosmin 1 (NPM1), which controls the liquid-like properties of the nucleolus. We find that NPM1 self-assembles through fuzzy interactions of its acidic tracts with its C-terminal chaperone domain. Ribosomal proteins and RNA tighten the association of NPM1 oligomers, blocking access to the C-terminus. Our study shows how self-assembly and chaperone function can be controlled simultaneously by involving the same domain. This principle reveals a blueprint for the "structure—function"-based regulation of membraneless organelles.

Introduction

The nucleolus, the site of ribosome biogenesis in the nucleus, is a membraneless compartment that responds to changes in cellular growth rate, metabolic activity, and stress (1). Its dynamic nature allows a constant exchange of proteins and nucleotides with the surrounding nucleus and the cytoplasm. For example, the nucleolus sequesters p14ARF and its binding partner human double minute 2 homolog (HDM2), which otherwise ubiquitinylates the tumor suppressor p53 to allow cell cycle progression (2). Its ability to recruit and release proteins has been attributed to the fact that nucleolar assembly is driven by liquid–liquid phase separation (LLPS) of a highly enriched subset of nucleolar proteins (3). By exhibiting different phase-separating properties, these proteins account for the coexistence of nucleolar regions with distinct functions (4). Nucleophosmin 1 (NPM1, also known as B23) is the main component of the outermost nucleolar phase, the granular com-

Competing Interests: The authors declare no competing interests

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ponent, where ribosomes are assembled. Mutations in NPM1 are commonly associated with acute myeloid leukemia (AML) (5). It is the main nucleolar interaction partner for the p14ARF tumor suppressor and the c-MYC oncoprotein, and knockdown experiments have identified NPM1 as a promising target for cancer therapy (6, 7).

NPM1 has a modular structure with folded N-terminal and C-terminal domains (NTD, residues 1 to 120, and CTD, residues 240 to 294) linked by an intrinsically disordered region (IDR, residues 120 to 240) (Fig. 1a). The N-terminal domain adopts a &-sheet sandwich fold and assembles into pentamers that can form end-to-end decamers (8). The NTD and IDR contain three highly acidic poly-E stretches (A1, A2, and A3), as well as two regions with predominantly basic residues (B1 and B2) (9). The CTD is a nucleotide-binding domain composed of three &-helices and is the site of AML-related mutations, which result in removal of a nucleolar localization signal (10). Full-length (FL) NPM1 self-assembles into large oligomers in isolation (9, 11) and in cancer cells (12). The addition of polyanionic or polycationic molecules, such as basic peptides or RNA, induce LLPS through heterotypic interactions with the opposing charges on FL NPM1 (9). The isolated NTD can also undergo LLPS by binding basic peptides via its acidic A1 region (13). In addition, oligomeric NPM1 engages in homotypic interactions through contacts between its charged regions, which can give rise to LLPS under crowding conditions and are modulated by ionic strength (9, 14).

Besides its role as a nucleolar scaffold, NPM1 has a chaperone activity in vitro and effectively prevents the aggregation of denatured proteins (15). Strikingly, the nucleic protein is enriched in disordered and aggregation-prone proteins (16). The nucleolus can sequester misfolded proteins during cellular stress and turn them over to HSP70 for refolding (17). It also stores protein aggregates in amyloid bodies that may function as “nucleolar detention centers” for potentially toxic aggregates (18). In cells, NPM1 colocalizes with misfolded proteins and amyloids, suggesting that it functions as a chaperone during cellular stress (17) (Fig. 1b). Importantly, deletion of individual domains of NPM1 reduces its ability to counteract thermal denaturation of proteins (11). Together, these findings raise the possibility that the chaperone function of NPM1 in the nucleolus is related to its self-assembly properties.

Results
Full-length NPM1 and its CTD display differential chaperone activity toward amyloid formation

Previous studies have established the ability of FL NPM1 to protect globular proteins from thermal and chemical denaturation (11, 15). However, several amyloidogenic proteins are targeted to the nucleus (19), and NPM1 associates with nuclear amyloid aggregates in vivo (17). We, therefore, tested the ability of NPM1 to prevent the aggregation of amyloid-β42 (Aβ42), which has been found in the nuclei of neurons from Alzheimer’s disease patients and is considered a model for amyloid formation (20, 21). It has to be noted that Aβ42 has no confirmed nucleolar localization and is not likely to be a physiological target of NPM1, but is widely used as a model system to study the generic anti-amyloid activity of chaperone proteins (22, 23).

Briefly, we incubated 3 μM Aβ42 with 0 to 6 μM NPM1 in the presence of the amyloid-specific dye Thioflavin T (ThT) and monitored fibril formation through the increase in fluorescence intensity (Fig. 1c). The addition of NPM1 delayed the half-time of fibrillation in a concentration-dependent manner, with the most pronounced effect at an NPM1 : Aβ42 ratio of 1:1 (Fig. 1c). The end-point fluorescence intensity was in all cases comparable to that of Aβ42 alone (Fig. S1a). Interestingly, NPM1 does not appear to specifically affect only primary or secondary nucleation, or fibril elongation, but rather all of these processes. Together, these observations suggest that NPM1 efficiently retards the amyloid assembly processes. To identify which parts of the modular NPM1 structure are responsible for delaying Aβ42 aggregation, we designed truncated variants: NPM1120, which contains only the NTD with the A1 tract, NPM1188, which includes the NTD and the acidic tracts A1-3, NPM1240, which encompasses the NTD and the entire IDR, and NPM1240-294, which represents the isolated CTD (Fig. 1d). We then tested the effects of all NPM1 variants on Aβ42 fibrillation under the same conditions as for FL NPM1 and determined the half-life of Aβ42 fibrillation (t1/2). Plotting the Aβ42 t1/2 in the presence of FL NPM1, NPM1120, NPM1188, and NPM1240 shows that full-length NPM1 affects Aβ42 aggregation in a concentration-dependent manner, whereas the C-terminally truncated variants have only minor effects (Fig. 1e). NPM240-294, i.e., the isolated CTD, a dose-dependent delay in ThT fluorescence at a concentration of 0.3 μM, and additionally resulted in a strong suppression of ThT fluorescence when the concentration was raised further (Fig. 1f). Strikingly, none of the other NPM1 constructs resulted in a similar decrease in end-point fluorescence (Fig. S1a). In agreement with the ThT data, electron microscopy showed no fibrils in the presence of the CTD at the reaction end-point. (Fig. S1b). Together, these observations indicate that the ability of NPM1 to delay fibril formation is related to its CTD but appears to be reduced in the FL protein. To test whether NTD and CTD work synergistically to delay Aβ42 fibrillation, we incubated Aβ42 with 3 μM NPM1 and 3 μM NPM1240-294. However, the combined effect of both NPM1 parts on fibril formation was far less pronounced than for FL NPM1 at the same concentration (Fig. S1b). This finding shows that the chaperone activity of the CTD is reduced in the presence of the NTD, but the effect is less pronounced when the NTD is part of the same polypeptide than when it is added in trans. These observations imply that inter-domain interactions in the FL protein regulate its chaperone activity towards Aβ42.

A covalent link between NTD and CTD is required for NPM1 self-assembly

To better understand the relationship between chaperone activity and interdomain interactions, we turned to native mass spectrometry (MS). Here, intact protein complexes are transferred from the solution to the gas phase using soft electrospray ionization (nESI). Since noncovalent interactions can be preserved during mass measurements, we can obtain information about the oligomeric states of protein complexes in solution (24, 25). In combination with collision-induced dissociation, where the intact complexes are subjected to high-energy collisions with an inert buffer gas inside the mass spectrometer, we can determine their composition. By comparing the collision voltages required to dissociate protein interactions, we can furthermore assess their relative stabilities (26) (Fig. 2a).

First, we subjected FL NPM1 to native MS analysis (Fig. 2b). Under gentle MS conditions (collision voltage 25 V), we could only detect an unresolvable “hump” that is characteristic of large assemblies with no well-defined stoichiometry (27). When raising the collision voltage to 75 V, we obtained well-resolved peaks corresponding in mass to monomers, dimers, trimers, and pentamers of FL NPM1. We did not detect higher oligomeric states than pentamers and consider that the monomers, dimers, and trimers

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Fig. 1. NPM1 delays Aβ42 aggregation. (a) NPM1 has a modular architecture, composed of a folded N-terminal pentamerization domain (NTD, residues 1 to 120), an intrinsically disordered region (IDR, residues 120 to 240) containing three acidic and two basic tracts (A1-3 and B1 and 2, respectively), and a C-terminal nucleotide-binding domain (CTD, residues 240 to 294). The NTD and the CTD are rendered based on PDB IDs 2P1B and 2VXD, respectively. (b) NPM1 pentamers associate with RNA, basic transcription factors, and ribosomal proteins to form the granular component of the nucleolus. Cellular stress induces the formation of nucleolar amyloid bodies. (c) The presence of increasing amounts of NPM1 shows that NPM1 delays the onset and the elongation of fibrillation, as judged by ThT fluorescence. Error bars indicate the SD of \( n = 4 \) experiments. (d) Truncated variants of NPM1 used in this study: NPM1120 encompasses the NTD. NPM1188 the NTD and the acidic regions of the IDR, NPM1240 the NTD and the entire IDR and NPM1240-294 only the CTD. (e) Fibrillation half-times (t1/2) of Aβ42 in the presence of 0 to 3 \( \mu \)M of NPM1 variants shows that only FL NPM1 and NPM1240-294, but no other truncated variants, affects fibrillation. The t1/2 of Aβ42 alone is shown as a dashed line. Error bars indicate the SD of \( n = 4 \) experiments. A one-way ANOVA was used to test for the overall significant difference between groups before running pairwise t-tests with Bonferroni correction. Asterisks indicate significant differences (\( P < 0.001 \)). (f) ThT fluorescence curves of Aβ42 in the presence of 0 to 3 \( \mu \)M NPM1240-294 show a dose-dependent delay in fibrillation and a decrease in fluorescence intensity. Error bars indicate the SD of \( n = 4 \) experiments.

likely are dissociation products. These findings indicate that FL NPM1 pentamers assemble into large oligomers, in agreement with previous reports (9, 11, 12). Next, we analyzed oligomerization of the truncation variants. NPM1120 yielded well-resolved spectra at a collision voltage of 50 V. Strikingly, we observed a broad range of oligomeric states, ranging from 5 to 50 subunits, but always in multiples of five, suggesting that the NTD pentamers can assemble into polymers (Fig. 2b). The charge state distribution of a protein ion is dependent on its surface area (28, 29), allowing structural information to be extracted. Plotting the average charge of each oligomeric state as a function of its molecular weight results in a correlation, which closely follows the trend expected for globular proteins (Fig. S2a). The crystal structure of NPM1120 shows side-by-side and end-to-end association of NTD pentamers in a crystal lattice via salt bridges (Fig. S2b), which leads us to speculate that crystal-like interactions drive multimerization of the NTDs. MS analysis of NPM1188, which includes the IDR region with acidic tracts A2 and A3 required moderately higher collision voltages (75 V). The resulting mass spectra show pentamers and decamers, but not any higher oligomeric states, indicating that the IDR disrupts the self-association propensity of the isolated NTDs. Native MS analysis of NPM1240, which includes the entire IDR, yielded essentially the same oligomeric state as NPM1188 but with a slightly lower amount of decamers. Last, we also examined the isolated CTD, NPM1240-294. Native mass spectra show monomeric protein, with no sign of higher oligomerization besides traces of dimers (Fig. 2d). Taken together, native MS reveals three types of NPM1 oligomerization, which are governed by the three domains: (1) the isolated NTD forms pentamers that self-assemble into ordered multimers; (2) the IDR disrupts these NTD multimers, yielding pentameric protein; (3) including the CTD induces the formation of large oligomers which can be dissociated into pentamers and pentamer fragments. We conclude that the formation of higher NPM1 oligomers requires full-length protein, suggesting direct interactions involving the CTD.

NPM1 pentamers connect via their CTDs

The observations from MS suggest that the CTD is involved in the higher oligomerization of NPM1. We therefore used fluorescence spectroscopy to test for interactions between the CTD and other regions of the protein. Importantly the only two tryptophane residues in NPM1 are in the CTD, which allowed us to use intrinsic fluorescence to probe its interaction in solution. We found that
Fig. 2. Native MS shows that higher oligomerization is specific for NPM1FL. (a) A schematic illustration of the native MS experiment, using collision-induced dissociation in the collision cell of the MS instrument. The illustration has been adapted from (43). (b) The native mass spectra of NPM1 at low collision energy show unresolved baseline humps, indicating large oligomers (top). Increasing the collision voltage releases NPM1 monomers, dimers, trimers, and pentamers (bottom). (c) The mass spectra of NPM1120 show a range of oligomeric states composed of multiples of five subunits, as indicated. NPM188 and NPM1240 form predominantly pentamers and minor decamer populations. (d) The isolated CTD (NPM1240-294) exists nearly exclusively as monomers. The respective collision voltage at which each spectrum was obtained is indicated on the right.

The addition of quadruplex DNA, a high-affinity ligand for the CTD (30), quenches the intrinsic fluorescence of NPM1240-294, indicating binding (Fig. 3a). We then tested whether the N-terminal region of NPM1 affected tryptophan fluorescence. The addition of NPM1120 resulted in a mild decrease in fluorescence intensity, whereas the addition of NPM1240 had a more pronounced effect. The presence of both DNA and NPM1240 gave an intermediate fluorescence reduction. Taken together, the change in tryptophan fluorescence suggests a direct association between NPM1240-294 and NPM1240, which is impacted by the presence of DNA.

To better understand how the CTD associates with other parts of NPM1, we turned to cryo-electron microscopy (cryo-EM). Using physiological conditions (pH 7.5 and 150 mM NaCl), we obtained electron density maps that allowed a reconstruction of the NTD with a resolution of 2.6 Å (Fig. S3, Table S1). Comparison to the crystal structure revealed a virtually identical fold, including sidechain orientations, except for the loop covering residues 34 to 39, which makes up the A1 tract. In the X-ray structure, it is stabilized via crystal contacts with the neighboring pentamer (Fig. S2). In the cryo-EM density maps, the loop is too flexible for a confident reconstruction, suggesting that not only the A2 and A3 tracts but even the A1 tract in the NTD is disordered (Fig. 3b). Free CTDs attached to the disordered linker are too small (less than 9 kDa) to be resolved on their own. Interestingly, we obtained two distinct 3D classes, one with an additional density above the acidic side of the pentamer, off-center from central cavity (Fig. 3c). Strikingly, nearly all particles in this class had another NTD pentamer in the proximity (Fig. S4a). Particles in the second class, on the other hand, did not show the extra density and had less nearby particles (Fig. 3d and Fig. S4b). The extra density is connected to the pentamer via the A2 tract of a single NTD and can therefore not be attributed to the disordered regions of each protomer (Fig. S4c). We speculated that the density could stem from interactions involving the CTD.

To test this possibility, we turned to AlphaFold (AF), a neuronal network that can predict the 3D structures of protein complexes with an accuracy that rivals experimentally determined structures (31, 32). Capitalizing on the ability of AF to dock short, disordered peptide segments with high confidence (33), we divided the IDR of NPM1 into peptides of 20 amino acids with a 10-residue overlap and predicted possible complexes with the CTD (Fig. 3e, Fig. S5). We then calculated the binding energies for the top-scoring complex for each peptide. We found that peptides covering the acidic tracts A2 or A3 exhibited weakly favorable interactions due to charge contacts with the basic residues in the CTD. Placement of the top-scoring complex between the CTD and the A2 tract into the EM map showed a good agreement between the unassigned density, the helical acidic region of the IDR, and the approximate location of the CTD (Fig. S4d). Considering the flexibility of the IDR,
Fig. 3. An NTD–CTD interaction in FL NPM1. (a) The fluorescence of tryptophane residues 288 and 290 in NPM240–294 is quenched by the addition of equimolar amounts of DNA or NPM240, and to a lesser extent by NPM120. In the presence of DNA and NPM240, intermediate quenching is observed. Error bars indicate the SD of \( n = 6 \) repeats. One-way ANOVA was used to test for the overall significant difference between groups before running pairwise t-tests with Bonferroni correction. Asterisk indicates significance (\( ** P < 0.01 \) and \( *** P < 0.001 \)). (b) The cryo-EM reconstruction of the NTD (gray) shows virtually no deviations from the crystal structure (PDB ID 5EHD, blue), except for the A1 tract (residues 34 to 39), which could not be modeled based on the density map. (c) The cryo-EM density map for FL NPM1 reveals an additional asymmetric density (green) above the NTD pentamer (gray). (d) A second 3D class with fewer neighboring particles does not show the additional density. (e) The helical A2 tract (orange) appears as a diffuse density in the EM map. AF predicts a complex between the helical A2 tract (orange) and the basic CTD (rendered as an electrostatic surface). (f) Native MS of the NPM1\_IDR variant lacking the disordered region between residues 120 and 240 reveals pentamers and a small fraction of decamers but no higher oligomers.

The CTD would be unlikely to occupy a more specific orientation, giving rise to a more diffuse density than the pentameric NTD.

We then confirmed interaction between the disordered region and CTD by recording mass spectra of NPM1\_\_IDR and NPM1\_240–294. We observed unresolvable peaks and a shift to the higher m/z region for NPM1\_\_IDR, indicating binding of the CTD with a mixed stoichiometry (Fig. S6a). Next, we designed a short NPM1 variant, NPM1\_ALM, in which NTD and CTD are linked without the IDR (Fig. 3f). Native MS analysis of NPM1\_ALM revealed almost exclusively pentameric protein and a small decamer population, irrespective of collision voltage (Fig. 3f). This result demonstrates that removing the IDR prevents higher oligomerization of NPM1 pentamers.

Taken together, fluorescence spectroscopy, AF, cryo-EM, and MS suggest that the CTD engages in weak, “fuzzy” interactions with the disordered region of neighboring pentamers. The IDR and CTD can be viewed as a “grappling hook” that links NPM1 into large oligomers.

Since the interactions between CTD and IDR affect nucleotide binding to NPM1, we recorded mass spectra of FL NPM1 in the presence of equimolar amounts of tRNA, which binds to NPM1 and is more homogeneous than rRNA and thus easier to detect by MS. We found that tRNA completely abolishes the NPM1 signal in MS even at high collision voltages (Fig. S6b), consistent with tRNA cross-linking the NPM1 pentamers into oligomers that are too stable for gas-phase dissociation. We also considered AML mutations that disrupt the third helix of the CTD and asked whether they may impact self-assembly. We purified an NPM1 variant with a mutation at the C-terminus (NPM1\_AML) in which the last seven residues (WQWRKSL) are exchanged for an 11-residue sequence lacking the nucleolar localization signal (CLAVEEVSLRK) and recorded mass spectra under native conditions. Strikingly, we
did not observe a pronounced difference between FL NPM1 and NPM1AML (Fig. S6c). We conclude that the AML variants do not exhibit altered self-assembly properties, and that its oncogenic potential can therefore be attributed to loss of the nucleolar localization signal in helix 3.

**Nucleolar components modulate NPM1 self-assembly and chaperone activity**

Having established that CTD interactions mediate NPM1 self-assembly, we then asked whether the same interactions affect its ability to delay A\(_{\beta}\)\(_{42}\) fibrillation. It is well established that NPM1 can undergo LLPS through heterotypic interactions with RNA or basic peptides located in the nucleolus. Peptides such as residues 21 to 37 of ribosomal protein L5 (rpL5) or residues 299 to 326 of SURF6, which both contain multiple arginine-rich motifs, bind to the A1 tract of NPM1 and induce LLPS (13). The same type of interaction has been found to mediate sequestration of p14\(^{ARF}\) by nucleolar NPM1 (34, 35). We speculated that such interactions would compete with higher oligomerization via the CTD.

To test this possibility, we selected rpL5\(_{21-37}\), a short peptide with two basic motifs and whose association with NPM1 has been investigated in detail (Fig. 4a) (13, 36). The AF model shows binding of the second basic motif of rpL5\(_{21-37}\) to the acidic A1 groove on the NTD, while the first basic motif is extended away from the protein, making it accessible for charge interactions with another NTD (Figure S7c). This model, in agreement with previous reports (13), thus suggests that rpL5\(_{21-37}\) cross-links acidic regions in NPM1. We monitored the effect of rpL5\(_{21-37}\) on NPM1 self-assembly by incubating FL NPM1 for 60 min at room temperature in the absence or the presence of increasing concentrations of the peptide and following the accumulation of sedimented macroscopic assemblies by light microscopy (Fig. S7a). After incubation of NPM1 alone, we detected a small number of spherical aggregates, which may be related LLPS of NPM1 in the presence of trace amounts of nucleotides not detectable by MS. Addition of rpL5\(_{21-37}\) at a ratio of 1:10 caused a notable increase in assembly size and fusion into larger, amorphous structures. After 60 min incubation at a 10-fold excess of rpL5\(_{21-37}\), exclusively amorphous assemblies were observed. Assemblies formed in the presence of rpL5\(_{21-37}\) stained positively for the phase separation-specific DroProbe dye (Fig. S7b) (37). Since rpL5\(_{21-37}\) can induce the assembly of both FL NPM1 as well as the isolated NTD (13), we recorded the mass spectra...
In the presence of rpL521-37 (Fig. 4a). We observed significant peak broadening, indicating binding of the 2.2kDa peptide to NPM1 oligomers, as well as a shift from predominantly decamers to oligomers composed of ≥25 subunits. Next, we subjected complexes between FL NPM1 and rpL521-37 to MS analysis. Notably, we did not detect pentamers or their fragments upon collisional activation, but only trace amounts of monomers (Fig. 4b). Together, the insights from MS suggest that rpL521-37 binding increases the size and stability of NPM1 oligomers by cross-linking acidic tracts. Importantly, these oligomers have a considerably closer association between NTD pentamers than those formed by interactions with the CTD, due to the short length of the rpL521-37 peptide.

The fact that the rpL521-37 peptide modulates higher oligomerization of NPM1 enabled us to test the impact of self-assembly on chaperone activity. We, therefore, performed aggregation assays with Aβ42 and FL NPM1 in the presence of increasing amounts of rpL521-37 (Fig. 4c). Although rpL521-37 alone had no effect on Aβ42 fibrillation, it significantly reduced the ability of NPM1 to delay the formation of ThT-positive aggregates (Fig. 4c, Fig. S7d). Interestingly, the effect was observed already at a 5-fold excess of NPM1, corresponding to one rpL521-37 per pentamer. These data suggest that inducing a tighter and more complete association of NPM1 subunits through basic nuclear peptides reduces its chaperone activity. To investigate whether the observations for rpL521-37 represent a general mechanism, we also tested the SURF639-326 peptide, which encompasses basic tract 4 of SURF6 and induces LLPs of NPM1 (13, 38). AF models indicate a binding mode similar to that of rpL521-37 (Fig. S7f), in good agreement with their comparable effects on NPM1 assembly (13, 38). Aggregation assays showed that SURF639-326 alone has no impact on Aβ42 fibrillation but reduces the efficiency of FL NPM1 chaperoning to a similar extent as rpL521-37 (Fig. S7e, g). We conclude that proteins or peptides with multiple basic motifs cross-link NPM1 oligomers, which reduces their ability to delay fibril formation in vitro.

**Discussion**

The nucleolar scaffold protein NPM1 controls p53-dependent tumor suppression by sequestering aggregation-prone transcription factors, including p14ARF and c-MYC (7) and is implicated in the formation of amyloid bodies under stress conditions (18). In this study, we have investigated the basis of NPM1 self-assembly and how it relates to its ability to prevent amyloid formation in vitro. The CTD of NPM1 can delay fibrillation of the model amyloid Aβ42, yet this chaperone activity is modulated by NPM1 self-assembly. Using a combination of native MS, machine learning, and cryo-EM, we show that direct interactions between CTD and the acidic IDR in NPM1 induce the formation of an NPM1 network and thus reduce its ability to delay Aβ42 aggregation. Our findings imply that NPM1 pentamers are linked by their CTDs and IDRs, although additional interactions of the CTDs with the acidic NTDs cannot be excluded, as AF models of 29 homologs from the Pfam with identical domain architecture as NPM1 with sequence identities below 50% show clustering of the CTDs around the acidic side of the NTDs (Fig. S8). The length of the IDR (approximately 120 residues) leaves considerable space between the NPM1 pentamers, creating flexible compartments lined with acidic and some basic residues. Previous studies have suggested diffuse interactions between the acidic and basic tracts in oligomers (13), which could regulate the degree of compaction of the NPM1 network. The addition of basic peptides such as rpL5 or SURF6 cross-links the acidic NTDs and potentially the other acidic tracts, to constrict the network, in line with the induction of LLPs of NPM1 by polyC (9, 13) (Fig. 4d).

Our findings raise the question of how exactly NPM1 oligomers interact with unfolded or aggregation-prone clients. The observation that NPM1 does not seem to specifically inhibit primary or secondary nucleation of fibril elongation but rather affects all these events indicates that NPM1 works by partially sequestering Aβ42. We speculate that a fraction of the Aβ42 species is trapped in the NPM1 network and released on an equilibrium basis. Importantly, Aβ42 has not been identified as a client of NPM1 in vivo and does not include highly charged motifs that would promote recruitment into the NPM1 network via interactions with basic or acidic tracts. The fact that the chaperone activity of the CTD is higher than that of FL NPM1 suggests that Aβ42 can interact with free CTDs, and that this interaction is reduced when the CTDs instead bind to IDRs of neighboring NPM1 pentamers. The artificial amyloid β17, which shares some characteristics with Aβ42, colocalizes with NPM1 in cells (17), underscoring the possibility that NPM1 may bind a wider range of aggregation-prone sequences. Importantly, the isolated CTD does not prevent aggregation of de-natured globular proteins, which instead requires FL NPM1 (11, 15). These differences indicate different mechanisms for chaperoning unfolded proteins and amyloidogenic peptides, a distinction that has been reported for other chaperone systems (23, 39, 40).

Unfolded proteins expose hydrophobic as well as charged patches, which may drive their recruitment into NPM1 oligomers. The fact that the nucleolus does not actively promote refolding but holds unfolded proteins before transferring them to the HSP70 network supports this concept (17).

We hypothesize, based on our findings, that changes in nucleolar composition could balance the ability of the NPM1 scaffold to chaperone unfolded proteins or amyloids. Basic proteins, including rpL5 and p14ARF, mediate tight connections between the NPM1 pentamers but are released under stress conditions. The resulting loosening of the NPM1 network serves to activate the chaperone function of NPM1, in line with its role as safeguard against stress-induced protein aggregation (Fig. 4d). In this context, NPM1 may be viewed as an extreme case of a polydisperse chaperone (Fig. 4e).

Considering the limited physiological relevance of the NPM1-Aβ42 model system, we speculate that other proteins with a similar combination of charged regions, disordered regions, and oligomerization potential may exert a similar anti-amyloid activity. Such a protein has recently been identified as a noncanonical chaperone: the death-domain associated protein (DAXX) contains a folded N-terminal domain as well as a disordered region with poly-glutamate and poly-aspartate stretches and undergoes LLPs through interactions with p62 (41). DAXX can prevent Aβ42 fibrillation as well as the aggregation of heat-denatured proteins and associates with p53 to ensure correct folding (22). It furthermore controls nucleolar integrity (42), raising the possibility that poly-D/E proteins constitute a class of chaperones that incorporate into nucleolar condensates. Identifying the physiological targets of these chaperones and which properties of poly-D/E proteins they recognize will be an important step toward understanding their biological function.

Although our study clearly shows that direct interactions involving the CTD modulate the chaperone activity of NPM1, it has some limitations, as it does not address the possibility of conformational changes in the disordered regions. Specifically, the experimental approach used here (cryo-EM, AF, and native MS) is best suited for detecting complexes between folded domains. For example, the density map outside of the NTD is too diffuse to allow any unambiguous reconstructions, and we could not ob-
observe a shift to larger oligomers of NPM120 in the presence of
effect is related to phase separation of NPM1. For example, we
study. Our results indicate that RNA and basic peptides increase
contributions to NPM1 chaperone activity within the scope of this
broadly, the self-assembly of NPM1 is affected by multiple types of
“fuzzy” interactions, including with polyamions, polycations, and
molecular crowders (9), but we cannot independently assess their
contributions to NPM1 chaperone activity within the scope of this
study. Our results indicate that RNA and basic peptides increase
stability of NPM1 oligomers, but do not reveal whether this effect is related to phase separation of NPM1. For example, we
observe a shift to larger oligomers of NFM120 in the presence of
rpL512-37. However, the isolated NTD does not undergo phase sep-
oration with rpL512-37 unless the disordered A2 tract is included
(13), which hints at a need for flexible connections between the
protomers for LLPS to occur. Similarly, RNA molecules could in-
crease the distance between NTD pentamers and thus modulate
the fluidity of the assembly. Further studies are therefore needed
to assess the specific role(s) of LLPS in nucleolar chaperoning.

Materials and Methods
The density maps for the N-terminal region (“CryoEM structure of
the human Nucleophosmin 1 core”) and for the two selected 3D
classes have been deposited in the EMDB (EMD-15,606). The re-
fined atomic coordinates for the NPM1 core have been deposited in
the PDB with (code 8AS5). Full materials and methods are de-
scribed in the Supplementary Material file.

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Supplementary Material
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Authors’ Contributions
M.S. and M.L. designed the study with input from T.M.A., M.A.H.,
D.P.L., and J.J. M.S. and M.K. produced NPM1 variants, and G.C. pro-
duced Ab42. M.S. and A.L. performed aggregation assays. M.S. and
C.S. performed light microscopy analysis. D.L. performed energy
calculations. G.V.G. and M.H. recorded and analyzed E.M. data.
M.L. and A.E. generated structure predictions. M.L. wrote the pa-
per with input from all authors.

Data Availability
The density maps for the N-terminal region (“CryoEM structure of
the human Nucleophosmin 1 core”) and for the two selected 3D classes have been deposited in the EMDB (EMD-15,606). The re-
fined atomic coordinates for the NPM1 core have been deposited in
the PDB with (code 8AS5).

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