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Small-angle X-ray scattering (SAXS) was used to monitor structural changes induced by heat treatment and acid gelation in milk matrices with added whey protein concentrates (WPCs) and nano-particulated whey protein (NWP). In general, heat treatment was found to mainly affect whey protein components while pure casein micelles remained largely unaffected. Conversely, acidification mainly affected caseins while leaving pure whey protein components intact. In mixed systems, the overall behaviour could be understood as a combination of the above effects, however, the type of the added whey protein components influenced the resulting structure formation and dynamics. NWP led to formation of larger structures compared to WPC components during heat treatment, although the latter showed faster aggregation dynamics. During acidification the NWP containing samples exhibited structural changes at slightly higher pH values than the WPC samples. The modeling of pure liquid whey protein (LWPC) samples showed that the heat induced denaturation and resulting aggregation of individual whey proteins is mainly a surface effect leaving the overall protein shape and dimensions unaffected. Schematic diagrams based on the current SAXS data and previous studies were constructed to illustrate the suggested interaction mechanisms between casein and whey proteins during both heating and acidification.

1. Introduction

The addition of whey protein ingredients to dairy products has become a common practice as they can provide favorable nutritional and functional properties, such as increased creaminess and viscosity (Janhøj, Petersen, Frost, & Ilsen, 2006; Torres, Janhøj, Mikkelsen, & Ilsen, 2011), or softer texture (Tamime, Kalab, Muir, & Barrantes, 1995) especially in low-fat yoghurt. Heat treatment/pasteurization and acidification are integral parts of industrial processing of yogurt. In general, heat treatment will cause denaturation of whey proteins leading to partial aggregation as well as interactions with the casein micelles, and the process of acidification can lead to considerable modifications in the milk components and especially change the structure of casein micelles. Therefore, the investigation of structural changes induced by heat treatment and acidification is integral parts of industrial processing of yogurt. In general, heat treatment was found to mainly affect whey protein components while pure casein micelles remained largely unaffected. Conversely, acidification mainly affected caseins while leaving pure whey protein components intact. In mixed systems, the overall behaviour could be understood as a combination of the above effects, however, the type of the added whey protein components influenced the resulting structure formation and dynamics. NWP led to formation of larger structures compared to WPC components during heat treatment, although the latter showed faster aggregation dynamics. During acidification the NWP containing samples exhibited structural changes at slightly higher pH values than the WPC samples. The modeling of pure liquid whey protein (LWPC) samples showed that the heat induced denaturation and resulting aggregation of individual whey proteins is mainly a surface effect leaving the overall protein shape and dimensions unaffected. Schematic diagrams based on the current SAXS data and previous studies were constructed to illustrate the suggested interaction mechanisms between casein and whey proteins during both heating and acidification.

A R T I C L E I N F O

Keywords:
Small-angle X-ray scattering
Casein micelle
Nano-particulated whey protein
Whey protein concentrate
Acidified gel
Interactions of milk proteins

A B S T R A C T

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1. Introduction

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The production proceeded under a controlled uniform temperature of 4°C in a 30 L pilot scale reactor equipped with 0.2 μm ceramic membrane (Lenntech, Delfgauw, the Netherlands) and a 0.1 μm ceramic membrane (Membrane Systems, Urdorf, Switzerland) equipped with 0.1 μm polyethylene membrane. MF separation was performed using a pilot-scale SW25 MMS system (MMS AG Membrane Systems, Switzerland), and UF separation was performed using a pilot-scale UF Lab-stack (Tetra Pak, Silkeborg, Denmark) fitted with GR61PP membranes (Alfa Laval, Lund, Sweden) of 20,000 Da nominal molecular mass cut-off. The obtained MF permeate was collected and used for dilution during sample preparation. Subsequently, LCC and LWPC samples were kept at –80°C, and permeate at 4°C until use. More details about the production can be found in our previous papers (Li et al., 2021a; 2021b).

For thawing of samples, the same procedure was adopted as in Li et al. (2021b), i.e., the frozen samples were initially kept in a refrigerator at 5°C for 3d, and subsequently moved to a water bath and kept at 30°C for 30 min.

2.2. Acidified milk model systems processing in lab scale

A total of 8 milk model systems were constructed using casein (LCC) and 3 different types of whey proteins (LWPC, WPC, NWP) in different ratios, including one reference (SM) mimicking the composition of commercial skim milk and which consisted of 2.8% LCC and 0.9% LWPC. Permeate from UF was used to dilute the samples, thus mimicking the milk environment in terms of minerals and lactose. Commercial skim milk (Com SM) was included as an additional reference. Except for the two references, all milk model systems had a total protein content of 4% (w/w) with casein and whey protein in the ratio of 1:1 (see Table 1 for more details).

Each of the mixed and diluted milk model systems of 300 mL were first pre-stirred at 800 rpm for 30 min with a magnetic stirrer (IKA™ RET Basic Magnetic Stirrer, Fisher Scientific, Roskilde, Denmark) at room temperature. Afterwards, they were continually stirred at a speed of 600 rpm overnight in a cold room at 5°C, using the same magnetic stirrer. Then, a pre-heat treatment (55°C, 5 min) was performed in a water bath before samples were subjected to a two-stage homogenization (20/5 MPa) using a high-pressure homogenizer (Panda plus 1000, GEA Niro, Soavi, Italy).

Subsequently, each of the samples were divided in aliquots of 20 mL in 50 mL centrifuge tubes and subjected to a heat treatment at 90°C for 10 min in a water bath. The selected condition of heat treatment followed our previous study and emulates what is applied in industrial yogurt production (85–90°C for 5–10 min) (Walstra et al., 2005; Li et al., 2021b). During this step, samples were gently stirred for 30 s and then kept in a water bath at 45°C until the pH reached 4.60 ± 0.05. This was monitored using a HQ411D Laboratory pH mV Meter with an Intellical™ PHC705 electrode (Hach, Düsseldorf, Germany) and pH was measured in triplicate. The relation between

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Experimental design and ingredients used in the non-fat acidified milk model systems.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total protein content (% w/w)</td>
<td>No.</td>
</tr>
<tr>
<td>(%)</td>
<td>(%)</td>
</tr>
<tr>
<td>4</td>
<td>1</td>
</tr>
<tr>
<td>2</td>
<td>LWPC</td>
</tr>
<tr>
<td>3</td>
<td>LCLW</td>
</tr>
<tr>
<td>4</td>
<td>W1</td>
</tr>
<tr>
<td>5</td>
<td>W2</td>
</tr>
<tr>
<td>6</td>
<td>N1</td>
</tr>
<tr>
<td>7</td>
<td>N2</td>
</tr>
<tr>
<td>3.5</td>
<td>8</td>
</tr>
<tr>
<td>9</td>
<td>Com SM</td>
</tr>
</tbody>
</table>

Note: LCC represent liquid casein concentrate, LWPC represents liquid whey protein concentrate, NWP is nano-particulated whey protein, and WPC is whey protein concentrate. The calculated protein composition ignores the small amount of whey protein and casein content existing in LCC and LWPC, respectively.
pH and time was found to be logarithmic (seen in equation (1) during acid gelation with $R^2$ values ranging from 0.995 to 0.999.

$$pH = A \ln(t) + B$$

where $t$ represents time in min. The values of $A$ and $B$ varied slightly for the different model systems (data not shown).

2.3. Small-angle X-ray scattering (SAXS)

The measurements during heat treatment were performed using a GANESHA instrument from SAXS-LAB (Lyngby, Denmark) with a Rigaku (Rigaku-Denki, Co., Tokyo, Japan) 40 W micro-focused Cu source producing X-rays. A wavelength of $\lambda = 1.54$ Å was detected by a moveable Pilatus 300 k pixel-detector from Dectris (Baden, Switzerland), which is capable of measuring different angular ranges and thus length scales. The two-dimensional scattering data were azimuthally averaged using standard reduction software (SAXSgui, Lyngby, Denmark). The scattering patterns for the radially averaged intensity (Int.) from the detector were recorded as a curve against the scattering vector $q$.

The log-normal distribution follows.

$$\ln(q) \sim \ln(q_p) - \frac{\ln(\Delta q)}{\sqrt{2\sigma}}$$

where

$$q_p = q^{\text{ref}}(\frac{qR_0}{\sqrt{6}})^{-1}$$

$R_g$ is the radius of gyration, $G$ is the Guinier prefactor, $B$ is a prefactor specific to the type of power-law scattering, and $P = -4$ in Porod’s law used in this study. For each level, the four parameters $G$, $R_g$, $B$, and $P$ were chosen.

The selected scattering data from heat treatment were fitted to a polydisperse sphere model in the $q$ range from 0.008 to 0.05 Å$^{-1}$. The following equation was applied for calculation (Yang et al., 2021).

$$F(q, \alpha) = \Delta \rho V_p \left[ \frac{3\sin(qR)' - q\cos(qR)R}{(qR)^2} \right]$$

where

$$r = \left[ R_p^2 \sin^2 \alpha + R_p^2 \cos^2 \alpha \right]^{1/2}$$

$\alpha$ is the angle between the axis of the ellipsoid and $q$, and $V = (4/3)\pi R_p^2 R_e^2$ is the volume of the ellipsoid. Here $R_p$ is the polar radius, and $R_e$ is the equatorial radius. $\Delta \rho$ (contrast) is the scattering length density difference between the scatterer and the solvent.

The unified_power_Rg model adopts the empirical multiple level unified Exponential/Power-law fit method, which is developed by Beaucage (1996). Here the intensity is given by.

$$I(q) = \text{background} + \sum_{i=1}^{N} [G_i \exp \left( -\frac{q^2 R_i^2}{3} \right) + B_i \exp(-\frac{q^2 R_{i\text{inel}}^2}{3})(\frac{1}{q_i})^P ]$$

2.3.1. Heat treatment

Fig. 1 presents an overview of background subtracted SAXS data from the different milk model systems listed in Table 1. These were all subjected to heat treatment (90 °C) for 0, 2, 5, and 10 min. From Table 1 we see that the casein-only system (LCC) and the pure whey protein sample (LWPC) can be considered the building blocks of the remaining samples so we will describe their behavior first and then turn to the mixed samples.

3. Results and discussion

3.1. Heat treatment

The casein-only system (LCC) only exhibited slight changes in the
scattering data, which is not surprising since the overall internal structure of casein micelles is not expected to be markedly disrupted at temperatures below 120 °C at the native pH of milk (pH 6.7) (Dalgleish & Corredig, 2012). However, the system containing only whey protein (LWPC), showed significant changes in the scattering data with increasing heat treatment time, particularly at low q. We attribute this to some level of denaturation or surface disruption and subsequent aggregation of whey protein at the applied temperature of 90 °C. A model combining a local ellipsoidal shape to describe the individual protein contour and the unified model to describe larger aggregates were applied to interpret the scattering of LWPC (Fig. 2). The ellipsoid can be taken to represent the local shape of whey proteins in LWPC with dimensions matching those of the known crystal structure of β-lactoglobulin (PDB ID 1BEB), which is the major whey protein. The scattering data show that this local structure is basically preserved during the heat treatment as evident from the unchanged scattering at q-values larger than ca. 0.07 Å⁻¹ during the entire heat treatment duration of 10 min (note the unchanged radius of the ellipsoid shape in Table 2). A partial denaturation where the overall shape is largely preserved, but where protein parts exposed on the surface are mainly affected and could induce aggregation, would explain the obtained scattering data.

The aggregation is evident from the behavior at q-values lower than ca. 0.07 Å⁻¹ where significant changes occur over the course of the heat treatment. At 0 min there is a slight upturn in the intensity at the lowest q-values, indicating the presence of a small amount of larger aggregates (Fig. 2), but overall the scattering here is fully described by the local
ellipsoidal shape. As the heat treatment progresses, the unified model fits indicate the presence of two size populations evolving as described by the fitted radii of gyration (Rg). These numbers go from 5.5 nm / 14 nm (2 min) to 7 nm / 45 nm (5 min) and finally 10 nm / 60 nm after 10 min (Table 2). The increasing sizes of aggregates observed here show that most of the native whey protein is affected by the heat treatment in a way that favors aggregation into larger structures, but whether the proteins are fully denatured is not clarified - if so, the overall nanoscale shape remains relatively unchanged as mentioned above. Corresponding proteins are fully denatured is not clarified - if so, the overall nanoscale nanoscale aggregate size is coarser. The fitted radii of gyration (Rg) show a dominant distribution between 20 nm and 80 nm, with a more dominant population at 20 nm in radius in the intermediate-q, apart from the pure LCC, which basically remained unchanged as mentioned above.

Two questions seem relevant: do the different whey protein components behave similarly, and to what extent do they interact with caseins? Fig. 2 B reveals the following progression of particle size formation between the mixed samples: SM/Com SM < W1/2 < LCLW < N1/2.

Apart from the self-interactions demonstrated above in the LWPC sample, whey proteins can aggregate together with micellar or serum α-casein and κ-casein, with sizes of formed complexes ranging from 30 to 100 nm (diameter) during heat treatment (Donato & Dalgleish, 2006; Donato, Guyomarc'h, Amiot, & Dalgleish, 2007), while deviation may occur due to the different measurements of particle sizes applied compared to the present study. It has also been reported that only around 20% of the denatured whey proteins attach to casein micelles and the rest exist as soluble complexes in the solution at the natural pH of milk (Kethireddipalli, Hill, & Dalgleish, 2010). Thus, it may be speculated that most of the particle sizes (radius around 3-20 nm) found in the present study are likely to represent soluble denatured whey protein-whhey protein aggregates although the association of whey protein and individual caseins may also occur. In particular, compared to the pure LWPC sample, the mixed system LCLW (2% LCC and 2% LWPC) presented a larger average radius of ~8 nm (versus ~6 nm). It can thus also be proposed that the formed whey protein aggregates (LWPC) were affected by the presence of casein (LCC), e.g., forming complexes with individual caseins during heating. Both W1 and W2 presented a similar particle size distribution as seen for LCLW, but with lower average radius than LCLW. This may suggest a similar aggregation behavior of WPC as for LWPC, but with more native whey protein in LWPC leading to more exposed bonds available for interaction with other proteins during heating. Furthermore, it was reported by the manufacturer that the WPC powder also contained ~17% caseinomacropeptide (CMP) as it was produced from whey stemming from cheese production. CMP will not denature/aggregate during heat treatment. Both N1 (0.5% LWPC

Table 2

<table>
<thead>
<tr>
<th>Time/ min</th>
<th>Ellipsoid</th>
<th>Unified power</th>
</tr>
</thead>
<tbody>
<tr>
<td>A-scale</td>
<td>A-radius-equatorial</td>
<td>B-level 1</td>
</tr>
<tr>
<td>(nm)</td>
<td>(nm)</td>
<td>B-B1</td>
</tr>
<tr>
<td>A-level 1</td>
<td>A-level 2</td>
<td>B-G1</td>
</tr>
<tr>
<td>(nm)</td>
<td>(nm)</td>
<td>B-G2</td>
</tr>
<tr>
<td>B-level 2</td>
<td>B-level 3</td>
<td>B-B2</td>
</tr>
<tr>
<td>(cm^-3)</td>
<td>(cm^-3)</td>
<td>(cm^-3)</td>
</tr>
<tr>
<td>0</td>
<td>0.0064</td>
<td>5</td>
</tr>
<tr>
<td>2</td>
<td>0.0064</td>
<td>5</td>
</tr>
<tr>
<td>5</td>
<td>0.0070</td>
<td>5</td>
</tr>
<tr>
<td>10</td>
<td>0.0070</td>
<td>5</td>
</tr>
</tbody>
</table>

Note: Scale: means scale factor or volume fraction; Rg is radius of gyration.

Fig. 3. Schematic of LWPC structural development with increasing heat treatment (90 °C) time from 0 to 10 min. LWPC represents liquid whey protein concentrate produced from ultra-filtration.
and 1.5% NWP) and N2 (1% LWPC and 1% NWP) demonstrated relatively narrow particle size distributions and increased particle sizes after heating compared to LCLW, W1 and W2. N1 exhibited the highest average radius with narrower distribution, which may imply that heating induced larger NWP-NWP, or NWP-LWPC, aggregates formed in N1. N2 presented lower average radius of ~8 nm compared to ~9.5 nm for N1 during heating. NWP has previously been shown to exhibit larger aggregate sizes compared to systems with added either micro-particulated whey protein (MWP) or WPC. It has been suggested that NWP has a large surface area because of its original small size (Li et al., 2021b).

A simplified quantitative view of the effect of the heating time can be obtained by looking at the intensity development at a single selected q-value of ~0.025 Å⁻¹ as shown in Fig. 5. The intensity values were normalized to the initial intensity value at time 0. The most significant effect after 10 min is seen for the LCLW sample, but interestingly only the W1/2 samples exhibit any sign of aggregation after 2 and 5 min of heating. As mentioned above, these samples show similarities in the size distributions after 10 min of heating, but the time evolution suggests that the W1/2 samples are slightly more susceptible to heat induced surface modifications. Apart from the W1/2 samples, all samples have to be heated for more than 5 min to show any detectable aggregation behavior.

3.1.3. Skim milk references SM and Com SM

The skim milk samples (SM and Com SM) did not show significant differences during heating. Two reasons for this seems likely: first, these samples contain only 0–0.7% whey proteins compared to 2% in the other samples, reducing the probability for whey proteins to associate in solution. Second, as seen in equation (2) above, the scattering intensity scales both with the square of the particle volume and the particle concentration, thus the casein micelles dominate the spectrum until whey protein aggregates reaches a certain level depending on size and number of particles.

3.2. Acidification

An overview of the SAXS scattering data obtained during acidification for each of the model systems can be seen in Fig. 6. Further, in Fig. 7 A and B, the intensity evolution as a function of pH is shown for two selected q-values. A quick glance of Fig. 7 B shows that the samples again fall into groups with similar behavior, namely the high casein containing samples LCC/SM/Com SM, the W1/2 and LCLW samples, the N1/2 samples and finally the pure whey sample LWPC.

3.2.1. LWPC sample

The most striking changes are the gradual smoothing of high-q features and an overall intensity reduction with decreasing pH for all the
model systems, except for the whey protein-only sample, LWPC, which almost maintains its initial local structure during the entire acidification process. Thus, opposite to what was observed for the heat treatment, the whey proteins are largely unaffected by the acidification as quantified by basically constant intensity values in Fig. 7 A and B.

3.2.2. LCC and SM/Com SM samples

For the LCC sample and the other two high casein containing samples SM/Com SM the situation is opposite as the scattering curves change both in intensity and shape. Changes in high-q (0.08–0.1 Å\(^{-1}\)) features represent primarily local protein inhomogeneities (De Kruif, 2014; Ingham et al., 2016) and were most pronounced in the casein-only sample LCC (4% casein), and the two references SM and Com SM (2.8% casein) compared to the other mixed systems (2% casein). Fig. 7 shows that the intensity development is different at high and low q. At high q the intensity drops roughly linearly with pH while at low q the intensity drops steeply around a pH of ca. 5.2. Again, with reference to equation (2), there can be several reasons for changes in the intensity: the size and number of objects, but also the contrast (\(\Delta \rho\)) given by the difference in electron density of the proteins and the surrounding medium. In this case, contrast changes can be caused by the release of calcium from the casein micelles into the serum phase as well as a general disruption of the micelles, creating a more uniform protein solution. However, there are also clear structural changes occurring: at the lowest q-values the shape of the curve initially contains a distinct signature of the overall casein micelle size from the curvature of the

Fig. 5. Changes of scattering intensity after heating 2, 5 and 10 min compared to the original sample (0 min) at q = 0.025 Å\(^{-1}\). Values were calculated by dividing the intensity of original sample (0 min heat treatment). Each error bar is the mean ± SD (n = 2); values with uppercase letters represent significant difference between heating time, and lowercase letters mean significant difference between different model systems at the same heating time (P < 0.05).

Fig. 6. Background-subtracted SAXS data of different milk model systems at different pH, as indicated in the legend, during acidification. The descriptions of the systems are given in Table 1.
data. This curvature is gradually lost indicating most likely a micellar breakdown or at least a loosening, potentially leading to larger structures outside the resolution of our instrument. Also, increasing aggregate size could also lower the intensity as large aggregates would sediment out of the x-ray beam (Ingham et al., 2016).

In fact, pH values of 5.2–5.1 corresponds to a region where the structure of casein micelle loosens, CCP is completely solubilized into the serum phase and phosphoproteins are redistributed. Ingham et al. (2016) also mentioned a similar phenomenon of broader high-q features appearing in skim milk at pH 5.15 during acid gelation using GDL. During milk acidification, a specific demineralization, i.e., a progressive release of CCP from the interior of the casein micelles and dissolving into the serum phase at pH dropping from 6.7 to 5.3 has been observed by Marchin et al. (2007). Casein micelles have been reported to become smaller, more homogeneous and have a higher average density from pH 6.5 to 5.2 during skim milk acidification (Moitzi, Menzel, Schurtenberger, & Stradner, 2011) with a subsequent disappearance of the high-q feature at pH 4.85. The explanation put forward was, that already released casein in the serum aggregated at the low pH, providing a more uniform structure in this high-q region (Ingham et al., 2016; Moitzi et al., 2011). These observations generally make sense also for our data.

3.2.3. Mixed systems, LCLW, W₁ and N₁

In comparison to LCC and the references (SM and Com SM), the mixed systems containing 2% of casein and 2% of whey protein (i.e., LCLW, W₁, W₂, N₁ and N₂) showed less evident high-q feature decreasing rate (Fig. 7A). The low-q behavior however, also shows a decrease similar to the LCC sample, but with reduced rate and for the N₁/₂ samples at a slightly higher pH, ca. 5.4.

In addition, the overall intensity reduction rates of mixed systems (i.e., LCLW, W₁, W₂, N₁ and N₂) seems less significant compared to LCC and the references (SM and Com SM). This may be related to both the different varying rate of reduced contrast and casein micelle structure disruption, and relatively smaller casein-whey protein aggregates forming during acidification compared to larger casein-casein aggregates in LCC and the skim milk references.

From the overview of SANS scattering in the low-q range (0.004–0.005 Å⁻¹), all the model systems, except LWPC, showed a small deviation at the natural pH (6.7), which was related to a comparatively rough surface of casein micelles based on the study by Liu et al. (2017a). They found that the small positive deviation followed Porod’s law at natural pH of skim milk. The interfacial scattering intensity after this deviation, i.e. the low-q range decayed by a q⁻⁴ power law during acidification with GDL, where a relatively smooth and sharp interface was gradually formed as pH was decreasing. Besides, they confirmed these structural characteristics of casein micelles before and after acidification using transmission electron microscopy (TEM) and explained the changes as a result of concomitant gradual shrinkage of κ-casein hairy layers. Thus, the interface of the casein micellar structure in the studied model systems may also become more smooth and sharp with decreasing pH in the present study. The scattering intensities of LCC, and the two references (SM and Com SM) were lower than for the other mixed systems (LCLW, W₁, W₂, N₁ and N₂) below pH 5.0 at a selected low-q around 0.005 Å⁻¹, seen in Fig. 7B. This might imply that larger casein-casein aggregates formed after demineralization. In addition, compared to LWPC, the intensities of LCLW were much higher even though they kept decreasing with decreasing pH. It cannot be directly confirmed that LWPC interacted with LCC, forming casein-whey protein aggregates larger than the whey protein-whey protein aggregates present, but smaller than casein-casein aggregates (present in LCC). The general decreasing intensities and rate of LCLW were similar to W₁ and W₂, except at lower pH values from 4.8 to 4.6, where its intensity was slightly lower. This could indicate slightly larger casein-WPC aggregates forming in this range. The intensities of systems with added NWP (N₁ and N₂) decreased more significantly than LCLW, W₁ and W₂ from pH 6.7 to 4.6 (Fig. 7B). NWP has been reported to self-associate at pH ≥ 5.5 caused by decreased electrostatic repulsion and enhanced hydrophobic interaction (Liu et al., 2017b).

The mixed systems (LCLW, W₁, W₂, N₁ and N₂) showed a second (less distinct) inflection point at around 0.009 Å⁻¹, which was not seen for LCC and the two references (SM and Com SM) (Fig. 6). The inflection points disappeared at around pH 5.2, when dense proteins disrupt (casein micelles loosen). This may indicate that more added whey protein may induce changes to the structure of casein clusters by interacting with caseins as seen from the intermediate-q range (0.009–0.04 Å⁻¹) from pH 6.7 to 5.2.

3.3. General discussion

Schematic diagrams of interactions between the proteins in the selected three different model systems (i.e., LCLW, W₁, N₁) during both heating and acidification are shown in Fig. 8. The diagrams are based on both the discussion of obtained SANS scattering data in the present study and on previous studies (Ingham et al., 2016; Li et al., 2021b; Liu et al., 2016), suggesting a possible interaction mechanism of the selected proteins from changes of their local protein structure. The casein micelle model chosen is the one suggested by Ingham et al. (2016) considering a hydrated sponge-like structure. K-casein on the surface ensures that the micelle is stable against aggregation, and the chains of casein-omacropete (CMP) are extended from the micellar surface and anchored as a hairy layer, providing steric stabilization of the micelles.
After heat treatment, some of the whey proteins are able to form disulphide bonds with κ-casein in the inner part of the casein micelle by penetrating the layer of CMP (Donato et al., 2007), and others aggregate themselves or with individual caseins in the serum, which are illustrated in Fig. 8. Meanwhile, the complexes of whey protein-κ-casein in the inner part of the casein micelle are reported to be also capable of preventing coagulation of casein micelles, resulting in their stability in milk solution (Dalgleish & Corredig, 2012). In addition, the complexes can facilitate additional attachment points between micelles during acid gelation (Ingham et al., 2016). Based on the discussion of SAXS changes for the different systems above, heat treatment did not markedly affect the overall structure of casein micelles (e.g. LCC). However, it caused significant changes to all mixed systems (LCLW, W$^1$, W$^2$, N$^1$, and N$^2$) with whey protein included, mainly in the intermediate-q range of 0.009–0.04 Å$^{-1}$ with particles sizes ranging from 1 to 20 nm (radius). Most soluble denatured whey protein aggregates can be assumed to form in the serum phase, which were observed in this intermediate-q range. More LWPC-LWPC aggregates were formed after heat treatment of 10 min, as the structure of native whey proteins changes to a greater degree when getting denatured and aggregating. The behavior of WPC was similar to LWPC, because WPC also contains a small amount of native whey protein, leading to formation of more disulphide bonds after heating than NWP. NWP mostly includes denatured whey protein aggregates produced in nano size and the small particles may already aggregate in the solution before heating because of higher surface area. Subsequently, the NWP aggregates continue to attach to other proteins, forming larger aggregates during heating. The higher average radius with narrower distribution in the system with added NWP may imply that larger NWP-NWP, NWP-LWPC, or even LWPC-LWPC aggregates were formed during heating. It has also previously been reported that the average particle size of a model system with added WPC was only slightly higher than that with added LWPC, but significantly lower than that with NWP, when they were mixed with the same amount of casein (Li et al., 2021b). NWP was also suggested to interact with other proteins, like casein and whey protein isolates (Liu et al., 2016).

During acid gelation, the steric stabilization of micelles will decrease with the decreasing pH, where the κ-casein layer collapses due to the decreasing charge of CMP (De Kruijff, 1999). This can be inferred from the small deviation at low-q features (0.004–0.005 Å$^{-1}$), and high-q features disappearing at pH ~ 5 for almost all model systems with added LCC in this study. The micelles can then connect at a closer range because of the attractive forces (Dalgleish & Corredig, 2012). As indicated in Fig. 8, the overall structure of micelle aggregates in the three model systems became more compact and denser during acidification. The decreasing overall intensity of SAXS scattering may also imply an increase in larger aggregates formed during acidification, which are sedimented out of solution and not detected by the X-ray beam for the model systems of LCLW, W$^1$, and N$^1$. The intensity of SAXS scattering decreased more

Fig. 8. Diagrams of interacting caseins with different types of whey proteins during processing of acidified gels (only the interacting parts are shown, not the whole particles). αs- and β-caseins are represented by yellow hairs. κ-casein is blue, the caseinomacropeptide chains are black, and calcium phosphate nanoclusters are represented by grey spheres. Inherent whey protein (LWPC) are in red hairs, NWP are purple spheres, and WPC are red spheres. Not drawn to scale.
significantly for N\textsuperscript{1} compared to LCLW and W\textsuperscript{1} at low q -range, meaning that NWP may form larger aggregates during acidification, as indicated in Fig. 8.

4. Conclusion

Structural changes of milk model systems with added whey protein ingredients induced by heat treatment and acid gelation were investigated using SAXS. Overall, heat treatment was found to mainly affect whey protein components, and acidification mainly affected caseins. In mixed systems of both casein and whey proteins, the structure formation and dynamics can be influenced by the form of the added whey protein components. The formation of a larger structure was obtained for systems containing NWP compared to WPC during heat treatment. The NWP containing samples also showed structural changes at slightly higher pH values than the WPC samples during acidification. The pure liquid whey protein (LWPC) samples was affected significantly through denaturation and resulting aggregation by heat treatment, the modeling of its SAXS data presented mainly a surface effect leaving the overall protein shape and dimensions unaffected. A mechanism for the protein interactions was suggested based on their different internal structural changes during both heat treatment and acidification, and combined with previous findings. SAXS was shown to be a promising non-destructive method to probe turbid samples, like the acidified gels in the present study, evaluating their original structures without disruption, which would not be possible with conventional light scattering techniques.

CRediT authorship contribution statement

Ruifen Li: Conceptualization, Methodology, Formal analysis, Investigation, Validation, Writing – original draft. Tanja Christine Jæger: Conceptualization, Writing – review & editing, Funding acquisition. Tijs A.M. Rovers: Conceptualization, Resources, Writing – review & editing, Funding acquisition. Birte Svensson: Conceptualization, Writing – review & editing, Funding acquisition. Richard Ilsen: Conceptualization, Methodology, Validation, Writing – review & editing, Supervision, Project administration, Funding acquisition. Jacob J.K. Kirkengaard: Conceptualization, Methodology, Validation, Writing – review & editing. Anny Bygræ Hougaard: Conceptualization, Resources, Validation, Writing – review & editing, Supervision, Project administration, Funding acquisition.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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