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Genomes shed light on the evolution of *Begonia*, a mega-diverse genus

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Summary

- Clarifying the evolutionary processes underlying species diversification and adaptation is a key focus of evolutionary biology. *Begonia* (Begoniaceae) is one of the most species-rich angiosperm genera with c. 2000 species, most of which are shade-adapted.
- Here, we present chromosome-scale genome assemblies for four species of *Begonia* (*B. loranthoides*, *B. masoniana*, *B. darthvaderiana* and *B. peltatifolia*), and whole genome shotgun data for an additional 74 *Begonia* representatives to investigate lineage evolution and shade adaptation of the genus.
- The four genome assemblies range in size from 331.75 Mb (*B. peltatifolia*) to 799.83 Mb (*B. masoniana*), and harbor 22 059–23 444 protein-coding genes. Synteny analysis revealed a lineage-specific whole-genome duplication (WGD) that occurred just before the diversification of *Begonia*. Functional enrichment of gene families retained after WGD highlights the significance of modified carbohydrate metabolism and photosynthesis possibly linked to shade adaptation in the genus, which is further supported by expansions of gene families involved in light perception and harvesting. Phylogenomic reconstructions and genomics studies indicate that genomic introgression has also played a role in the evolution of *Begonia*.
- Overall, this study provides valuable genomic resources for *Begonia* and suggests potential drivers underlying the diversity and adaptive evolution of this mega-diverse clade.

Key words: *Begonia*, evolution, genomes, introgression, shade adaptation, whole-genome duplication.

*These authors contributed equally to this work.
Introduction

The mechanisms underlying the diversification of large clades of closely related species (often designated taxonomically as genera) remain one of the biggest mysteries in plant biology (Frodin, 2004). Although speciose genera have received widespread attention from evolutionary biologists, typically few genomic resources are available for these closely related, species-rich clades. Representative completely assembled nuclear genomes of only three of the 10 largest angiosperm genera (Frodin, 2004) have been published, namely Solanum (A. Bolger et al., 2014; Song et al., 2019), Dendrobium (Yan et al., 2015) and Begonia (Griesmann et al., 2018). However, these genomic studies either focused on the specific characteristics of the reference species (A. Bolger et al., 2014; Yan et al., 2015; Song et al., 2019) or were part of a large comparative genomic project (Griesmann et al., 2018); none of them used genomic data to explore the evolutionary patterns in these mega-diverse clades.

Begonia L. (Begoniaceae, Cucurbitales) is well known for a huge diversity of leaf shapes, patterns and textures (Fig. 1). The genus is pantropical and comprises more than 2000 currently recognized species, as well as many hybrids (Hughes & Hollingsworth, 2008). We compared the four Begonia species presenting 37 of the 70 recognized sections of Begonia L. (Begoniaceae, Cucurbitales) across all three major continental clades (Supporting Information Fig. S1). We also examined cytonuclear incongruences detected in our study, and investigated the molecular basis of shade adaptation in Begonia.

The interpretation of Begonia genomic diversity in an evolutionary context will not only contribute to a better understanding of the origin, evolution and shade adaptation of this megadiverse clade, but also provide valuable reference genomes for molecular breeding of these highly valued ornamental plants.

Materials and Methods

Sample collections and DNA/RNA extraction

All Begonia samples were collected from the glasshouse in Fairy Lake Botanical Garden (Shenzhen, China) where plants were cultivated at 26°C: 18°C (day : night) with a relative humidity of 65–80%. Specimens have been deposited in the Herbarium of Fairy Lake Botanical Garden. For WGS, genomic DNA from young leaves of each individual was extracted using the cetyl trimethylammonium bromide (CTAB) method (Porebski et al., 1997). For single tube long fragment read (stLFR) sequencing, high-molecular-weight genomic DNA was isolated using the IrysPrep® Plant Tissue DNA Isolation kit (RE-014-05; Bionano Genomics, San Diego, CA, USA) following the manufacturer’s instructions. DNA quality and quantity were evaluated using pulsed field gel electrophoresis and Qubit® 3.0 Fluorometer (Thermo Fisher Scientific, Waltham, MA, USA). For transcriptome sequencing, total RNA from different tissues (root, stem/rhizome, leaf, peduncle and flower) from four Begonias was isolated using TRIzol® reagent (Invitrogen, Carlsbad, CA, USA), respectively. All the RNA samples were quality controlled using a NanoDrop™ One UV-Vis spectrophotometer (Thermo Fisher Scientific) and a Qubit® 3.0 Fluorometer.

Library preparation and sequencing

All DNA libraries for WGS were constructed using the MGIEasy FS DNA Library Prep Set (1000006988) with 300–500 bp fragment sizes, and sequenced on an Illumina HiSeq2000 platform to generate paired-end (PE) reads of 150 bp. Transcriptome libraries were constructed with a MGIEasy RNA Library Prep Kit (1000006384) with inserts of 200–400 bp and sequenced with PE reads of 100 bp. More than 5 Gb of sequence data were generated for each library. The stLFR library was prepared with the MGIEasy stLFR Library Prep Kit (1000005622) (Wang et al., 2019) and sequenced with PE reads of 100+42 bp, generating >150 Gb of raw sequence data for each library. 10 × Genomics Chromium® Genome libraries with insert sizes of 350–500 bp were prepared with Chromium Genome Reagent Kit (v2 Chemistry, Pleasanton, CA, USA) following the manufacturer’s protocols with modified PCR primers to introduce sequencing primers suitable for the BGISEQ-500 platform and then sequenced with PE reads of 150 bp. SMART library preparation and sequencing details are given in Methods S1.
Fig. 1 Phylogenetic tree showing the topology and divergence times for 78 newly sequenced species of Begonia. Maximum-likelihood tree inferred with RAxML based on SNPs within regions of nuclear single-copy genes. Divergence times are indicated at the internodes; the range of the blue bars indicates the 95% confidence interval of the divergence times. Representative images on the right show Begonia diversity. Three major geographically circumscribed clades are colored blue, red and green for the African, Neotropical and Asian clade, respectively. Taxa in bold highlight the four species with chromosome-scale genomes generated in this study. Branches are maximally supported unless otherwise indicated.
Genome assembly

For assembly of the sequences from 10 × Genomics Chromium and stLFR libraries, the clean reads were obtained using an in-house script and de novo assembled using SPERNova (v.2.1.1) (Weisenfeld et al., 2017) with default parameters. A minimum fasta record size of 100 bp was specified at the ‘mikoutput’ stage for outputting the assembly in the ‘pseudohap’ style. De novo assemblies of the PacBio long reads for B. masoniana and B. darthvaderiana were conducted by Canu (v.0.1) (Koren et al., 2017). Subsequently, two rounds of iterative corrections were performed with PacBio long reads using the software RACON (v.1.2.1) (Vaser et al., 2017), and two rounds of corrections with Pilon (v.1.22) (Walker et al., 2014) using 10 × Genomics reads (see details in Methods S1).

Variant analysis

A total of 468 Gb 150 bp PE Illumina reads were generated, yielding an average coverage of 7 × per accession. Raw reads were quality controlled using Trimmomatic (A. M. Bolger et al., 2014) to remove adaptors and low-quality bases. The clean reads were aligned against the reference genome of B. masoniana using BWA-Mem (v.0.7.10) (Li, 2013) with default parameters. Variant detection was performed using the genome analysis toolkit (GATK; v.3.5-0-g36282e4) (Mckenna et al., 2010) following the best practices workflow for variant discovery. The resulting BAM files were locally realigned using the IndelRealigner to remove erroneous mismatches around small-scale insertions and deletions. Variants were called in each accession separately using the HaplotypeCaller and individual gVCF files were merged using GenotypeGVCFs. This two-step approach includes quality recalibration and regenotyping in the merged vcf file, ensuring variant accuracy. Single nucleotide polymorphisms (SNPs) were filtered based on the following criteria: SNPs in repeat regions; SNPs with read depth > 1000 or < 5; SNPs with missing rate > 40%; SNPs with < 5 bp distance with nearby variant sites; and nonbiallelic SNPs were removed. Phylogenetic reconstruction, admixture analysis, principal component analysis (PCA), diversity statistics and ABBA-BABA analysis based on the SNP data are detailed in Methods S1.

Phylogenetic analysis

For nuclear phylogenetic analysis, SNPs within 4000 Begonia single-copy nuclear genes identified using the software OrthoFinder (Emms & Kelly, 2015) with four newly sequenced Begonia genomes with default settings were extracted from vcf files and filtered based on sequence length (> 100 bp) and taxon occurrences (> 50%), aligned with MAFFT (Katoh et al., 2005), and trimmed with GBLOCKS (Talavera & Castresana, 2007). A supermatrix method was used to infer the nuclear phylogeny using RAxML (v.7.2.3) (Stamatakis, 2006). The maximum-likelihood tree inferred from concatenated nuclear SNPs was used as a starting tree to estimate species divergence time using MCMC TREE as implemented in PAML (Yang, 2007). One calibration point of the Begonia crown group (24 million years ago (Ma) ± 3.57 million years with a normal distribution) was defined following Moonlight et al., 2018). For plastid phylogenetic analysis, we newly generated 78 Begonia plastid genomes with NovoPlasty (Dierckxsens et al., 2017) using the seed sequence of rbcL. These plastid genomes were annotated and the conserved 83 plastid protein-coding genes were extracted for phylogenetic inference in Geneious 10.0.2 (Biomatters, Auckland, New Zealand). The concatenated nucleotide dataset was evaluated with PartitionFinder (Lanfear et al., 2012) for the optimal data partition scheme and the associated nucleotide substitution models, with an initial partitioning strategy by both locus and codon positions, resulting in 13 partitions. The concatenated dataset was analyzed using RAxML (v.7.2.3) (Stamatakis, 2006) with 500 bootstrap replicates.

Genome synteny

The syntenic blocks between two species were defined by MScan (Tang et al., 2008) based on core-orthologous gene sets identified by Blastp (E-value ≤ 1e-5; number of gene pairs required to call synteny ≥ 5). The resulting dot plots were inspected to confirm the paleoploidy level of Begonia in relation to the other genomes by counting the syntenic depth in each genomic region. The synonymous Ks value for homologous gene pairs was calculated using the software PAML (Yang, 2007) and a custom perl script (https://ftp.cngb.org/pub/CNSA/data3/CNP0001056/CNS0227982/CNA0013976/), respectively (see details in Methods S1).

Chlorophyll fluorescence analysis

For Chl fluorescence measurement, the plants were dark-adapted for 30 min before the measurements with the MAXI version of the Imaging-PAM M-Series Chl fluorescence system (Heinz-Walz Instruments, Effeltrich, Germany), as described by Jin et al. (2018). For measurements of the light-response curves of photosystem II (PSII) quantum yield (ΦPSII), plant leaves were illuminated at the following light intensities: 0, 1, 21, 56, 111, 186, 281, 336, 396, 461, 531, 611, 701 and 801 μmol photons m-2 s-1. The PSI electron transport rate (ETRI) was measured using light gradients of 0, 5, 13, 31, 89, 167, 209, 325, 496, 754 μmol photons m-2 s-1.

Identification and phylogenetic analysis of light-harvesting Chl a/b-binding proteins superfamily

For the identification of LHCs, all LHCs previously described from Arabidopsis and Oryza sativa (Umate, 2010) were retrieved from the database of TAIR (www.arabidopsis.org) and NCBI (www.ncbi.nlm.nih.gov/protein/), respectively. Representative members of the subfamilies of Arabidopsis were used as queries to perform Blastp searches against the protein database of each species with an E-value cutoff of 1e-5. Candidate sequences
identified as LHC orthologs were then aligned using MAFFT (Katoh et al., 2005) to remove those that did not contain the intact domain (PF00504). For phylogenetic analysis, sequences of LHC orthologs of four Begonias, Crocus sativus, as well as Arabidopsis thaliana and O. sativa were aligned using MAFFT (Katoh et al., 2005), followed by phylogenetic reconstruction with PhyML (v.3.1) (Guindon & Gascuel, 2003).

Results

Genome sequencing and genome characteristics

Seventy-eight Begonia species were sequenced to acquire genome skirn data for comparative genomic studies (Fig. 1; Table S1). As there is already a draft genome for the Neotropical B. fuchsioides (Griesmann et al., 2018), we selected four species, including one from Africa (B. loranthoides, 2n = 38) and three from Asia (B. masoniana, 2n = 30; B. darthvaderiana, 2n = 30; B. peltatifolia, 2n = 30), for reference genome sequencing (Fig. S2). K-mer analyses based on 10 × Genomics Chromium reads data indicated an estimated genome size of c. 724, c. 806, c. 797 and c. 349 Mb for B. loranthoides, B. masoniana, B. darthvaderiana and B. peltatifolia, respectively (Table S2). The genomes of B. masoniana and B. darthvaderiana had the highest heterozygosity levels (Fig. S3), 0.96% and 0.98%, respectively, compared to 0.19% for B. loranthoides and 0.27% for B. peltatifolia (Table S3). We combined multiple sequencing and assembly technologies (Fig. S4; Table S4), including linked reads from stLFR and 10 × Genomics Chromium for four species, PacBio single-molecule real-time (SMRT) to assist in assembly of the more heterozygous B. masoniana and B. darthvaderiana genomes, and Hi-C scaffolding strategies for chromosome assembly of all the four genomes. For B. loranthoides and B. peltatifolia, the genomes were assembled into 716.44 Mb (scaffold N50: 6.73 Mb) and 334.09 Mb (scaffold N50: 3.20 Mb) using 10 × Genomics data, with c. 88.55 and 87.13% of assembled sequences anchored onto 19 and 15 pseudochromosomes, respectively (Tables S4, S5). For B. masoniana and B. darthvaderiana, the genome assemblies yielded 799.40 Mb (contig N50: 0.44 Mb) and 771.67 Mb (contig N50: 0.32 Mb) using PacBio long reads data, with c. 98.83 and 97.55% of the assembled sequences anchored onto 15 pseudochromosomes, respectively (Table S5).

To evaluate the quality of the assemblies, RNA-sequencing reads from root, stem/rhizome, flower, peduncle and leaf tissues were mapped to their cognate assemblies (Table S6). About 90.92–98.83% of the reads were aligned to their corresponding genomes (Table S7). The completeness of the assemblies in terms of gene content was assessed with Benchmarking Universal Single-Copy Orthologs (BUSCO) analysis. Of the core 1614 conserved plant genes evaluated, 97.00, 91.00, 92.20 and 96.80% were complete in the assemblies for B. loranthoides, B. masoniana, B. darthvaderiana and B. peltatifolia, respectively; c. 0.80–2.50% of the genes were fragmented (Table S5). Collectively, these results demonstrated that our four genome assemblies were of high quality in terms of contiguity, base accuracy and genome completeness.

Repeat annotation and gene prediction

Repetitive elements were estimated to represent 66.52%, 68.40%, 70.33% and 51.47% of the genome assemblies in B. loranthoides, B. masoniana, B. darthvaderiana and B. peltatifolia, respectively (Table S5). Most of these repeats were TE s that were further subclassified into nine groups (Table S8). Long-terminal repeat retrotransposons (LTR-RTs) represent 42.80–65.60% of the genome assemblies, with Gypsy elements being the most abundant transposon superfamily in all four Begonia species (30.39–48.60%), followed by the Copia superfamily (7.32–18.36%) (Table S8). The pattern of LTR distribution patterns varied across the genomes for different elements (Fig. 2a). The density of Gypsy scaled negatively with that of the genes whereas Copia was distributed more evenly across the genome and showed no obvious correlation with gene elements (Figs 2a, S5). This is expected since it is known that Gypsy elements accumulate predominantly in heterochromatin and centromeres, whereas Copia elements are normally scattered across the genome (Neumann et al., 2011).

De novo and homology-based approaches were combined to predict protein-coding genes. In total, 22,059, 22,861, 23,444 and 23,010 complete genes were predicted for B. loranthoides, B. masoniana, B. darthvaderiana and B. peltatifolia, respectively, with the highest gene density being 69 genes per Mb in B. peltatifolia, and 28–31 genes per Mb in the other three species (Table S5; Fig. S6), which correlated with the relatively small genome size of B. peltatifolia among the four analyzed Begonia species. The numbers of protein-coding genes are relatively consistent within Cucurbitales (18,292–32,203), but except for B. peltatifolia the gene densities in Begonia (28–31 genes per Mb) are near two-fold lower than those in Cucurbitaceae (64–117 genes per Mb), probably due to higher transposon content (Table S9).

Whole genome duplication and gene evolution

The fraction of synonymous substitutions per synonymous site (Ks) distributions of paralogs clearly showed two peaks (Fig. 2e), one around 1.5 representing the γ hexaploidization event shared by the core eudicots (Jaillon et al., 2007; Chanderbali et al., 2017), and the other around 0.5 indicating that a lineage-specific WGD event occurred in Begonia. By performing a comparative genomic analysis of Begonia with Vitis vinifera, we identified a 2:1 syntenic depth ratio (Fig. 2b), which confirms the WGD previously reported in Brennan et al. (2012). We speculate that the WGD event occurred 35 ± 8 Ma (Fig. 2c; Methods S1), and hence before the split of Begonia (median, c. 25 Ma) and its monotypic sister Hillebrandia, the only other genus of Begoniaceae (Moonlight et al., 2018). This is supported by the fact that Hillebrandia has also been found to possess the WGD, probably indeed shared with Begonia (Martínez, 2017). Following the WGD event, 2850 gene families were retained in the common ancestor of the four species of Begonia we sequenced. The retained gene duplicates shared by the four species were considered as core retained genes. This set was enriched for terms such
Fig. 2 Synteny and lineage-specific whole-genome duplication (WGD) in *Begonia*. (a) Circular view of the *Begonia loranthoides* genome. a: Lines in the inner circle represent links between synteny-selected paralogs. b: Gene density, c: Gypsy density, d: Copia density, e: Gene expression. (b) Syntenic blocks in homologous chromosomes between *B. loranthoides* – *B. masoniana*, *B. masoniana* – *B. darthvaderiana*, and *B. darthvaderiana* – *B. peltatifolia*. (c) A simplified phylogenetic tree showing the lineage-specific WGD in *Begonia*. The other generally accepted WGDs shown are based on Jiao et al. (2011) and Zhang et al. (2020). (d) Macrosynteny patterns show that three typical ancestral regions in the grape genome can be mapped to six regions in the *Begonia* genome. Gray wedges in the background highlight major syntenic blocks spanning > 30 genes between the genomes (highlighted by one syntenic set shown in color). (e) Synonymous substitution rate ($K_s$) distributions of syntenic blocks for the paralogs of four *Begonia*is and orthologs between either two *Begonia*is are shown in different colors, as indicated. Note the $K_s$ unit in the range 1.0–4.0 is ten-fold of that in the range 0–1.0.
as ‘carbohydrate biosynthetic process’ and ‘nucleotide binding’, and many metabolism pathways such as ‘inositol phosphate metabolism’, ‘starch and sucrose metabolism’, and ‘galactose metabolism’ (Fig. S7).

Individual species retained some specific groups of duplicated genes (Figs S8–S11); for instance, genes annotated as involved in the ‘cutin, suberine and wax biosynthesis’ pathway were differentially retained in *B. loranthoides*. This gene retention might be associated with the characteristic waxy leaves of this species. Specific retention of genes involved in ‘phenylpropanoid biosynthesis’ and ‘flavonoid biosynthesis’ might be responsible for the colorful leaves of *B. masoniana* and *B. darthvaderiana* (Fig. S2).

As variegated leaves are commonly found among Begonias and are largely attributed to the accumulation of anthocyanins, we looked more closely at the anthocyanin biosynthesis pathway gene families. We found that in contrast to expansion of gene families of the upstream general phenylpropanoid pathway from Cucurbitaceae, *Begonia* species show significant expansion of gene families related to anthocyanin biosynthesis, especially for *Chalcone synthase* (*CHS*) in *B. masoniana* and *Flavanone 3-hydroxylase* (*F3H*) and *Dihydroflavonol 4-reductase* (*DFR*) in *B. darthvaderiana* (Fig. S12), and there is recent evidence for relaxed selective constraints and differential expression of paralogs in *CHS* in *Begonia* (Emelianova et al., 2021).

Based on a high-confidence phylogenetic tree reconstructed by 193 single-copy nuclear gene families of 13 angiosperm species including *V. vinifera*, *Populus trichocarpa*, *Glycine max*, *Prunus mume* and five species from Cucurbitaceae, we identified gene families that have experienced significant expansions and contractions during the evolution of *Begonia* and related species (Fig. S13). Twenty gene families, including 1071 genes, were significantly expanded (*P* < 0.05) in *Begonia* species compared to the other groups. GO and KEGG enrichment analyses found these to be enriched in terms including ‘zinc ion binding’, ‘transition metal ion binding’ and ‘metal ion binding’ (Table S10), which are primarily involved in the ‘Oxidative phosphorylation’, ‘Endocytosis’ and ‘Pyrimidine metabolism’ pathways (Table S11). Surprisingly, many resistance- and defense-related gene families such as ‘NB-ARC’ were significantly contracted in the *Begonia* lineage. The TIR-NBS-LRR (TNL) disease resistance gene family (Kim et al., 2009) was completely lost (Fig. S14). We looked for expansion of other disease-related genes and found that only the Autophagy 17 (APG17) family showed significant expansion in *Begonia* (Table S12). Other GO terms which were underrepresented in the set of contraction gene families included the ‘protein kinase domain’, ‘Cytochrome p450’ and ‘Terpene synthase’ gene families (Table S13).

**Chromosome evolution**

To reconstruct the evolutionary events leading to current genome structures in *Begonia*, homologous chromosome segments between different species were identified. There were 122 shared syntenic blocks in the four species of *Begonia* sequenced here, which accounted for 74.6%, 78.6%, 67.0% and 74.6% of the *B. loranthoides*, *B. masoniana*, *B. darthvaderiana* and *B. pelatitifolia* genomes, respectively. The lowest percentage of syntenic blocks in *B. darthvaderiana* among those of the four species was consistent with the low TE proportion in these regions compared to *B. masoniana* and *B. loranthoides* (Fig. S15). Synteny analyses between them showed that each chromosome had a nearly one-to-one syntenic relationship with chromosomes from other species (Fig. 2b); the relationship was especially strong for those three species with the same chromosome number. Some large inversions could be inferred for each species. One translocation was detected in chromosomes 2 and 17 in *B. loranthoides*. Chromosome fissions and fusions were identified in the genomes of *B. loranthoides* and *B. masoniana*. We suggest that chr9 and chr12, chr1 and chr18, chr3 and chr11, and chr8 and chr19 in *B. loranthoides* experienced breakage with fusion to chr4, chr1, chr11 and chr15 in *B. masoniana*, respectively (Fig. 2b).

Conserved gene adjacencies suggest that the ancestral *Begonia* karyotype reconstructed based on the four species noted above consisted of 22 conserved ancestral regions (CARs), following an ancestral WGD that occurred early in the history of Begoniaceae characterizing all extant members (Fig. S16). From the 22 CARs of the ancestral karyotype, the 15 chromosomes of *B. masoniana* might be derived by three fusions, and four deletions, while the 15 chromosomes of *B. darthvaderiana* were shaped through one fission, four fusions and three deletions, the 15 chromosomes of *B. pelatitifolia* through four fissions and 11 fusions, and the 19 *B. loranthoides* chromosomes through seven fissions and 10 fusions (Fig. S16). Although *B. pelatitifolia* has the same chromosome number as *B. masoniana* and *B. darthvaderiana*, it appears to have undergone a large number of chromosome fissions and fusions after the split from their common ancestor. This suggests that genomic rearrangements may be even more frequent in *Begonia* than apparent from the highly variable chromosome numbers (*2n* = 16–156) (Dewitte et al., 2009, 2011).

**Transposable elements evolution and distribution**

Transposable elements generally comprise the bulk of plant genomic DNA and their numbers show a positive correlation with genome size (Wendel et al., 2016). In our *Begonia* samples, this also appears to be the case: *B. pelatitifolia* has both the smallest genome and the smallest number and proportion of TEs (Fig. S17). Amongst the most abundant superfamilies of TEs, the number of *Gypsy* and *Copia* LTR elements were most strongly and positively correlated with genome size (Fig. S17). As the four *Begonia* species have similar numbers of protein-coding genes (Table S5), genome size variations between them are essentially attributed to the variation of TE abundance between the different *Begonia* species.

The investigation of TE representation in our four *Begonia* genome assemblies showed that they had different compositions of TE superfamilies (Fig. 3a; Table S8), and are quite variable for full-length *Gypsy* and *Copia* families (Fig. S18). The analysis of full-length LTR-RTs indicated several transposon bursts occurred during the last 8 million years, including recent expansions in all species, especially in *Gypsy* elements compared to *Copia* (Fig. 3b). When the full-length *Gypsy* and *Copia* families were analyzed in
the four *Begonia* species, they showed an expansion event at 0–2 Ma, with the *Reina* subgroup of elements expanding 3–4 Ma in *B. loranthoides* (Fig. 3c).

To determine the historical dynamics of the different lineages of *Gypsy* and *Copia* elements in the *Begonia* genomes, the divergence of their reverse transcriptase (RT) sequences was analyzed. Evolutionary analysis revealed different patterns among different LTR lineages in the four species (Fig. S19). For example, *SIRE* elements of *Copia* show a recent activity burst from a few ancestor sequences in *B. masoniana*, *B. loranthoides* and *B. darthvaderiana*, but no burst is observed in *B. peltatifolia*. A few species-specific bursts were also observed for the *Gypsy Tekay* element in all four genomes investigated (Fig. S19). Furthermore, several *Copia Ivana* and *Gypsy CRM* copies from the common ancestor of the four *Begonia* species we investigated have been maintained and are still active (Fig. S19). These findings show that *Begonia* has a long and ongoing history of active TE elements.

Based on the presence and abundance of TE elements in each species, PCA recovered three well-circumscribed *Begonia* lineages, corresponding to the three major geographical groups of the genus, indicating similar TE compositions in geographically restricted clades (Fig. 3d). Closely related species showed similar TE abundance, even in some species that have diverged more than 10 Ma (Fig. S20). Our investigation reveals a congruence of TE abundance with the phylogenetic tree, indicating that TEs are specifically accumulating across clades of species.

To look for effects of TE activity on gene function, we analyzed TE distribution upstream and downstream of genes. The numbers of genes with adjacent *Copia* and *Gypsy* elements

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**Fig. 3** Transposable element (TE) evolution in the *Begonia* genomes. (a) TE composition of the four *Begonia* genomes. Stacked histograms represent the contribution of each TE superfamily to the four genomes. The eight most abundant TE superfamilies are shown. (b) Estimation of insertion time of *Gypsy* and *Copia* based on analyses of full-length LTRs in four *Begonia* species. (c) Activity of 12 prominent full-length retrotransposon families in four *Begonia* species. Concentric circles indicate timescale per million years from 6 Ma (center) to the present (outer circle). For each family, the circle size shows the retrotransposon number in four species. Each data point represents the peak activity of that element. (d) Principal component analysis (PCA) based on TE abundances of 74 *Begonia* species originated from Africa (green), America (orange) and Asia (red).
insertions was very similar for the four Begonia species (Fig. S21a). However, comparison of other enriched TE families surrounding genes indicated different composition patterns in these four Begonia species (Fig. S21b). About 743–2751 (3.23–12.03%) and 1705–2378 (7.41–10.78%) genes have TE insertions in their intron and promoter regions, respectively (Fig. S22). Functional enrichment analysis of those genes with TE insertions identified stress-related and metabolic process pathways as over-represented in the set (Tables S14, S15), with distinct differences between the basal African lineage represented by B. loranthoides and the three Asian species (Fig. S23).

**Evolution of shade adaptation**

As classical shade-dwelling plants, all Begonias have lower total Chl and lower ratios of Chl/Chl a ana (Fig. 1). As expected, the two deep shade species (B. masoniana and B. darthvaderiana) on phylogenetic relationships, the deep shade adaptations of other plant species (Figs 4d, S24 Begonia and UV Resistance Locus 8 (UVR8) were obviously expanded in Cryptochromes (CRYs), Phototropins (PHOTs), Phytochromes belonging to the core components of light perception; that is, comparative genomic analysis, we found several gene families over-represented in the set (Tables S14, S15), with 2378 (7.41–10.78%) and 1705 other plant species (Fig. S22). Functional enrichment analysis of those genes with TE insertions identified stress-related and metabolic process pathways as over-represented in the set (Tables S14, S15), with distinct differences between the basal African lineage represented by B. loranthoides and the three Asian species (Fig. S23).

**Phylogenomic incongruences and hybridizations**

Species of Begonia are known to hybridize in nature (Peng & Ku, 2009; Hughes et al., 2018), and previous work (Goodall-Copestake et al., 2010) identified possible hybridization events early in Begonia evolution. To investigate this further, we compared phylogenetic inferences between plastid and nuclear phylogenies. The plastid tree supports the African origin of Begonia and shows successive divergences of four major clades, corresponding to the African, Neotropical I, Neotropical II and Asian clades (Figs 5c, S30, S31). Our plastid phylogeny differs from previous phylogenetic studies based on three plastid markers (Moonlight et al., 2018) in the position of the yellow-flowered African Begonia (YFAB) clade. The YFAB clade forms a sister group with the Fleshy-fruited African Begonia (FFAB) clade in our study (Fig. 5c) whereas in previous multilocus studies it diverged at the base of Begonia (Moonlight et al., 2018). The nuclear trees (Figs S32–S34) and TE topology (Fig. S20) in our study consistently recovered a topology with three major geographically restricted clades: the African, Neotropical and Asian clades. Some conspicuous incongruences between nuclear and plastid trees can be identified within the Neotropical clade: the well-resolved EB (East Brazil) clade containing sections Trachelocarpus, Perea, Astrontrix, Solananthera, Gaerdtia and Latistigma in the nuclear tree is split into three independent lineages (EB1, EB2, EB3) diffusely distributed between the two Neotropical clades in the plastid tree (Fig. 5c). The position of the two SDAAB (Seasonally dry adapted African Begonia) accessions also show strong cytonuclear incongruence, suggesting hybridization, introgression or incomplete lineage sorting (ILS).

We observed strong discordance for the Neotropical species in the species tree constructed with ASTRAL-III (Fig. S32).

Genetic variation and admixture patterns

Begonia originated in Africa and spread across all the tropical regions except Australia (Neale et al., 2006). We selected 78 accessions (Fig. 5a) that cover the full distribution of Begonia, representing 37 out of 70 sections, to investigate patterns of genetic variation across the genus. We detected 1 137 696 SNPs and 66 862 small indel variants (< 10 bp). Phylogenetic analysis using a subset of 926 407 SNPs within regions of putatively single-copy genes (SCGs) clearly differentiate Begonia accessions into three distinct clades (Fig. 5c). A weakly supported African clade is sister to a clade consisting of two monophyletic lineages including one consisting of largely Neotropical accessions and one consisting of exclusively Asian accessions.

Genetic clustering analysis with ADMIXTURE showed an optimal value of $K=3$ subpopulations (Fig. 5d), which is consistent with the PCA (Fig. 5b). We observed evidence of interspecific admixture within the Neotropical and African Begonia accessions, respectively, and the highest nucleotide diversity ($\pi$) in the Neotropical accessions (0.0005755) compared with that of the African (0.0002595) and Asian (0.0002434) accessions (Fig. S29).
Hybridization or ILS are possible explanations for this and are also suggested by the SplitSTREE network analysis which revealed a reticulate evolution for these Neotropical accessions (Fig. 5f).

To identify possible causes of genetic introgression among Begonia species, we calculated Patterson’s D-statistics for every triplet (a combination of P1, P2 and P3) in the Begonia phylogeny. The
Fig. 5 Phylogenomic incongruences and hybridization. (a) Geographic localities of sequenced Begonia individuals and a proposed migration route. (b) Principal component analysis (PCA) of the sequenced 78 Begonia accessions. (c) Cytonuclear conflicts between chloroplast (left) and nuclear (right) phylogenetic trees among 78 sequenced Begonia species with three Cucurbitales species as the outgroup. Branches are maximally supported unless otherwise indicated. (d) Ancestry results from ADMIXTURE under the K = 3 model supported by an examination of cross-validation. (e) Detection of introgression events using Patterson’s D statistics among different Begonia species. (f) SPLITSTREE network for 78 Begonia individuals. MB, Malagasy Begonia; EB, East Brazil; YFAB, Yellow-flowered African Begonia; FFAB, Fleshy-fruited African Begonia; SDAAB, Seasonally dry adapted African Begonia; MG, Mexico section Gireoudia.
ABBA-BABA analyses revealed significant introgression in the Neotropical clade between the lineage containing MG (Mexico sect. Gireoudia), and the aforementioned EB clade, the EB clade and the African SDAAB B. wollastonii. We also observed strong genetic introgression between the SDAAB accession B. dreggei and the MG clade (Fig. 5e), suggesting that hybridization and introgression might play some roles in the evolution of the Neotropical Begonia. Phylogen network results suggested a hybrid origin of a clade consisting of B. bullatifolia and B. santos-limae (from the EB clade) from B. radicans and the early Neotropical Begonia colonizers, as well as a hybrid origin of the lineage consisting of Sect. Wageneria and B. soli-mutata (Fig. S35), lending support for the ABBA-BABA introgression results.

In contrast to extensive cytonuclear incongruence and putative hybridization and introgression in the Neotropical clade, only a few topological incongruences were detected within Asian and African clades (Fig. 5e). Nonetheless, significant introgression might have occurred between the MB clade and the FFAB clade (Fig. 5e). One independent introgression event was also inferred for the Asian clade, namely the introgression between B. dipetala and the ancestor of the other Asian Begonias (Fig. 5e). These introgressions were also supported by the corresponding Phylogen network results (Fig. S35). Together, these putative introgression and hybridization events are generally in good agreement with the instances of cytonuclear incongruence and may have played a role in the evolution of Begonia.

Discussion

Putative WGDs have been identified across the eukaryote tree of life, especially in the green plant clade. Many of these WGDs are considered as driving forces contributing to species diversification and evolutionary innovations (Van de Peer et al., 2017; Ren et al., 2018; Leebens-Mack et al., 2019; Wu et al., 2019). WGD may be followed by lineage-specific loss of duplicated genes, contributing to adaptation to new niches, survival in response to environmental stress and subsequent rapid accumulations of species diversity (Landis et al., 2018; Ren et al., 2018; Van de Peer et al., 2021). In this study, we confirmed the occurrence of a lineage-specific WGD event in the common ancestor of Begoniaceae (c. 35 Ma), before the split of cosmopolitan Begonia (median, c. 25 Ma) from the Hawaiian endemic Hillebrandia (Moonlight et al., 2018) (Fig. 2c). As a shade plant, shade adaptation is the key driving force underlying the diversification of Begonia. We provide evidence that the expansion of light signaling pathway genes retained following WGD may have contributed to shade adaptation of Begonia (Fig. 4).

However, WGD is not always associated with species diversification (Landis et al., 2018), as shown in the stark contrast of species diversity between the two genera. The present lack of species diversity in Hillebrandia on the Hawaiian Archipelago is potentially linked to its relict status on the older islands (Clement et al., 2004) and highly homozygous genome (Martinez, 2017). It is tempting to speculate that H. sandwicensis is a dying ember of a once much more species-rich clade, with diversity having been extinguished in the scramble to colonize the archipelago as islands sank and emerged during its geological evolution.

In addition to WGD, hybridization and introgression have also contributed to the species diversity of Begonia. Through population genomic analysis, we detected several putative hybridization events, especially in the Neotropical clade (Fig. 5). These events may have partially contributed to the exceptional species diversity and genetic diversity of Neotropical Begonia though novel combinations of genotypes, introgression and hybridization-based genome rearrangements or TE activation. Further genomic studies on Neotropical Begonia might help elucidate which factors have contributed to this high species diversity.

Plant genomes tend to accumulate large amounts of LTRs, and these have been shown to create different landscapes across closely related taxa. The presence and activity of TEs in plant genomes has been widely observed in many other plant groups, from largely studied taxa such Brassicaceae (Joly-Lopez & Bureau, 2014; Rogivue et al., 2019), Solanaceae (Parisod et al., 2012; de Assis et al., 2020) and Poaceae (Ma et al., 2004; Altinkut et al., 2006; Wyler et al., 2020), to nonmodel plant groups such as Quercus (Mascagni et al., 2019), Passiflora (Sader et al., 2021), Anacyclus (Vitales et al., 2019) or Melampodium (McCann et al., 2020), among many others. We show that transposons are also an important source of genetic variation in Begonia. Two thousand genes in Begonia genomes have TE insertions in their promoter regions (Fig. S22). KEGG functional annotations of these genes with TE insertions in the promoter regions revealed a similar pattern for the three Asian Begonias with enrichment in the pathways of carbohydrate and energy metabolism (Fig. S23). This consistency suggested that TE insertions in the promoter regions might be under some selection constraints rather than neutral and random processes (Baduel et al., 2019). Moreover, the GO enrichment analyses found these genes with TE insertions to be specifically enriched in the function of photosynthesis, negative regulation processes, response to biotic stimulus and stress, and defense response (Tables S14, S15). This result suggests that TE insertions into the regulatory regions in Begonia genomes might play some adaptive role, as has been demonstrated in Arabidopsis (Li et al., 2018; Baduel et al., 2019) and maize (Freeling et al., 2015).

In summary, we have assembled for the first time four chromosome-level genome assemblies of Begonia, and also provide WGS data for 74 representative species within the genus. Through comparative genomics, we confirmed that a lineage-specific WGD event pre-dates the radiation of Begonia and may have provided substantial genetic materials for the phenotypic evolution and shade adaptation. Moreover, we found considerable variation in the compositions and abundance of TEs, and strong phylogenetic signal in TE feature clustering. Species-specific patterns of TE insertions in promoters and introns might have played a role in the adaptive evolution of Begonia. Furthermore, we provide evidence for introgression during the evolution of Begonia, especially for the Neotropical clade. This study not only provides high-quality genomic resources for Begonia, but...
also reveals new insights into the evolution mechanisms of a mega-diverse clade.

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Author contributions

SZ and HL conceived and initiated the study. LL, XC, SD and XG designed the major scientific objectives and managed the project with SZ, HL and DT. HY, XX and Xin Liu coordinated the project. XC, CW, Wenguang Wang and LL conducted the sequencing experiments. DF performed the genome assembly and annotation; XC, LL and SD carried out the repeat analysis. LL, NL, DF, XC, MLIu, WM, ZJL, LZ, TY and JS carried out the comparative genomic analysis and analyzed the gene families. DF, LL, NL, XC, SD, YJ and FC were involved in the WGD analysis. DF, LL, XC, SD, LCD, XG and YG performed the gene annotation and transposable element analysis. SD, XG, NL, DF, XZ, AL, EW and Wei Wang coordinated the phylogenetic analysis. SD, YL, DF, SKS and XC annotated the chloroplast genomes. LL, Xiaofei Liu, CZ and XLang performed Chl fluorescence measurements. LL, YP and LY contribute to cytology analysis. SZ, XLang, Wenguang Wang, Suzhou Zhang, JY and LL coordinated and collected the samples. LL, DF, XG, SD and XC drew and modified the figures. LL, XC, SD, XG, DF, NL, CK, LCD, DCT, DES, YVdp, MH and MLisby wrote and edited most of the manuscript. All authors read and approved the final manuscript. LL, XL, DF, SD, XG and NL contributed equally to this work.

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Data availability

All of the raw sequence data including whole-genome sequencing and transcriptome have been deposited in China National GeneBank Sequence Archive (CNSA) database (https://db.cngb.org/cnsa) under accession number CNP0001056 and National Center for Biotechnology Information (NCBI) under accession number PRJNA791490. The assemblies, gene sequences and annotation data are available at the CNSA database with accession nos. CNA0013973, CNA0013974, CNA0013975 and CNA0013976 for B. dartthadierana, B. loranthoides, B. masoniana and B. petalitifolia, respectively.

References

Fig. S4 Flowchart of sequencing and assembly for the four *Begonia* species.

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Fig. S7 Analyses of post-whole-genome duplication (WGD) retained gene families in *Begonia*.

Fig. S8 Analysis of post-WGD retained gene families specific to *B. masoniana*.

Fig. S9 Analysis of post-WGD retained gene families specific to *B. darthvaderiana*.

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Fig. S13 Gene family expansions and contractions along a dated angiosperm phylogeny of 13 selected species.

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Fig. S16 Reconstruction of the paleognome of four sequenced *Begonia* species.

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Fig. S18 Number of shared full-length LTR families across four *Begonia* species.

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Fig. S34 Coalescent super tree inferred with ASTRAL-III using SNPs in 1343 nuclear single gene trees.

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Table S12 Number of genes in families related to defense in Begonia and other selected genomes.

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Table S17 Comparisons of the gene numbers for the light signaling genes in 10 angiosperm genomes.

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