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Predicting Protein Content in Grain Using Hyperspectral Deep Learning

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Abstract

We assess the possibility of performing regression analysis on hyperspectral images utilizing the entire spatio-spectral data cube in convolutional neural networks using protein regression analysis of bulk wheat grain kernels as a test case. By introducing novel modifications of the well-known convolutional neural network, ResNet-18, we are able to significantly increase its performance on hyperspectral images. Our modifications consist of firstly applying a 3D convolution layer enabling learning of spectral derivatives that 2D spatial convolution is unable to learn, and secondly, the application of a (1 × 1) 2D convolution layer that downsamples the spectral dimension. Analysis of the responses learned by the convolution kernels in our modifications reveals meaningful representations of the input data cube that reduce noise and enable the subsequent ResNet-18 to perform more accurate regression analysis.

1. Introduction

Recently, near-infrared (NIR) hyperspectral imaging (HSI), combining spectroscopy with image analysis, has been applied for grain quality assessment. Caporaso et al. [8] highlights that NIR-HSI combines the advantages of spectroscopy - the rapidity and non-destructive nature - with the possibility of analyzing the spatial distribution of grain quality parameters. Feng et al. [13] mention that deep learning has great potential of using HSI to assess seed quality. Several parameters for grain quality exist, such as Hagberg Falling Number (HFN) [14], grain sprouting, kernel hardness, moisture content, mycotoxin levels, and protein content.

Various studies have shown how NIR-HSI can be applied to solve different grain quality assessment problems. Serranti et al. [24] use NIR-HSI to distinguish between oat and groat kernels and achieve an accuracy of almost 100%. Yu et al. [28] combine NIR-HSI and deep learning employing stacked autoencoders (SAE) and a fully connected network (FCN) to predict the Nitrogen content of oilseed rape leaves. Nitrogen compounds are related to protein content [17, 27]. They achieve an RMSEP of 0.31% on a dataset with ground truth values ranging from 3% – 6% and conclude that deep learning is feasible for HSI. Caporaso et al. [6] use NIR-HSI for HFN regression analysis on batches of wheat grain kernels achieving an RMSEP of 62 s (HFN is measured in seconds) with a dataset with ground truth values ranging from 62 s − 318 s. Apan et al. [2] apply partial least squares (PLS) regression on wheat crops and achieve an RMSEP of 0.50% on a dataset with ground truth values ranging from 9.4% − 16.0%. Caporaso et al. [9] use PLS regression on single wheat kernels to determine the protein content and achieve an RMSEP of 0.94% on a dataset with ground truth values ranging from 6.2% − 19.8%. Pedersen et al. [21] apply PLS regression on single wheat kernels and achieve an RMSEP of 0.47% on a dataset with ground truth values ranging from 6.8% − 17.0%.

We analyze the possibility of combining deep convolutional neural networks (CNNs) with NIR-HSI for regression analysis within bulk grain sample quality assessment. The combination of NIR-HSI and CNN makes it possible to simultaneously utilize spatial and spectral information important for grain quality parameters which are correlated with both chemical and physical changes in grain kernels. In this paper, we use protein content of bulk wheat grains as a test case to examine possible methods for employing regression analysis with CNNs directly on the NIR-HSI data. Thus, the main focus of this article is to extend the current knowledge on deep learning for regression analysis on hyperspectral images of bulk grain kernels. Previous studies have performed regression analysis using CNNs on one-dimensional spectral data [18]. To the best of our knowledge, we are, however, the first to apply end-to-end trained CNNs for regression analysis on hyperspectral images utilizing the complete spatio-spectral data cube.
We present novel architectural modifications to the well-known CNN, ResNet-18 [15], that allows the model to better utilize the spatio-spectral features of hyperspectral images than its original design which was made for RGB images. The modifications are primarily inspired by those from Dreier et al. [12]. We use PLS on the spectral dimension as a baseline for comparison with the original ResNet-18 and our extensions. Additionally, as PLS uses purely spectral information, we also include a comparison with ResNet-18 using purely spatial information by averaging the spectral dimension, yielding a grayscale NIR image.

In this paper we first present, in Sec. 2, the dataset and preprocessing used for our experiments. In Sec. 3, we present the different deep learning models that we investigate in Sec. 4. In Sec. 5, we analyze and provide some interpretation to what the deep learning models learn from the dataset. Finally, we provide a discussion of our results in Sec. 6 and conclusions in Sec. 7.

2. Dataset

2.1. Acquisition

For our study, we use wheat grain obtained from reference samples of harvests from The FOSS European Grain Network (EGN) and The FOSS World Grain Network (WGN). The samples are made available for our study by Foss Analytical A/S [3]. The harvests are divided by year, with between 6 and 10 harvests from different parts of the world belonging to each year. A total of 63 samples from harvests divided non-uniformly over seven years have been processed.

From each harvest, between 100 and 130 grams of grain kernels were randomly sampled without replacement and all sampled kernels were imaged twice in two sequences. A sequence was constructed in the following way: From each grain sample, 4 images of sparsely packed grain were taken with the remaining grain from that sample being used for images of densely packed grain. This typically yielded between 5 and 7 images of densely packed grain per sample for each sequence. The samples were imaged in a random order for each sequence in order to eliminate any systematic bias that might be introduced in the hyperspectral images due to changes in camera or light source characteristics during use. Examples of densely and sparsely packed images can be seen in Figure 1.

The hyperspectral images were acquired with a Specim FX17 [26] line-scan camera with a spatial sampling of 640 pixels and 224 spectral channels in the near-infrared spectrum in the 900 nm - 1700 nm range. The grains samples were presented to the camera in a 3D-printed sample tray that was placed on a conveyor belt. The sample was illuminated using 6 halogen light bulbs from which light was reflected onto the camera’s line of focus by two mirrors, yielding a uniform and very bright illumination of the focus line. The setup can be seen in Figure 2.

The reference protein contents for each grain sample were measured with the FOSS Infratec™Nova [4]. It uses NIR transmission spectroscopy to measure the mean protein content of bulk grain samples which we use as the ground truth value for all hyperspectral image crops from that sample.

2.2. Preprocessing and data augmentation

To account for electronic noise, variations in illumination intensity, and detector response in the camera, the measured intensity spectrum \(I_{i,j}\) for image pixel \((i,j)\) was transformed into an absorbance spectrum

\[
A_{i,j} = -\log \left( \frac{I_{i,j} - D_i}{W_i - D_i} \right),
\]

where \(W_i\) is the mean spectrum of the measured reflectance of the white PTFE foil in the front and back of the sample tray by camera spatial detector pixel \(i\). \(D_i\) is the spectrum of the reflectance of a dark image taken with the camera’s shutter closed as measured by camera spatial detector pixel \(i\). \(D_i\) was measured before imaging of each sample.

The hyperspectral images are cropped using a window size of 128 \(\times\) 128 pixels with a stride of 64 \(\times\) 64 pixels. Following this, we sanitize the dataset to contain only those image crops containing at least 10\% grain pixels. This is determined by a segmentation mask computed by an Otsu threshold [20] on the mean spectrum of each uncropped hyperspectral image and yields a threshold on the mean spectrum that separates background from grain. This procedure yields 17,754 hyperspectral image crops from 13 harvests for test dataset, and 69,706 crops from 50 harvests used for 5-fold cross-validation with each validation split containing samples from 10 different harvests. The test split contains samples with a range of protein contents between 9.975\% and 16.390\%. The training/validation set contains samples with a range of protein contents between 8.660\% and 17.780\%. A histogram of the sanitized crops and the distribution of ground truths for each validation and test split can be seen in Figure 3.

Signoroni et al. [25] offer a review on deep learning for HSI and mention that the main problem for the interpretation of hyperspectral data comes from the curse of dimensionality. In this paper, this issue is addressed by reducing the number of spectral channels in the dataset. The first and last 10 spectral channels are discarded as the Specim FX17 camera has reduced sensitivity near the limits of its spectrum. After this, the spectral channels are downsampled by averaging the closest pairs of channels, which finally reduces the spectral dimension to 102 channels. The
reduction is formalized by
\[
A_{i,j,c}^{\text{reduced}} = \frac{A_{i,j,2c+10} + A_{i,j,2c+11}}{2} \text{ for } c = 0, 1, ..., 101 .
\]
(2)
The results spectral channel width covers a wavelength range of 7.1 nm similar to the 8 nm average spectral resolution of the Specim FX17 camera [26].

For the CNN models, we apply centering and scaling per spectral channel and independently for each different hyperspectral image crop by applying
\[
A_{i,j,c}^{\text{final}} = \frac{A_{i,j,c}^{\text{reduced}} - \mu_c}{\sigma_c} \text{ for } c = 0, 1, ..., 101 ,
\]
(3)
where the channel mean and standard deviation is computed by
\[
\mu_c = \frac{1}{n} \sum_{i=0}^{127} \sum_{j=0}^{127} \mathbb{1}(A_{i,j} = \text{grain}) A_{i,j,c} \quad \text{for } c = 0, 1, ..., 101 ,
\]
(4)
\[
\sigma_c = \sqrt{\frac{\sum_{i=0}^{127} \sum_{j=0}^{127} \mathbb{1}(A_{i,j} = \text{grain}) (A_{i,j,c} - \mu_c)^2}{n}} \quad \text{for } c = 0, 1, ..., 101 ,
\]
(5)
where \( n = \sum_{i=0}^{127} \sum_{j=0}^{127} \mathbb{1}(A_{i,j} = \text{grain}) \) and \( \mathbb{1}(A_{i,j} = \text{grain}) \) is the segmentation mask for the image crop as computed by the Otsu threshold. Thus, the centering and scaling is done relative to the grain spectra regardless of the amount of background in any given image crop.

The PLS models predict the protein content of each image crop based on the average spectra of pixels containing grain. Prior to averaging the spectra of each image crop, a standard normal variate (SNV) [5] standardization of each pixel spectrum was applied, followed by a transformation of the spectrum into a derivative spectrum using a Savitzky–Golay (SG) smoothing [23]. The pre-processing is applied to mitigate the effect of light scattering artifacts, primarily from Rayleigh scattering occurring when the radiation interacts with particles that are small compared with the wavelength of the light [22]. The best prediction result was found using a 2nd derivative SG transformation with a 3rd order polynomial and kernel width of 7 reduced spectral channels.

We apply augmentation to the training set by rotating each hyperspectral image crop by \{0, 90, 180, 270\} degrees and flipping them left-right. This increases the variability of the training data and thus increases the robustness of the model. The augmentation is done uniformly at random and independently for each hyperspectral image crop and each epoch.

Figure 1: Examples of densely (left) and sparsely (right) packed grain images.

3. Models

All models are modifications of the ResNet-18 CNN model from the well-known family of ResNet models [15]. The modifications consist of adding new layers before the original ResNet-18; changing the ordering of rectified linear units (ReLUs) and batch normalization (BN) layers such that BN layers come after ReLU instead of before to avoid having the weights updated in a suboptimal way as argued by Chen et al. [10]; and changing the final layer to be a fully connected layer with a single neuron using linear activation, enabling interpretation of the output as a regression prediction. We compare the raw ResNet-18 model with models containing different \( 1 \times 1 \) 2D convolution layers, called downsamplers, that serve to downsample the spectral dimension. For some models, we also add a 3D convolution layer before the downsamplers to facilitate the learning of spectral gradients such as those computed by SG derivative filters [23] which are paramount to apply for the success of PLS [22]. In the case that we apply 3D convolution as the initial layer, the output is 4D. However, as the downsamplers expect 3D input, we concatenate the features yielded by each of the 3D convolution filters to reduce the 4D tensor to 3D. The downsamplers and 3D convolution layers have linear activation and a BN layer is applied after each for training stability. The 3D convolution layers use linear activation to mimic the linearity of the SG derivative filters. We experimented with using ReLU as activation for the downsamplers but that did not lead to increased performance.

We compare these spatio-spectral models with models utilizing purely spatial or purely spectral information. For using purely spatial information, we average the spectral dimension of the hyperspectral image crop, yielding a grayscale image crop, and feed it to the ResNet-18 skeleton. For using purely spectral information, we perform PLS
regression on the mean grain spectrum of the hyperspectral image crop using 3 components which compares to using 3 downsamplers in the CNNs. An overview of the neural models used in this paper is shown in Table 1.

### 3.1. Model implementation

The CNN models are implemented using the deep learning libraries, Keras [11] and TensorFlow [1]. Similar to the original implementation of ResNet [15], we use stochastic gradient descent (SGD) with a momentum of 0.9, L2-regularization with a regularization parameter of $10^{-4}$, and initialize all weights with the Kaiming He Normal Distribution [16]. The loss function is root mean squared error (RMSE). The learning rate is multiplied by a factor of 0.1 if validation loss plateaus for 4 epochs. Training halts if the validation loss plateaus for 12 epochs or if the number of epochs exceeds 50 - in practice, all training runs were halted by the plateau criterion. We train with a mini-batch
Here, we analyze both of these layers. In Figure 5, we show not. Furthermore, applying a 3D convolution filter before employing downsamplers perform better than those that do.

5. Analysis

Table 1: Overview of the layers applied before the ResNet-18 skeleton. Conv3D filters are always applied before downsamplers if they are both used.

<table>
<thead>
<tr>
<th>Model</th>
<th>Conv3D filters, kernel size</th>
<th>Downsampler filters</th>
</tr>
</thead>
<tbody>
<tr>
<td>Model 1</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>(Grayscale input)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Model 2</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>Model 3</td>
<td>None</td>
<td>3</td>
</tr>
<tr>
<td>Model 4</td>
<td>1, (1 × 1 × 7)</td>
<td>3</td>
</tr>
<tr>
<td>Model 5</td>
<td>3, (1 × 1 × 7)</td>
<td>3</td>
</tr>
<tr>
<td>Model 6</td>
<td>1, (7 × 7 × 7)</td>
<td>3</td>
</tr>
<tr>
<td>Model 7</td>
<td>3, (7 × 7 × 7)</td>
<td>3</td>
</tr>
</tbody>
</table>

Table 2: Mean RMSE ± standard error on the mean on the validation and test sets for each of the models. (best on val) is defined as the model which achieved the lowest RMSE on its validation set.

<table>
<thead>
<tr>
<th>Model</th>
<th>Validation [%]</th>
<th>Test [%]</th>
<th>Test [%] (best on val)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Model 1</td>
<td>1.23 ± 0.07</td>
<td>1.44 ± 0.04</td>
<td>1.45</td>
</tr>
<tr>
<td>Model 2</td>
<td>1.09 ± 0.11</td>
<td>1.21 ± 0.08</td>
<td>0.97</td>
</tr>
<tr>
<td>Model 3</td>
<td>0.98 ± 0.09</td>
<td>1.05 ± 0.07</td>
<td>0.98</td>
</tr>
<tr>
<td>Model 4</td>
<td>1.91 ± 0.56</td>
<td>434 ± 384</td>
<td>2152</td>
</tr>
<tr>
<td>Model 5</td>
<td>0.85 ± 0.06</td>
<td>0.90 ± 0.05</td>
<td>0.83</td>
</tr>
<tr>
<td>Model 6</td>
<td>0.89 ± 0.06</td>
<td>0.99 ± 0.03</td>
<td>1.03</td>
</tr>
<tr>
<td>Model 7</td>
<td>0.85 ± 0.06</td>
<td>0.92 ± 0.04</td>
<td>0.82</td>
</tr>
<tr>
<td>PLS</td>
<td>0.76 ± 0.02</td>
<td>0.75 ± 0.01</td>
<td>0.77</td>
</tr>
</tbody>
</table>

The responses learned by the three (1 × 1 × 7) convolution filters from Model 5 validated on split 4. The filters seem to approximate spectral gradient filters, which implies that the model is able to learn representations of the input hyperspectral image crop that is similar to the SG derivative filter applied in the preprocessing for the PLS models.

The responses learned by the three downsampling filters applied by the same model following the 3D convolution filters are shown in Figure 6. As each 3D convolution filter yields an output that consists of a learned representation of the entire input hyperspectral image, the concatenation of these outputs will have three consecutive representations of the input hyperspectral image. Each (1 × 1) kernel in the downsamplers is applied to a specific wavelength offset, consisting of the response learned by the (1 × 1 × 7) 3D convolution filter applied at that wavelength. Nitrogen (N) compounds are related to protein content [17, 27]. Therefore, in Figure 7 we zoom in on the last third of the downsampling filters from Figure 6. These parts of the downsampling filters operate on the representation of the hyperspectral image as output by Filter 3 in Figure 5. Here we have plotted the expected wavelength centers of light absorption caused by NH and CH overtones in the 938 nm - 1662 nm range (this is the spectrum remaining after we cut off the first and last 10 channels of the Specim FX17) [19] along with the responses learned by the three downsampling filters. The downsamplers learn spikes in their responses near these expected wavelength positions of NH overtones. However, they also learn spikes in their responses at other wavelengths, noticeably, where CH overtone bands are located (between 1100 nm and 1250 nm, between 1350 nm and 1500 nm, and above 1600 nm) [19]. Similarly, absorbance values located at wavelengths related to CH overtones are also found to be of importance in the PLS model used for purely spectral prediction in this paper as well as previous attempts of predicting protein content with NIR.
spectroscopy and PLS [7], indicating that CH content correlates with protein content.

We examine the representation of the hyperspectral image crops from training split 4 as output by Model 5 (trained and validated on split 4) after its downsamplers in Figure 6. We use the data from the model’s training to analyze if the model is able to learn to distinguish protein content at this early stage. As the downsampling layer utilizes three $(1 \times 1)$ convolution filters, the output will have each spatial pixel represented by a 3D vector, that is a reduced representation of the 102 input wavelength channels. In Figure 8a we perform a PCA on this representation on a representative subset of training split 4 to reduce the representation of each spatial pixel to two dimensions. Here, we color black the spatial pixels containing background and color the spatial pixels containing grain according to their protein content. Clearly, the variance between background and foreground dominates the principal components. Therefore, we apply PCA only to the subset spatial pixels containing grain of the same data and plot this in Figure 8b. We observe that this representation shows that there is a gradient from low protein contents (blue) near the edges of the cluster through middle protein contents (red) to high protein contents (green) in the center.

6. Discussion

Considering the results shown in Table 2 and Figure 4 it is evident that the modified ResNet does not achieve a better RMSE than PLS. While the predictions of Model 1 in Figure 4a (pure ResNet-18 with grayscale input) reveal
that there is some correlation between spatial feature distribution and protein content indicated by the Pearson correlation coefficient of $\rho = 0.76$, the PLS model using only spectral information vastly outperforms Model 1. Model 2 (pure ResNet-18 with spatio-spectral input) outperforms Model 1 with a large margin but is outperformed with an even larger margin by our modified ResNet-18 from Model 5, indicating that our modifications to ResNet-18 are highly favorable.

From the distribution of the average predictions shown in Figure 4 it is evident that the variance in the mean predictions of the same model validated on different splits is relatively higher for the CNN models when compared to PLS. This indicates that the CNNs will benefit from a larger dataset and have not yet reached their full potential.

For PLS, we apply the segmentation mask yielded by the Otsu threshold in order to determine which pixels contain grain and which contain background. For the CNN models, we do not apply the segmentation mask. This implies that the CNNs have to learn to distinguish foreground from background. We experimented with applying the mask to nullify the spectra of any spatial pixels containing background. This did, however, not lead to increased performance, but warrants more future investigation.

Indeed, analyzing our modifications to ResNet-18 revealed that they were able to autonomously learn highly relevant responses to the problem on which they were applied including the segmentation of foreground and background but also the learning of spectral derivatives which have to be manually applied for PLS. Thus, from an end-to-end learning perspective, our modified ResNet-18 is able to extract relevant spatio-spectral features for the application and solve it to a reasonable performance level without the need for application of any advanced pre-processing or prior knowledge of the problem. Additionally, this indicates that it may not be necessary to swap the model or the simple minimal preprocessing that we apply in order to perform regression analysis on a different parameter than protein content, although it may be necessary to fine-tune the number of downsamplers used.

Although our modified ResNet-18 provides a clear advantage over the pure ResNet-18, we expect that its full potential is yet to be uncovered on other problems where there is a higher correlation between spatial features and the tar-
get parameter of interest than for protein content.

7. Conclusion

The work presented in this paper shows that CNNs can be applied to hyperspectral images for regression analysis on bulk wheat grain kernels and achieve reasonable results. While seemingly not as relevant as the spectral information, we show that there is some correlation between the spatial features of wheat grain kernels and their protein content. We applied ResNet-18, which is known to perform well on RGB images, to hyperspectral images. By incorporating the two concrete extensions of ResNet-18 presented in this work, we are able to significantly boost its performance to a level that is close to the performance achieved by PLS which is known to solve the problem of protein content regression analysis well. We show that our extensions of ResNet-18 are able to learn representations of the input data that resemble the preprocessing that humans with extensive knowledge of the problem would apply before applying PLS to perform regression analysis. Furthermore, we show that our extensions are able to learn representations of the hyperspectral data that are highly relevant to the task of protein regression analysis on which they are applied. This end-to-end approach indicates that using our modifications, CNNs should be applied to other regression problems of interest within NIR-HSI, especially where both spatial and spectral information is relevant.

References


Figure 8: PCA reductions of the output of the downsamplers shown in Figure 6 using pixels with grain and background (a) and using only pixels with grain (b). The protein content in % is measured by the Infratec™Nova.


