The Transcriptional Network That Controls Growth Arrest and Macrophage Differentiation in the Human Myeloid Leukemia Cell Line THP-1

Gazova, Iveta; Lefevre, Lucas; Bush, Stephen J.; Clohisey, Sara; Arner, Erik; de Hoon, Michiel; Severin, Jessica; van Duin, Lucas; Andersson, Robin; Lengeling, Andreas; Hume, David A.; Summers, Kim M.

Published in:
Frontiers in Cell and Developmental Biology

DOI:
10.3389/fcell.2020.00498

Publication date:
2020

Document version
Publisher's PDF, also known as Version of record

Document license:
CC BY

Citation for published version (APA):
The Transcriptional Network That Controls Growth Arrest and Macrophage Differentiation in the Human Myeloid Leukemia Cell Line THP-1

Iveta Gažová1†, Lucas Lefevre1†, Stephen J. Bush1†, Sara Clohisey1, Erik Amer2, Michiel de Hoon2, Jessica Severin2, Lucas van Duin3, Robin Andersson3, Andreas Lengeling4, David A. Hume5* and Kim M. Summers1,5*

1 The Roslin Institute, The University of Edinburgh, Edinburgh, United Kingdom, 2 RIKEN Center for Integrative Medical Sciences, Kanagawa, Yokohama, Japan, 3 Bioinformatics Centre, University of Copenhagen, Copenhagen, Denmark, 4 Max Planck Society – Administrative Headquarters, Munich, Germany, 5 Mater Research Institute – University of Queensland, Translational Research Institute, Brisbane, QLD, Australia

The response of the human acute myeloid leukemia cell line THP-1 to phorbol esters has been widely studied to test candidate leukemia therapies and as a model of cell cycle arrest and monocyte-macrophage differentiation. Here we have employed Cap Analysis of Gene Expression (CAGE) to analyze a dense time course of transcriptional regulation in THP-1 cells treated with phorbol myristate acetate (PMA) over 96 h. PMA treatment greatly reduced the numbers of cells entering S phase and also blocked cells exiting G2/M. The PMA-treated cells became adherent and expression of mature macrophage-specific genes increased progressively over the duration of the time course. Within 1–2 h PMA induced known targets of tumor protein p53 (TP53), notably CDKN1A, followed by gradual down-regulation of cell-cycle associated genes. Also within the first 2 h, PMA induced immediate early genes including transcription factor genes encoding proteins implicated in macrophage differentiation (EGR2, JUN, MAFB) and down-regulated genes for transcription factors involved in immature myeloid cell proliferation (MYB, IRF8, GFI1). The dense time course revealed that the response to PMA was not linear and progressive. Rather, network-based clustering of the time course data highlighted a sequential cascade of transient up- and down-regulated expression of genes encoding feedback regulators, as well as transcription factors associated with macrophage differentiation and their inferred target genes. CAGE also identified known and candidate novel enhancers expressed in THP-1 cells and many novel inducible genes that currently lack functional annotation and/or had no previously known function in macrophages. The time course is available on the ZENBU platform allowing comparison to FANTOM4 and FANTOM5 data.

Keywords: macrophage, monocyte, THP-1 cells, differentiation, transcriptome, cell cycle, p53
INTRODUCTION

11q23 chromosomal translocations fusing the mixed lineage leukemia (MLL) gene (official gene symbol: KMT2A) with any one of >60 partners lead to aggressive leukemias with poor prognosis (Liang et al., 2017; Prange et al., 2017). One of the two most common fusion partners AF9 (official gene symbol: MLLT3) is involved in the primary translocation in the widely studied acute myeloid leukemia (AML) cell line, THP-1. THP-1 cells were originally established from peripheral blood cells of a 1 year old boy with an AML-M5 leukemia carrying a t(9;11)(p22;q23) translocation (Tsuchiya et al., 1980). A subsequent study demonstrated that upon treatment with phorbol esters, THP-1 cells become adherent and acquire many functional characteristics of mature macrophages (Tsuchiya et al., 1982). Partial differentiation could also be elicited in these cells with the therapeutic agonist all-trans-retinoic acid (ATRA) (Chen and Ross, 2004). Antisense down-regulation of the MLL-AF9 fusion transcript in THP-1 cells leads to growth inhibition but does not drive differentiation (Pession et al., 2003; Suzuki et al., 2009). On the other hand, differentiation with either phorbol myristate acetate (PMA) or ATRA was associated with down-regulation of MLL-AF9 and growth inhibition (Pession et al., 2003; Chen and Ross, 2004).

The KMT2A/MLL gene encodes an epigenetic modifier, histone lysine methyl-transferase 2A (Hess, 2004). The oncogenic activity of the MLL-AF9 fusion product is mediated in part through another epigenetic modifier, DOT1L, a histone 3 lysine 9 (H3K9) methyltransferase that in turn regulates target genes in the HOX gene clusters and MEIS1 (Nguyen et al., 2011). Genome-wide analysis of MLL-AF9 binding in THP-1 cells revealed a substantial overlap with enhancers bound by RUNX1, a transcription factor that regulates myeloid differentiation and is itself commonly involved in leukemogenic translocations (Prange et al., 2017). These studies identified a novel target of MLL-AF9, the transcription factor ZNF521. In mice, ZNF521 was enriched in hematopoietic stem cells (HSC) and germ line mutation impacted stem cell self-renewal. Knockdown of ZNF521 in THP-1 cells led to cell cycle arrest and partial differentiation (Garrison et al., 2017; Germano et al., 2017). Other genes that apparently contribute to dysregulated proliferation downstream of MLL-AF9 in either THP-1 cells or in mouse models include those encoding the transcription factor SALL4 (Yang et al., 2017) and the protooncogene EVI1 (Bindels et al., 2012).

Differentiation therapy involves forcing cells to cease proliferation and undergo terminal differentiation (Sachs, 1982). Such therapy with ATRA is one of the success stories in leukemia treatment but is applicable to only around 10% of AML cases (Ma et al., 2017). THP-1 cells provide a model system to investigate other potential differentiation therapy agents in aggressive AML. The process of differentiation of THP-1 cells has been studied in detail at the transcriptomic level as a model both of inhibition of leukemic proliferation and of macrophage differentiation. Differentiated THP-1 cells are commonly used as a tractable model for human monocytes (Bosshart and Heinzemann, 2016), recently exploited in functional genomics using CRISPR-Cas9 deletion (Goetze et al., 2017; Osei Kuffour et al., 2018; Palazon-Riquelme et al., 2018).

The original THP-1 line became adherent in response to PMA within 3 h, but with progressive adaptation to tissue culture the cells became more resistant to differentiation with adherence delayed until 48 h of stimulation (Tsuchiya et al., 1982). The line is epigenetically unstable; the relative proportion of cells expressing markers such as CD4 (associated with undifferentiated cells) and undergoing differentiation in response to PMA changes with time in culture (Cassol et al., 2006). Subclones can be selected from the parent line currently available from ATCC that restore the original phenotype and either do, or do not, respond to PMA. In order to study the process of differentiation in a population in which the majority of cells respond synchronously, the FANTOM4 consortium cloned THP-1 cells obtained from ATCC by limiting dilution and chose one subclone in which >90% of cells became adherent within 48 h of addition of PMA (Suzuki et al., 2009).

Alongside microarrays, the consortium used CAP Analysis of Gene Expression (CAGE) to identify regulated promoters across a time course of differentiation. These studies identified a cohort of transcription factor genes rapidly down-regulated following PMA addition. SiRNA knockdown of a subset of these genes (CEBPG, CEBPA, FLI1, GFI1, HOXA9, MYB and the oncogenic MLL-AF9 fusion transcript) produced changes in gene expression that partly mimicked the effects of PMA (Suzuki et al., 2009). A subsequent study revealed combinatorial impacts of several inducible miRNAs that also contribute to cell cycle arrest (Forrest et al., 2010).

The central conclusion of the FANTOM4 analysis (Suzuki et al., 2009) was that numerous regulated genes contribute to a complex network in which reduced expression of anti-differentiation/pro-proliferation genes is as essential as increased expression of regulators that promote differentiation. The FANTOM5 consortium extended the use of CAGE to generate a promoter-based transcriptional atlas for humans and mice (Forrest et al., 2014) and recognized that with sufficient depth of sequencing, CAGE could also detect RNAs derived from active enhancers, termed eRNAs (Andersson et al., 2014). CAGE profiling enabled analysis of enhancer profiles of human monocyte subsets (Schmidl et al., 2014) and a dense time course of the response of human monocyte-derived macrophages to lipopolysaccharide (Baillie et al., 2017). In the macrophage time course, and in several other systems studied (Arner et al., 2015) a transient pulse of eRNA from transcribed enhancers was detected prior to the detection of promoter activity of inducible genes.

One limitation of the earlier FANTOM4 study of THP-1 differentiation (Suzuki et al., 2009) was that the depth of sequencing of CAGE libraries was not as high as in subsequent studies so that transcription from enhancer loci could not be detected. In addition, the time course was insufficient to support resolution of the temporal of waves of transcriptional regulation. The FANTOM5 studies (Forrest et al., 2014) prompted us to reanalyze the time course of differentiation of THP-1 cells using CAGE with a view to further dissection of regulated promoters, enhancers and the complex transcriptional network that drives differentiation in this model.
The cellular commitment to differentiation or division usually occurs in the G1 phase of the cell cycle and differentiation of THP-1 cells is associated with cell cycle inhibition, with accumulation of cells in G1 and in G2/M (Chen and Ross, 2004; Gray et al., 2005; Gutsch et al., 2011). The FANTOM4 analysis revealed down-regulation of cell cycle related genes in response to PMA, with promoters enriched for E2F motifs (Suzuki et al., 2009). A set of 700 genes transcriptionally regulated in the human mitotic cell cycle has recently been generated based upon transcriptional network analysis of multiple datasets, including FANTOM5, and detailed curation and validation (Giotti et al., 2018). Some of these genes have more than one promoter and have other functions unrelated to the cell cycle in specific cell types. The set includes many genes associated with DNA repair and feedback regulation. Since the hallmark of leukemia is dysregulated proliferation we have examined in detail the coordinated expression of these cell cycle genes in THP-1 cells during PMA-induced growth inhibition. The objective was to further validate some of the novel cell cycle-related genes and to identify potential loss or gain of genes or gene expression within this co-expression cluster that might contribute to leukemogenesis and could assist with the development of differentiation therapies.

**MATERIALS AND METHODS**

**Cell Culture and RNA Extractions**

THP-1 cells (high differentiation clone 5, from the FANTOM4 consortium, passage number 8 (P8), provided by Dr. Mark Barnett, The Roslin Institute, United Kingdom) were cultured in sterile-filtered (0.2 μm) RPMI1640 medium with HEPES modification (25 mM HEPES + NaHCO₃) (RS886 Sigma-Aldrich, Gillingham, United Kingdom), 10% heat inactivated fetal bovine serum (FBS, GE Healthcare, PAA laboratories, Pasching, Austria), 1x Glutamax (35050-038, Gibco), 1 mM sodium pyruvate (11360-039, Gibco), 1x heat inactivated fetal bovine serum (FBS, GE Healthcare, PAA laboratories, Pasching, Austria), 1x Glutamax (35050-038, Gibco), and 1x MEM non-essential amino acids solution (11140-035, Gibco), 50 μM 2-mercaptoethanol (31350-038, Gibco) and penicillin-streptomycin (16 U/ml and 16 μg/ml, Gibco) (THP-1 medium). Cells were incubated at 37°C, 5% CO₂ and their concentration was maintained between 2 × 10⁵ – 8 × 10⁵ cells per ml by splitting in half three times a week. For the THP-1 differentiation time course, THP-1 cells were grown up at a low passage number (typically 2 or 3 passages from the P8 stock). The day before the start of the differentiation, cells were counted by hemocytometer and for each time point between 3 × 10⁶ and 5 × 10⁶ cells were pelleted and resuspended in 10 ml fresh medium. THP-1 cells were then differentiated by adding 30 ng/ml (48.6 nM) phorbol 12-myristate 13-acetate (PMA; P1585, Sigma-Aldrich) in DMSO. The cells were plated on a Sterilin bacteriological plate and at appropriate time points, the cells were lifted off by flushing with a blunt-end needle syringe, resuspended in 1 ml of RNA Bee (AMS Biosciences, Frienswood, TX, United States) and frozen at −80°C for later extraction. Medium was not changed for the duration of the experiment to avoid initiation of a serum-specific response during the time course. For each experiment, one culture was used to set up two batches of cells. The first batch was treated with PMA on the morning of the first day. Time points between 0 and 12 h were taken from this batch. The second batch from the same culture was treated with PMA in the evening of the first day (which is the 0 h time point from this batch, taken at the same time as the 12 h time point of the first batch). All cells were then incubated overnight. On the morning of the second day, the 24 h time point of the first batch and the 12 h time point of the second batch were taken, followed by time points between 12 and 24 h taken from the second batch. Batches had several overlapping time points (0, 12, 24, and 36 h) to ensure that cells were at similar state of differentiation. RNA was extracted from the RNA Bee lysate according to manufacturer's instructions. Extracted RNA was treated with DNase I (Ambion DNase kit AM1906, Thermo Fischer Scientific) and the quantity and quality assessed using a NanoDrop spectrophotometer ND-1000 (Nanodrop technologies, Wilmington, DE, United States) and Agilent RNA ScreenTape System (Agilent Technologies, Santa Clara, CA, United States) according to manufacturers’ instructions. Only RNA samples with RIN® values (Imbeaud et al., 2005) around or above 8 (out of 10) were used for CAGE sequencing.

**Cap Analysis of Gene Expression (CAGE)**

Time points chosen for CAGE sequencing were hourly from 0 to 12 h, two hourly from 12 to 18 h, and then at 24, 36, 48, and 96 h. At least three replicates of each time point were included. Eight samples of 5 μg RNA each were made into one CAGE library following the protocol adapted from Takahashi et al. (2012). All the primer sequences were taken from Takahashi et al. (2012) and primers were synthesized by IDT (Coralville, IA, United States). A detailed description of the CAGE protocol and associated bioinformatic analysis is provided in Supplementary File S1. Results for 66 samples were processed using the R/Bioconductor package CAGEr (Haberle et al., 2015). All 66 samples were normalized to a power law distribution (Supplementary File S1), with expression levels presented as tags per million (TPM). The full quality assurance process for CAGE sequencing is outlined in Supplementary File S1. TSS were mapped to the human genome, version GRCh38.p9. The normalized and aggregated groups of TSS (clusters of transcription start sites; CTSS) were then annotated with gene names as described in Supplementary File S1.

**Network Analysis of Gene Expression**

Patterns of gene expression during differentiation were visualized using the network analysis tool Graphia Professional version 2.0.1. Mean expression values across replicates for CTSS with at least 10 TPM in at least one sample were uploaded into the Graphia Professional software. Sample-to-sample analysis was performed using a correlation coefficient threshold of r ≥ 0.95, as indicated in the Results. Two samples, both at the 4 h time point, were found to be outliers, probably due to low level of mapped reads (2.7 and 6.2 × 10⁶ reads). These were removed from subsequent analysis. Gene-to-gene analysis was performed using a correlation coefficient of r ≥ 0.75. Clusters of genes with
similar expression patterns were determined with the Markov Clustering Algorithm (MCL) at an inflation value (determining granularity of the clusters) of 1.7. A list of genes associated with cell proliferation was obtained from Giotti et al. (2018) and a list of human transcription factors from Lambert et al. (2018). Further network analysis was performed on these subsets of genes, with correlation coefficients and inflation values as described in section “Results”. DAVID (Database for Annotation, Visualization and Integrated Discovery) was used to identify functionally enriched biological themes based on gene ontology (GO) annotations, using the Functional Annotation Clustering tool (Huang et al., 2007). High annotation scores indicate significant enrichment for groups of related GO terms.

Analysis of Transcription Factor Binding Motifs
Activities of transcription factor binding motifs were estimated using MARA (Suzuki et al., 2009) using all CTSS with at least 10 TPM in at least one sample, excluding the mitochondrial chromosome and considering conserved motifs in the region −300 to +100 bp of the midpoint of the CTSS. Motif names are based on the set of consolidated position weight matrices for human transcription factors curated for the earlier project (Suzuki et al., 2009), available at http://swissregulon.unibas.ch/sr/fantom4.

Analysis of Putative Enhancer Expression
Candidate enhancers were identified as bidirectional TSS as described (Andersson et al., 2014) at two levels of stringency. The more stringent set (~1000 loci) required at least two libraries with more than one CAGE tag for each CTSS, and for the enhancers an absolute directionality score of less than 0.8, and bidirectional tags in more than two samples. For the less stringent set (~2500 loci), the criteria were >1 tag per CTSS in at least one sample, and bidirectional tags for each enhancer in at least one sample. All bidirectional TSS regions closer than 500 bp to an annotated TSS and closer than 100 bp to an exon were removed to avoid detection of promoters of protein-coding genes. The analysis was done with use of the CAGE analysis package CAGEfightR (Thodberg et al., 2019), and the ported enhancer detection scripts from Andersson et al. (2014).

Validation of Gene Expression Levels With Quantitative Reverse Transcriptase PCR
Gene expression levels detected by CAGE were validated for representative genes using quantitative reverse transcriptase polymerase chain reaction (qRT-PCR) on the same RNA samples. cDNA was synthesized using Superscript III Reverse Transcriptase (Invitrogen) according to manufacturer’s instructions, using 500 ng of total RNA and 100 ng of random primers. The ribonuclease inhibitor used was RNasin Plus (Promega, Madison, WI, United States; 20 units per reaction) and extension at 50°C was for 60 min. The resulting cDNA was diluted 1:1 with water. Standard curves were established using four concentrations of starting RNA: 12.5, 6.25, 3.13, and 1.56 ng/µl. qRT-PCR was carried out using SYBRGreen (Roche, Mannheim, Germany) in a Roche Light Cycler 480 according to manufacturer’s instructions. Samples were pre-incubated at 95°C for 5 min (ramp rate 4.40°C/s), then amplification steps were repeated for 45 cycles. Amplification steps were 95°C for 10s (ramp rate 4.40°C/s), 60°C for 15 s (ramp rate 2.20°C/s) and 70°C for 30 s (ramp rate 4.40°C/s). Afterward, the melting curve for primers was measured by incubating at 95°C for 5s (ramp rate 4.40°C/s), 65°C for 1 min (ramp rate 2.20°C/s) and then the temperature was increased to 97°C by 0.11°C/s. At the end, the plate was cooled for 30 s at 40°C. Calculations were performed using the Roche Light Cycler 480 software, with the Advanced Quantification setting. ΔCt was calculated with previously established values of primer efficiencies from the standard curves (calculated using the same software, by using Abs Quant/2nd Derivative Max setting). Primer sequences and efficiencies are available in Supplementary File S1.

Analysis of the Cell Cycle in Differentiating THP-1 Cells
Propidium iodide staining was used to examine the cell cycle. 1 × 10⁶ cells were pelleted at 400 g for 5 min and resuspended in 1 ml of ice-cold 70% ethanol and then left at 4°C for at least 24 h (up to 2 months). On the day of the flow cytometry, cells were spun down at 16,000 g for 5 min and then washed twice with PBS. 300 µl of the Cell Cycle Staining Solution [38 mM sodium citrate, 10 µg/ml pancreatic RNase A (Sigma-Aldrich), 68 µM propidium iodide (Sigma Aldrich)] was added to the pellet. Samples were incubated in the dark for 1–2 h and 300 µl of PBS was added right before flow cytometry. Samples were analyzed on a Fortessa cytometer (BD Biosciences, Franklin Lakes, NJ, United States) using a 610 nm laser. Results were analyzed using FlowJo software version 10.0.8r1 (Flow Jo LLC, Ashland, OR, United States) by gating single viable cells by excluding doublets. The final window showed the histogram of the area of propidium iodide channel, which was examined using the cell cycle option of FlowJo software by choosing the Watson (pragmatic) type of analysis (Watson et al., 1987).

RESULTS

Characterization of the THP-1 Time Course of Differentiation
PMA treatment of THP-1 high differentiation clone 5 cells was carried out multiple times to harvest RNA for CAGE sequencing. To ensure that differentiation from monocyte to macrophage phenotype had occurred, changes in morphology and attachment to the culture vessel surface were examined for each replicate. Undifferentiated THP-1 monocyte-like cells grew in suspension, with most cells not attached to the bottom surface of the vessel. The THP-1 cells started to adhere around 6–8 h after addition of...
30 ng/ml of PMA. They then began to develop more macrophage-like morphology, with the most dramatic difference observed at 48–96 h. Their shape changed from being round to more variable and the cytoplasm contained more phagocytic vacuoles. At each time point there were a few cells where the morphology indicated they were more macrophage-like than the majority of cells. More than 99% of the cells were adherent and had similar morphology by the end of the time course at 96 h, indicating that this culture of THP-1 clone 5 had retained its high differentiation phenotype.

Prior to addition of PMA, the majority of the THP-1 cells were in G1 (growth) phase with approximately one third in S (DNA synthesis) phase (Figure 1A). After 10 h, the profile of DNA content detected by FACS was clearly changed. The number of cells in S phase was dramatically reduced and by 24 h there were <10% of cells with intermediate DNA content between 2N (G1 phase) and 4N (G2 phase) (Figures 1B,C). This low proportion was maintained for the rest of the time course (Figures 1D,E). The proportion of cells with 4N DNA content (G2) actually increased by 24 h as cells that had already entered S phase completed DNA synthesis. The proportion of cells in G2 phase gradually declined thereafter. These results indicate that differentiation of THP-1 cells to a macrophage morphology was preceded by a rapid decline in proliferation due to blocks at both entry to S phase and the well-documented G2-M phase (cytokinesis/cell division) checkpoint, most commonly associated with DNA damage (Stark and Taylor, 2004).

To ensure that the differentiation of THP-1 cells from promonocytic to macrophage-like phenotype was supported by changes in gene transcription, gene expression levels derived from qPCR were assessed for two relevant markers, MYB which encodes a transcription factor associated with proliferating hematopoietic cells (reviewed by Zhou and Ness, 2011) and monocyte differentiation antigen gene CD14 (Bazil et al., 1982; Wright et al., 1990). MYB was down-regulated during THP-1
differentiation in the earlier FANTOM4 study (Suzuki et al., 2009). In the present study, MYB expression was also found to decrease after PMA stimulation in all biological replicates, with the lowest level at around 2:30–3 h (Supplementary Figure S1). CD14, which was not detected in undifferentiated THP-1 clone 5 cells, consistent with FACS analysis of the parent THP-1 line (Bosshart and Heinzelmann, 2016), was not up-regulated until the last few points of the time course, from 24 h (Supplementary Figure S1).

Changes in Gene Expression During Differentiation of THP-1 Cells

The time course of THP-1 differentiation for all of the replicates is available on the ZENBU genome browser. The browser enables gene-specific search and includes the FANTOM5 promoter and enhancer tracks and the UCSC CpG island track. To compare the detailed time course with the earlier study (Suzuki et al., 2009) that had fewer time points, CAGE sequencing results were first analyzed for index genes highlighted as markers in that study. Figure 2A shows the time courses of representative macrophage differentiation marker genes. As with the qPCR results, CD14 increased late in the time course. Consistent with the previous data, CD14 was most strongly induced between 48 and 96 h. ICAM1 was induced maximally by around 24 h, ITGAM (encoding CD11B) increased after 12 h and APOE increased from around 18 h. By contrast to the earlier study, but in keeping with other results (Datta et al., 1992), the transcript encoding the growth factor receptor CSF1R was highly expressed by THP-1 cells without differentiation (200 TPM) but slowly induced a further 7-fold by PMA, reaching a maximum around 96 h. CSF1R is set as the default view on the ZENBU browser. The time course of CSF1R induction resembled the induction of CD36, proposed as a marker of differentiation in this system (Maess et al., 2010). CD36 mRNA was induced by 1 h and continued to increase throughout the time course. THP-1 cells respond to CSF1R ligand, CSF1 (Datta et al., 1992) and were previously shown to express CSF1 mRNA (Rambaldi et al., 1988). In the present study, CSF1 mRNA was induced with a very similar time course to CSF1R suggesting the likely involvement of an autocrine stimulatory loop in differentiation. CSF1 mRNA is also induced during the differentiation of primary human monocytes (Horiguchi et al., 1986) and indeed the highest level of expression in the FANTOM5 dataset is in monocyte-derived macrophages.

The transcriptional regulation of SPP1 (encoding osteopontin), representative of a cluster of genes induced between 12 and 24 h and associated with adherence, was dissected in detail previously (Suzuki et al., 2009). SPP1 was also massively induced by PMA in each time course replicate generated for this study. The timing of induction varied between replicates, so that the expression was highly variable between 6 and 24 h. Figure 2A shows mean data, averaged across replicates. The variation suggested that osteopontin is induced during the process of adhesion and that cluster analysis of the replicates independently might identify other genes that are involved in this key event.

A key feature of the PMA time course also dissected previously was the rapid down-regulation of genes for transcription factors involved in cell cycle regulation, notably MYB and E2F1, associated with growth arrest and induction of differentiation. Knockdown of some of these factors was sufficient to induce growth arrest and partial differentiation (Suzuki et al., 2009). Figure 2B shows a subset of these transcription factors. CAGE sequencing showed that MYB was expressed from two promoters, both of which were down-regulated rapidly in response to PMA. The related transcription factor gene, MYBL2 was highly expressed at the start of the time course (16 fold higher than the combined expression of the two MYB promoters) and declined later than MYB. MYBL2 was not assessed in the previous analysis. E2F1 was maximally repressed by around 12 h. GFI1 has been identified as a transcriptional repressor of target genes of the macrophage transcription factor, PU.1 (encoded by SPI1) (Wilson et al., 2010; Barth et al., 2019). In THP-1 cells, GFI1 was expressed from 4 separate promoters and rapidly down-regulated by PMA (Figure 2B). Consistent with repression of entry into the cell division cycle, the G1/S cyclin gene CCNE2 was rapidly repressed to undetectable levels (Figure 2C). Cell cycle-associated transcripts for genes associated with S phase and mitosis (Giotti et al., 2018) such as PCNA, BUB1, AURKA, CDK4 and the M phase cyclin, CCNB1, declined more slowly and progressively (Figure 2C), presumably as individual cells completed the cycle (Figure 1). Many of these transcripts showed some evidence of re-expression at 48–96 h (for example E2F1, CCNE2 and PCNA in Figure 2), which may reflect expansion of the small subset of cells that is resistant to PMA-induced differentiation even in this cloned line.

The time course of the loss of positive cell cycle regulators in response to PMA suggests that this may be a consequence rather than a cause of the block to S phase entry (Figure 1). The FANTOM 4 project did not analyze the expression of inducible cell cycle repressors. Others reported that the cell cycle inhibitor CDKN1A (p21WAF) was induced by PMA at the mRNA and protein level in THP-1 cells (Akashi et al., 1999; Traore et al., 2005). The induction of CDKN1A was associated with the loss of phosphorylated CDK2 protein, which was linked in turn to selective inhibition of phosphorylation of RB (retinoblastoma protein). Figure 2C shows that CDKN1A was induced from almost undetectable levels within 1 h and thereafter sustained at high levels throughout the time course.

These initial studies indicate that the current dataset broadly reproduces the temporal cascade of cell cycle arrest and monocyte-macrophage differentiation seen previously (Suzuki et al., 2009) and show that the increased temporal resolution of the present study was informative. We therefore undertook a refined network analysis of the data.

Patterns of Gene Expression Throughout Differentiation

CAGE sequencing data values (means for each replicate; N = 2–6) were loaded into the network analysis tool Graphia Professional to find patterns of gene activation and repression
FIGURE 2 | Gene expression during differentiation of THP-1 cells. X axis shows time points from 0 to 96 h after administration of PMA; Y axis shows average expression by CAGE analysis in tags per million (TPM). (A) Macrophage differentiation marker genes CD14, ICAM1, ITGAM, APOE, CSF1R, CD36, SPP1, CSF1. (B) Transcription factor genes MYB (two CTSS), MYBL2, E2F1, GFI1 (four CTSS). SPI1. Multiple CTSS associated with the same gene are shown by lighter colored sections of the bars. (C) Inducible cell cycle repressor genes CCNE2, PCNA, BUB1, AURKA, CDK4, CCNB1, CDKN1A. (D) Color code for time points on the X axis.
across the time course. To look for general trends across all replicates, a sample-to-sample network at 18 different time points was generated using a correlation coefficient of $\geq 0.95$. The sample-to-sample network (Figure 3A) highlights the global transcriptomic transition as the time after treatment increased. The transcriptome of samples at 0 h (deep red) were most different from those at 96 h (deep purple). Samples from other time points lie between these two, moving from red (1 h) through orange and yellow (2–6 h), green (7–12 h), and blue (14–48 h).

Next, CTSSs were correlated with each other based on the mean expression at 18 time points (to create a gene-to-gene network). The analysis was done with a correlation coefficient threshold of 0.85, which included 10,168 nodes out of 11,360 possible CTSS (see section “Materials and Methods”). Expression patterns of excluded CTSS were not correlated with any other CTSS at a correlation coefficient of 0.85. A network graph comprising one large element of 10,046 nodes and 53 smaller unconnected elements of between 2 and 11 nodes was generated using Graphia Professional. Markov clustering with an inflation value of 1.7 partitioned the graph into clusters of CTSS with related expression profiles. Figure 3B shows the result for the main element. There were 92 clusters with 10 or more nodes. A number of clusters showed distinct patterns of gene expression during differentiation. The largest of these are shown in Figure 3C. The clusters in which the key genes discussed above were found are shown in Table 1. A full list of all the clusters and the mean expression patterns of those with 10 or more nodes is presented in Supplementary Table S1.

The largest cluster, Cluster001, contained genes which code for proteins associated with mature macrophage functions, including surface markers (CD14, CD163, ITGAM), lysosome activity, transmembrane location and integrin complex. The most significantly enriched GO term was lysosome (enrichment score 14.8). This cluster contained genes with expression that steadily increased throughout differentiation, peaking at the end of the time course. Cluster002 declined progressively across the time course and showed high enrichment scores for terms associated with cell cycle (enrichment score 31.7) and mitochondria (enrichment score 21.1). Three hundred and sixty seven (17%) of the genes in this cluster were contained in an annotated list of 701 cell cycle genes (Giotti et al., 2018). These two clusters are consistent with the major phenotypic impacts of PMA, growth arrest and macrophage differentiation.

Aside from these two clusters, the network analysis revealed that many transcripts were induced or repressed transiently. By 1 h after PMA treatment, early response genes with functions such as transcriptional activation were highly expressed (Cluster020 peaking at 2 h then declining rapidly and Cluster007 peaking at 4 h). Both these clusters were associated with GO terms relating to regulation of transcription (enrichment scores 2.7 and 3.9, respectively). Expression of the genes in Cluster003 began to rise after 1 h and peaked between 5 and 8 h. They were associated with both the cell cycle (enrichment score 4.9) and cell-cell adhesion (enrichment score 4.1). Expression of genes in Cluster004 slowly increased from the 2 h time point, peaking between 10 and 18 h, and this cluster was enriched for GO terms relating to lysosome (enrichment score 5.3). Images of the average expression profiles are shown in Figure 3C.

Cell Cycle Gene Expression Throughout Differentiation
Cancer is characterized by abnormalities in cell proliferation, particularly the escape of the cancer cells from the normal controls on cell division (Sachs, 1982). In the analysis of the whole dataset, the second largest cluster (Cluster002) contained many genes associated with cell proliferation and the averaged expression profile of this cluster showed a decline during the time course with a small increase at 96 h. Note that by contrast to earlier studies (Nguyen et al., 2011; Garrison et al., 2017; Germano et al., 2017; Prange et al., 2017) (see section “Introduction”) the set of down-regulated transcripts did not include KMT2A (MLL), DOT1L, RUNX1 or ZNF521, implicated in primary leukemogenic transformation in AML. Each was expressed constitutively by THP-1 cells and unchanged upon differentiation. The cell cycle-associated cluster was explored in more detail by further clustering the expression profiles of the 2,158 genes separately, at higher stringency. A sample-to-sample analysis ($r \geq 0.9$) showed that for this group of genes, the later time points were quite distinct, but the earlier time points (from 0 to 14 h) were relatively similar with a weak correlation between time point and position in the network (Figure 4A). The gene-to-gene analysis was run at a correlation coefficient threshold of 0.95, which included 1540 nodes connected by 17,446 edges. The network was then clustered with an MCL inflation value of 1.7. 618 nodes of the original Cluster002 were excluded at this threshold. We hypothesized that genes involved in different phases of the cell division cycle, or with different mRNA stability, might decay with distinct temporal profiles Consistent with that view, the annotated cell cycle genes appeared in specific subclusters with distinct profiles of decay with time (Figure 4B). For example, Subcluster002 with 99 nodes had none of the reported cell cycle genes (Giotti et al., 2018), a highly significant departure from expected ($P = 7 \times 10^{-8}$). Overall expression in this cluster dropped very early in the time course. This cluster contains monocyte-associated transcription factors (IRF8, GFI1). These factors, as well as MYB and MYBL2 (which are also in the larger Cluster002 but did not form a part of a subcluster) have myeloid-specific roles in proliferation and down-regulation using siRNA can drive growth inhibition (Suzuki et al., 2009). Subcluster003 and Subcluster004 had a significant excess of cell cycle genes ($P < 1 \times 10^{-5}$ and $P = 3 \times 10^{-5}$ respectively). For these clusters, average expression declined slowly after 10 h and both had a small rise at 96 h, not seen in Subcluster002. Both these clusters had equal numbers of genes associated with G2, S phase and G2/M phase of the cell cycle. In contrast Subcluster010 with 17 nodes had 10 G2/M genes and no S phase genes ($P = 5 \times 10^{-6}$). Average expression was relatively flat until 18 h when it dropped rapidly. Using the Bonferroni method to correct for multiple testing, a $P$-value of 0.002 would be equivalent to the accepted threshold for significance of 0.05, so these differences are highly significant. The full list of subclusters and averaged expression profiles are provided in Supplementary Table S2.
FIGURE 3 | Network analysis of patterns of gene expression throughout differentiation of THP-1 cells. (A) Sample-to-sample analysis. Each node (sphere) represents the average of replicates at a time point, colored as shown in Figure 2D, indicating hours after addition of PMA. Edges between nodes show correlations of ≥0.95. (B) Gene-to-gene analysis showing the main element of the full network. Nodes represent genes (TSS) and the edges between them represent correlations of ≥0.85. MCL clustering was performed at an inflation value of 1.7 and nodes are colored according to their membership in a cluster. (C) Clusters of nodes that are differentially expressed across the time course. Orientation is the same as (B) but edges and other clusters have been removed for ease of visualization. Color of nodes shows the cluster to which nodes were allocated; histograms (colored the same as the nodes in the cluster) show the average expression patterns of the genes within the clusters. Color code for the X axis is as for Figure 2.
TABLE 1 | Clusters associated with key genes from network analysis.

<table>
<thead>
<tr>
<th>CTSS ID</th>
<th>Cluster number</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD14</td>
<td>001</td>
</tr>
<tr>
<td>ICAM1</td>
<td>001</td>
</tr>
<tr>
<td>ITGAM</td>
<td>001</td>
</tr>
<tr>
<td>APOE</td>
<td>001</td>
</tr>
<tr>
<td>CSF1R</td>
<td>001</td>
</tr>
<tr>
<td>CD36</td>
<td>001</td>
</tr>
<tr>
<td>SPP1</td>
<td>001</td>
</tr>
<tr>
<td>MYB p2</td>
<td>002</td>
</tr>
<tr>
<td>MYBL2</td>
<td>002</td>
</tr>
<tr>
<td>E2F1</td>
<td>002</td>
</tr>
<tr>
<td>GFI1 p1</td>
<td>002</td>
</tr>
<tr>
<td>GFI1 p2</td>
<td>002</td>
</tr>
<tr>
<td>GFI1 p3</td>
<td>002</td>
</tr>
<tr>
<td>GFI1 p4</td>
<td>002</td>
</tr>
<tr>
<td>CCNE2</td>
<td>002</td>
</tr>
<tr>
<td>PCNA</td>
<td>002</td>
</tr>
<tr>
<td>BUB1</td>
<td>002</td>
</tr>
<tr>
<td>CDK4</td>
<td>006</td>
</tr>
<tr>
<td>CDKN1A</td>
<td>012</td>
</tr>
<tr>
<td>SPI1</td>
<td>014</td>
</tr>
<tr>
<td>AURKA</td>
<td>273</td>
</tr>
<tr>
<td>CCNB1</td>
<td>273</td>
</tr>
<tr>
<td>MYB p1</td>
<td>332</td>
</tr>
</tbody>
</table>

The analysis was based on averaged values over the whole dataset. Where present, p1 – pn indicate the detection of additional CTSS attributed to the same gene by the CAGE analysis.

A previous study (Giotti et al., 2018) identified 701 genes that were expressed/induced in multiple cell types specifically in S phase and G2/M phases of the cell cycle. Of these, 561 (80%) were expressed in THP-1 cells, represented by 617 promoters. These 617 promoters were selected from the filtered dataset of mean values and analyzed separately. When clustered at these 617 promoters were expressed in THP-1 cells, represented by 617 promoters. These 617 promoters were selected from the filtered dataset of mean values and analyzed separately. When clustered at $R \geq 0.75$, 599 promoters were included in the analysis and 512 of these fell into the largest cluster (named CC-cluster01) with an average expression pattern which declined slowly until 10 h and then more steeply to 48 h, after which there was a small increase in average expression (Figure 4C). This group included the classic proliferation markers mentioned earlier. This finding provides further support for the annotation of these genes as cell cycle-related. A second cluster of 21 nodes (CC-cluster02) showed a small decline in expression from 0 h, with a large increase at 96 h. Other smaller clusters containing 2 – 11 unique genes showed similar expression patterns to the whole dataset (Figure 4C). The full CC-cluster lists and averaged expression profiles are provided in Supplementary Table S3. The set of 150 cell-cycle genes for which promoter activity was not detected in THP-1 cells is provided in Supplementary Table S4. In most cases, the apparent lack of expression is an issue of promoter annotation, and there is, in fact, expression of promoter activity in the region of the corresponding gene evident in the FANTOM5 dataset. For example, our data, and the FANTOM5 data, reveal that the transcript encoding the key regulator CDC25B, which is essential for G2/M cell cycle progression in AML (Didier et al., 2008) is expressed in THP-1 cells but is primarily driven by a cluster of three promoters that are internal to the annotated TSS in RefSeqs NM_001287516, NM_001287517, and NM_001287518. This can be seen in the data available on the ZENBU browser.

In summary the analysis of known cell cycle genes is consistent with the shut-down of entry into S phase prior to the 10 h time point as indicated by the cell cycle analysis (Figure 1) and subsequent slow decay of S phase-associated transcripts as cells complete the DNA synthesis phase of the cell cycle and are blocked in G2/M.

Transcription Factor Gene Expression Throughout Differentiation

In cells undergoing transitions in state, transcription factor gene expression precedes the expression of the target genes being regulated (Arner et al., 2015). Of 1639 transcription factors curated recently (Lambert et al., 2018), 647 (39%) were expressed in the filtered THP-1 dataset. The expression profiles of these genes were used for further analysis. In a sample-to-sample analysis including all time points ($R \geq 0.61$), the 0 h time point was an outlier, linking only with the 3 h time point (Figure 5A). A gene-to-gene analysis of the 647 transcription factor genes (796 promoters) was performed at a relatively stringent correlation coefficient of $R \geq 0.85$, which included 540 nodes (transcription factor genes) and 3026 edges (connections between them at $R \geq 0.85$). Clustering was performed at an inflation value of 1.7 and clusters were named as TF-cluster01 etc. The clusters are shown in Supplementary Table S5 which also contains histograms of mean activity of each cluster. The much denser time course analyzed herein, compared to the FANTOM4 project (Suzuki et al., 2009) highlights a temporal cascade in which the large majority of the transcription factor promoters exhibit regulated expression. Broadly speaking, there is a set of clusters where the transcripts are progressively down-regulated with different dynamics, a reciprocal set progressively up-regulated and a third set in which expression is regulated transiently (Figure 5B).

The transiently regulated transcription factors include the so-called immediate early genes (IEG) that have been analyzed in detail in other differentiation/activation time courses by the FANTOM 5 consortium (Vacca et al., 2018). That analysis identified a set of 42 protein-coding genes with promoters that consistently drive a rapid, but transient, wave of transcription in response to stimulus. A subset of these IEG was expressed by differentiating THP-1 cells. TF-Cluster15 contains transcription factors EGR1, EGR3, EGR4, JUNB, KLF2, KLF10 and MEF2D which were fully induced within 1 h and thereafter declined slowly. Other IEG had distinct patterns. EGR2, which has specific functions in myeloid differentiation including regulation of CSF1R transcription (Laslo et al., 2006; Krysinska et al., 2007), is part of TF-cluster10 (also including ELK3, FOXP2, JUNB, TBX3, and ZEB2) that peaks later around 8–10 h, around the time of the first appearance of adherent cells. ZEB2 was shown recently to be required for the establishment and maintenance of tissue macrophage identity (Scott et al., 2018). By contrast to other systems in which it is an IEG, and consistent with its known...
FIGURE 4 | Network analysis of cell cycle genes. (A) Sample-to-sample analysis of Cluster002, containing genes associated with the cell cycle and proliferation. Nodes show averaged values for each time point and edges show correlations of $\geq 0.90$ between nodes. Hours after addition of PMA are indicated. (B) Expression profiles of subclusters generated by gene-to-gene analysis of Cluster002. Averaged values for each time point. Correlation coefficient was $\geq 0.95$ and MCL inflation value was 1.7. (C) Gene-to-gene analysis of 561 curated cell cycle genes (617 TSS) (Giotti et al., 2018) expressed during THP-1 differentiation. Nodes show averaged values for each gene and edges show correlations of $\geq 0.75$ between them. MCL inflation value was 1.7. Histograms show the averaged expression profile for each cluster. Color code for the X axis is as for Figure 2.
FIGURE 5 | Expression of transcription factors during THP-1 differentiation. (A) Sample-to-sample analysis of 647 curated transcription factors (796 TSS) (Lambert et al., 2018) expressed during THP-1 differentiation. Edges show correlations of $\geq 0.61$. Hours after addition of PMA are indicated. (B) Expression profiles of clusters generated by gene-to-gene analysis of curated transcription factors. Averaged values for each time point. Correlation coefficient was of $\geq 0.85$ and MCL inflation value was 1.7. (C) Network analysis of transcription factor motif activity. Histograms showing average activity (as detected by MARA) of transcription factor motifs, clustered at a correlation coefficient of $\geq 0.60$ and MCL inflation value of 1.7. Color code for the X axis is as for Figure 2.
functions in macrophage differentiation (Shen et al., 2016), JUN was not induced rapidly but instead formed part of a progressive differentiation cluster (see below).

Mean expression of transcription factor genes in the largest cluster (TF-Cluster01) was maximal in untreated THP-1 cells and declined rapidly in response to PMA. This down-regulated cluster contains multiple transcripts associated with immature myeloid cells and maintenance of the proliferative state, including MYB, GFH1 (both shown in Figure 2B), IRF8 and RXRA. Average expression of transcripts within TF-Cluster03 declined more slowly; they include the core regulators of cell cycle gene expression, E2F1 (Figure 2B) and FOXM1 (Giotti et al., 2018). Several TF clusters show progressive up-regulation with distinct profiles. TF-Cluster04 (including JUN, MAF, NR1H3) increased across the whole time course, TF-Cluster11 (including ATF3, TCF7L2) was induced earlier, and TF-Cluster02 (including IRF1, FOXP1) was induced late, associated with terminal differentiation. One highly induced transcription factor with a unique expression profile that did not form part of a cluster was MAFB, discussed further below.

MARA Analysis
Promoters contain motifs that are recognized by DNA binding proteins. Where promoters share such motifs it is likely that they are regulated by the same proteins, and expression correlates with the number of copies of the relevant motif in the promoter region. The analysis tool MARA (Suzuki et al., 2009), which calculates motif activity based on this correlation, was used to analyze the response of THP-1 cells to PMA. The results are shown in Supplementary Table S6 and histograms of activity of selected motifs across the time course are shown in Supplementary Figure S2. A preliminary clustering analysis ($r \geq 0.6$, MCL inflation 1.7) showed four patterns of activity: declining or increasing throughout the time course, transiently up-regulated and transiently down-regulated (Figure 5C). These patterns confirm the conclusions of the earlier work (Suzuki et al., 2009). The imputed transcriptional activity of MYB and E2F1.5 motifs, and of TFDP1, which binds cooperatively with E2F (Rubin et al., 2005), was down-regulated progressively over the time course. The activity of related motifs recognized by AP1 family members (FOS; FOSB, JUN; D; FOSL2; JUN) increased rapidly, as did the activity of the NFE2L2 and TGF1 motifs (both factors induced by PMA).

There was some evidence of increased activity of the IRF1.2,7 motif, consistent with the induction of interferon target genes in Cluster032. Kubosaki et al. (2010) reported that the same genes were expressed constitutively in THP-1 cells and were targets of IRF8. Consistent with this and other studies (Phanstiel et al., 2017), IRF8 was highly expressed by THP-1 cells (the most abundant transcription factor mRNA) and repressed by PMA, but the putative target genes were low in undifferentiated cells and strongly induced. PMA also initiated clear and transient increased activity of the EGR1.3 motif. Also consistent with FANTOM4 data there was a decrease in activity of the NFYA[A,B,C] and octamer (POU2F1.3) motifs, although NFYB and NFYC mRNAs (both highly expressed in THP-1 cells) were not regulated by PMA.

The greater resolution of the current dataset permitted the detection of regulated activity of additional motifs, notably the NR4A2 (also recognized by NR4A1) and TFEB motifs. Both NR4A1 and NR4A2 were amongst immediate early genes induced by PMA. The TFEB motif, recognized by members of the microphthalmia transcription factor family, is enriched in the promoters of lysosome-associated genes (Sardiello et al., 2009; Hume et al., 2010). Two members of this family, TFEC and TFE3, were constitutively expressed in THP-1 cells but two others, TFEF and MITF, were progressively induced in response to PMA in parallel with lysosomal hydrolases (e.g., ACP2, CTSD, LIPA) and components of the vacuolar ATPase (e.g., ATP6V0A1). Conversely, we did not reproduce evidence of inducible activity of SPI1 (PU.1), RUNX1.3 or SNAI1.3 motifs although transcripts encoding the related transcription factors TWIST1 and SNAI1 were induced by PMA.

Enhancer Activation During THP-1 Differentiation
The increased depth of sequencing available in this dataset also enables for the first time the detection of enhancer RNA (eRNA), which derives from the transcriptional activity of regulated enhancers, and inferred relationships with inducible promoters. The eRNAs are rapidly degraded by the exosome complex (Andersson et al., 2014) and accordingly, they are generally detected at <1 TPM. Detection is therefore subject to stochastic noise with median mapped reads of around 10 million per library. Nevertheless, integration of the entire dataset enabled detection of regulated bidirectional transcription associated with enhancers annotated in the FANTOM5 dataset and identified additional candidate enhancers. The identified enhancers were compared to those identified in FANTOM 5. 1353 of the less stringent set and 656 of the more stringent set were found in the full set of ~63,000 enhancers (Andersson et al., 2014), indicating that this analysis has discovered some potential novel enhancers. Both sets were compared to the set of ATAC-seq peaks identified in analysis of chromatin architecture of differentiation (Phanstiel et al., 2017). Overlap was detected for 88 and 79% enhancers in the more and less stringent sets, respectively. One novel example highlighted in the chromatin analysis is the complex BCL6/TPRG1 locus. BCL6 is implicated in cell cycle regulation and interacts with TP53 and CDKN1A (Phan et al., 2005), whereas there is little known of the biology of TPRG1. The promoters of these two transcripts are separated by >200 kb (see ZENBU Browser). The TSS of TPRG1 is located upstream of the annotated RefSeq transcript NM_198485 at Chr3:188947237, downstream of a TATA box. The differentiation time course data confirm the rapid (within 1 h) and synchronous continued induction of both transcripts, and the detectable induction of enhancers with the intervening LPP locus. Figure 6 shows the profile of the activity of each of these promoters/enhancers across the time course. A similar co-regulated pattern was evident at the MITF locus. The induction of MITF with time in response to PMA was correlated with induction of the adjacent FRMD4B gene and evidence of bidirectional transcription initiation in the intervening genomic region.
To further confirm the validity of the enhancers found, they were compared to every individual FANTOM5 enhancer set. The THP-1 FANTOM5 library was the top hit for both sets, and other myeloid libraries were overrepresented in the top of the list. 219 out of 1172 FANTOM5 THP-1 enhancers were confirmed in the more stringent time course set. The results are available in Supplementary Tables S7A,B.

The macrophage-specific transcription factor, PU.1 (encoded by SPI1) is required for differentiation and controls the expression of many macrophage-enriched genes in THP-1 cells (Suzuki et al., 2009). SPI1 was expressed constitutively by THP-1 cells and was not highly regulated by PMA. As noted above, MARA did not reveal regulated activity of the PU.1/SPI1 motif across the time course. The expression of SPI1 in THP-1 cells depends upon an enhancer around 16 kb upstream of the TSS that binds the transcription factor SATB1 (Steidl et al., 2007). The CAGE data (see ZENBU Browser) reveal that there is abundant unidirectional transcription initiation from this enhancer in THP-1 cells, likely driving an annotated IncRNA, that is marginally up-regulated by PMA. SATB1 mRNA, on the other hand, was progressively down-regulated in response to PMA.

Amongst other genes of interest, the MYB gene contains two upstream and two intronic enhancers. A candidate enhancer in the first intron was progressively down-regulated in response to PMA. This same region, conserved across species, has two enhancers including the highly conserved FMS intronic regulatory element (FIRE) (Rojo et al., 2017). Finally, the FANTOM5 data identified candidate enhancers between the ITGAM (encoding CD11B) and ITGAX (CD11C) loci, both genes induced late in differentiation by PMA, and the CAGE data detect bidirectional transcription in the intergenic region (Supplementary Figure S3).

**DISCUSSION**

This study used CAGE to analyze the transcriptional regulatory networks that are initiated in response to phorbol esters in a high differentiation clone of the THP-1 myeloid leukemia line. The analysis of the cell cycle arrest is consistent with earlier reports. Using the parent line, Traore et al. (2005) found a similar proportion of cells in S phase in undifferentiated cells and reported a complete inhibition of DNA synthesis (assessed by BrdU uptake) in response to PMA within 24 h. They therefore implicated a block to G1/S entry which is also evident from the rapid loss of intermediate (S phase) DNA content in our study (Figure 1). The persistence of cells with 4N DNA content in both their study and ours indicates at least some of the cells are then blocked at the G2/M checkpoint, as also seen in AML cells treated with genotoxic agents (Didier et al., 2008).

Compared to the earlier analysis by the FANTOM4 consortium (Suzuki et al., 2009) we have analyzed a much denser time course, sequenced at greater depth using next generation sequencing platforms, and taken advantage of the denser time course, sequenced at greater depth using next generation sequencing platforms, and taken advantage of the depth of data to perform a network-based analysis of the temporal cascade. Minor differences between our study and FANTOM4 in detection of specific genes (e.g., TP53, CSF1R) are likely due to differences in sequencing technology/depth/error rates/mapping and refinement of the human genome assembly in two updates since the previous study (which was mapped to hg18, NCBI Build 36.1, released in 2006). Where there are differences, our data are validated by multiple replicates at multiple time points and the precise location of the TSS. The FANTOM5 consortium generated similar data for a number of differentiation/activation cascades in other cell types (Arner et al., 2015). Those data enabled the identification of eRNA derived from active enhancers and provided evidence that transient activation of eRNA transcription preceded the activation of target genes. The data generated for THP-1 cells complement and extend the detailed chromatin analysis published by others (Phanstiel et al., 2017), which compared only the unstimulated and fully differentiated states (after 72 h) and used the parent
FIGURE 7 | Expression of MYB intron 9 putative enhancer during THP-1 differentiation. (A) Screenshot of Zenbu browser showing MYB intron 9 putative enhancer (circled in red). (B) Expression of two MYB promoters and the putative intron 9 enhancer. Y axis shows expression levels in TPM; negative value (gray bars in panel on right) indicates expression from the reverse strand. X axis shows the time points (hours after administration of PMA; color code as shown in Figure 2).
THP-1 line in which only a subset of cells responds to PMA. They emphasized the role of AP1 transcription factors in the formation of chromatin loops between distal enhancers and macrophage-specific promoters, a conclusion that is supported by the MARA analysis herein.

One of the cellular systems analyzed in FANTOM5, in which there was a similar extensive change in gene expression, was the activation of human monocye-derived macrophages by bacterial lipopolysaccharide (LPS) (Baillie et al., 2017). Network-based analysis of that response identified waves of transiently induced promoters/transcripts extending across the entire time course up to 72 h. Each cluster of LPS-induced or repressed transcripts contained both candidate feedback repressors of the previous transcriptional cluster and transcription factors and autocrine growth factors that contributed to the regulation of subsequent clusters. The same pattern of transient waves of inducible transcripts is evident in this PMA time course in the high differentiation THP-1 cell line. For example, in the network analysis (Supplementary Table S1), Cluster020 contains immediate early transcription factors as well as feedback regulators such as ZFP36, which encodes an enzyme that degrades unstable AU-rich mRNAs. The related gene, ZFP36L1, was even more highly induced by PMA, peaking after 2–4 h and remaining elevated. Over-expression of ZFP36L1 was shown to positively-regulate differentiation in THP-1 cells. This was attributed to degradation of CDK6 mRNA (Chen et al., 2015), but this transcript was actually induced by PMA.

Cell cycle arrest in THP-1 cells probably involves multiple mechanisms. THP-1 cells were reported to lack functional TP53 and indeed TP53 mRNA was low in THP-1 cells in the FANTOM 4 and FANTOM5 data (Suzuki et al., 2009; Forrest et al., 2014). In the current study, TP53 mRNA was readily detectable in multiple replicates and formed part of Cluster001. Consistent with functional expression of TP53, PMA was previously shown to induce the TP53-responsive cell cycle regulator p21WAF1/CIP1 (encoded by CDKN1A). Induction of both CDKN1A mRNA and protein in published studies was detected after 24 hrs of PMA treatment (Traore et al., 2005). Our study shows that induction of CDKN1A by PMA is a much earlier event and provides insights into the mechanism. Transcriptional activation of CDKN1A in these cells has been linked to GC-rich binding sites for SP1 in the proximal promoter (Traore et al., 2005; Pivorunias et al., 2007). CDKN1A has also been implicated in cell cycle arrest in response to PMA in U937 myeloid leukemia cells (Vrana et al., 1998). CDKN1A formed part of Cluster012, and was already strongly induced 1 h after PMA addition, well in advance of cell cycle blockade and consistent with the existence of a poised initiation complex at the CDKN1A promoter (Espinosa et al., 2003). The FANTOM5 data reveal the presence of regulated enhancers both upstream and downstream of the CDKN1A promoter that were also implicated in other human cell types (Espinosa et al., 2003). SGK1 is another TP53 target gene (You et al., 2004) that was also immediately and massively induced by PMA. Meta-analysis of many large datasets identified only a limited set of consensus TP53-dependent genes (Fischer, 2017). Amongst the most highly validated targets, GDF15, PLK2, PLK3, BTG2, SESN1, and SESN2 (each of which can contribute to cell cycle arrest; Fischer, 2017; Yuniati et al., 2019) were also induced rapidly during the THP-1 time course of differentiation presented here and most likely also controlled by poised promoters, whereas BAX, GADD45A, MDM2, RRMM2, and SUS6 were constitutively expressed.

Accordingly, it is unlikely that CDKN1A induction provides the sole mechanism of cell cycle inhibition in PMA-stimulated THP-1 cells. The expression of retinoblastoma (RB1), itself a tumor suppressor gene (Dyson, 2016), and associated with G2/M cell cycle arrest (Stark and Taylor, 2004) was rapidly induced from two separate promoters by PMA. The set of inducible genes in Cluster012 also contains AB12 which suppresses FLT3-induced proliferation (Kazi et al., 2017) and the tumor suppressor gene DOCK4 (Sundaravel et al., 2015). Cluster003, which peaks transiently around 6 h, contains transcripts encoding cycle-dependent kinases encoded by CDK9 and CDK20 which have complex functions in cell cycle regulation (reviewed in Malumbres, 2014). By association, proteins encoded by other transcripts within these clusters could also contribute to cell cycle arrest. Amongst the most highly induced transcripts in Cluster001 are candidate regulators of proliferation including the MAP kinase inhibitor DUSP1 and GADD45G, which can drive growth arrest and differentiation in hematopoietic stem cells (Thalheimer et al., 2014).

The first detected response to PMA, as in other cellular state transitions, was the induction of immediate early genes. In each case, the mean gene expression increased to a peak within 1–2 h; the patterns differed depending upon how rapidly the expression decayed. As noted in the results, many IEG encode transcription factors. One key immediate early gene that is specific to myeloid lineages is MAFB. MAFB can drive monocyte commitment when transduced into progenitor cells (Gemelli et al., 2006). In the FANTOM 5 dataset, MAFB mRNA was most highly expressed in monocytes and macrophages and induced in the transition from committed granulocyte-macrophage progenitors (Joshi et al., 2015). In the current study, MAFB was an IEG, maximally up-regulated after 2 h and sustained thereafter. MAFB mRNA and protein were reportedly also up-regulated in THP-1 cells in response to a different differentiation signal, ATRA, in combination with tumor necrosis factor (Zhang et al., 2012). Given the rapidity of induction in our study, and the fact that the MAFB locus is in open chromatin in THP-1 cells (Suzuki et al., 2009; Phanstiel et al., 2017), transcriptional activation of MAFB most likely involves a poised initiation complex. Zhang et al. (2012) predicted target genes for MAFB based upon the presence of MAF-response elements (MARE) in their promoters. The MARE contains a consensus AP1 element and is partly captured in our MARA analysis. The majority of the 64 putative MAFB target genes (including AIM2, CCL2, LPXN, PRDM1, SLC15A3, SPOCK1) were also progressively induced in our study. However, although knockdown of MAFB compromised expression of target genes such as PRDM1 and SPOCK1, it did not prevent differentiation (Zhang et al., 2012).

Several other proteins encoded by IEG, notably JUN and EGR2, also have specific functions in macrophage differentiation (Rojo et al., 2017). NR4A1 is required specifically for the maturation of blood monocytes to form the non-classical (in mice, Ly6C+, in humans CD16+ subset (Thomas et al., 2016).
Deletion of part of an enhancer in the mouse Nr4a1 locus selectively ablated expression in monocytes (Thomas et al., 2016). The FANTOM5 data reveal multiple upstream enhancers within Nr4a1 and the present THP-1 time course reveals at least two regions of inducible bidirectional promoter activity.

Whereas there are numerous DNA-modifying transcription factors up- and down-regulated across the time course of differentiation, few regulated transcripts encoding chromatin modifying enzymes were identified that could contribute to the large-scale chromatin looping reported by Phanstiel et al. (2017). The exception is KDM6B (also known as JMJD3), which was almost undetectable in unstimulated cells and rapidly induced alongside IEG transcription factors. In mice, KDM6B is required for normal macrophage differentiation (Satoh et al., 2010). Consistent with the induction during monocytic differentiation in THP-1 cells, KDM6B was highly expressed in human monocytes and induced further with adhesion (Forrest et al., 2014). KDM6B is also recruited to TP53-dependent promoters in many other cell types, so likely contributes also to growth arrest (Williams et al., 2014).

The large set of transcripts within Cluster001 that was progressively up-regulated across the time course of THP-1 differentiation provides a rich resource for the identification of novel genes that are involved in cell cycle arrest on the one hand and macrophage differentiation on the other. Their inferred function is implied by stringent coregulation with many genes of known function including macrophage differentiation markers such as CD14, CD163, CSF1R, ITGAM, and MERTK and by cross-comparison with primary cells [stem cells, committed monocytes and induced further with adhesion (Forrest et al., 2014). KDM6B is also recruited to TP53-dependent promoters in many other cell types, so likely contributes also to growth arrest (Williams et al., 2014).

The large set of transcripts within Cluster001 that was progressively up-regulated across the time course of THP-1 differentiation provides a rich resource for the identification of novel genes that are involved in cell cycle arrest on the one hand and macrophage differentiation on the other. Their inferred function is implied by stringent coregulation with many genes of known function including macrophage differentiation markers such as CD14, CD163, CSF1R, ITGAM, and MERTK and by cross-comparison with primary cells [stem cells, committed monocytes and induced further with adhesion (Forrest et al., 2014). KDM6B is also recruited to TP53-dependent promoters in many other cell types, so likely contributes also to growth arrest (Williams et al., 2014).

Conversely, in parallel with the loss of cell cycle-associated transcription, the set of transcripts that is down-regulated progressively in response to PMA includes genes that are specific to monocytes and repressed upon macrophage differentiation. One such gene is IRF8. IRF8 mutations in humans are associated with monocye deficiency (Crosslin et al., 2013). Other monocye-specific transcripts down-regulated by PMA include the chemokine receptor, CCR2, the receptor for granulocyte colony-stimulating factor (CSF3R), the tetraspanin MS4A3 (recently identified as a marker of monocytic progenitors; Liu et al., 2019), P selectin ligand (SELPGL), surface markers (e.g., FCGR2A, CD38, CD302, CD320, GPR160) and S100A8/S100A9, encoding the cytoplasmic calgranulin complex. MS4A3 is expressed by committed human myeloid progenitors and down-regulated in monocytes (Forrest et al., 2014). CSF3R, CCR2, CD302, and S100A8/S100A9 are amongst the large set of genes...
down-regulated during differentiation of macrophages from blood monocytes (Bailie et al., 2017).

CONCLUSION

This study describes a detailed time course of growth arrest and differentiation of the AML line, THP-1. Whereas these processes are commonly portrayed as linear sequences with a beginning and an end point, the time course reveals a sequential cascade of transient changes in gene expression. Even at 96 h post-stimulation, there are macrophage-associated genes that appear to be increasing their expression (such as CD14), most likely still responding to signals from CSF1R and adhesion to the substratum. In mixed population of monocyte-macrophages in tissues, differentiating asynchronously, this sequential cascade might be (mis)interpreted as evidence of the existence of distinct monocyte subpopulations. Hundreds of signaling molecules, transcription factors and feedback regulators are involved. The analysis reveals many novel inducible genes (e.g., TPRG1, FRMD4B) for which there is currently no functional annotation. THP-1 has been studied as an archetypé of the subset of AML that can respond to differentiation therapy (Tagliafico et al., 2006). A detailed understanding of the process of differentiation and growth arrest may highlight new therapeutic targets and opportunities.

DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found at http://fantom.gsc.riken.jp/zenbu/glyphs/#config=Gazova_THP1_diff entiation. The raw data files have been uploaded to the European Nucleotide Archive, BioProject ID PRJEB38762.

AUTHOR CONTRIBUTIONS

IG performed the cell culture, CAGE analysis and validation, and wrote the first draft of the manuscript. LL was involved in CAGE library preparation. SB and SC helped with the bioinformatic analysis of CAGE sequencing results. EA performed the MARA analysis. MH and JS created the visualization platform and helped prepare the data for visualization. LD and RA received funding from the Danish Council for Independent Research (Grant 6108-00038) and the European Research Council (ERC) under the European Union’s Horizon 2020 Research and Innovation Program (Grant 638173).

REFERENCES


in transitioning mammalian cells. Science 347, 1010–1014. doi: 10.1126/science.1259418


**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Gažová, Lefevre, Bush, Clohisey, Arner, de Hoon, Severin, van Duin, Andersson, Lengeling, Hume and Summers. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.