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Kinetic $[^{18}\text{F}]$-Fluoride of the Knee in Normal Volunteers

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**ORIGINAL ARTICLE**

Purpose: $[^{18}\text{F}]$-sodium fluoride ($[^{18}\text{F}]$NaF) is a well-established bone-seeking agent that has shown promise to assess bone turnover in a variety of disorders, but its distribution in healthy knee joints has not been explored. This study aimed to investigate parametric values for $[^{18}\text{F}]$NaF uptake in various bone tissues of the knee and their spatial distributions.

Methods: Twelve healthy subjects were hand-injected with 92.5 MBq of $[^{18}\text{F}]$NaF and scanned on a 3-T PET/MRI system. Listmode PET data for both knees were acquired for 50 minutes from injection simultaneously with MRI Dixon and angiography data. The image-derived input function was performed to obtain $K_i$ ($K_i^c$) values and nonlinear regression analysis to obtain $K_i^\text{NL}$, $K_i$, $k_1(k_2 + k_3)$, and blood volume. Comparisons for the measured kinetic parameters, SUV, and SUVmax were made between tissue types (subchondral, cortical, and trabecular bone) and between regional subsections of subchondral bone.

Results: Cortical bone had the highest $[^{18}\text{F}]$NaF uptake differing significantly in all measured parameters when compared with trabecular bone and significantly higher SUVmax and $K_i$ than subchondral bone. Subchondral bone also had significantly higher SUV, SUVmax, and $K_i$ than trabecular bone tissue. Regional differences were observed in $K_i$ and $k_1(k_2 + k_3)$ values.

Conclusions: Quantitative $[^{18}\text{F}]$NaF PET is sensitive to variations in bone vascularization and metabolism in the knee joint.

Key Words: bone, fluoride, hybrid imaging, kinetics, knee, MRI, NaF, PET, PET/MRI

The coils were used because of their lower at-
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128, slice 18 is irreversibly bound to
2 minutes with the same
plasma concentration to the extravascular
the voxels centered in the middle of
=0 ). Data from 10 to 50 minutes were reconstructed using time of flight
2 seconds and 24
1s e c o n d s ,1 3 and diffuses back
to the blood compartment at a rate defined by k3. The
eventual rate of dissociation of the fluoride from the bone
matrix is described by k4.

\[^{18}\text{F}]\text{NaF}. The study was performed in compliance with the local in-
stitutional review board (Stanford University, Administrative Panels
for the Protection of Human Subjects) regulations, and all subjects
provided written consent prior to the study. One subject repeated
the trial on 3 separate days to evaluate reproducibility.

**PET/MRI Scanning**

Subjects were scanned on a 3-T whole-body time-of-
flight PET-MR hybrid system (GE Healthcare, Milwaukee, Wis).
Each subject was positioned feet first with a 16-channel
flexible phased-array receive-only coil (NeoCoil, Pewaukee, Wis)
around each knee. The coils were used because of their lower at-
etenuation effect on PET photons. Both knees were scanned using 1
PET bed (field of view = 25 cm) in list mode starting with the injec-
tion of NaF for 50 minutes, and all MRI data were acquired simul-
taneously with PET imaging.

Magnetic resonance angiography data were acquired using a
3-dimensional gradient echo sequence with imaging parameters:
repetition time/echo time (TE) = 21/2.1 milliseconds, slices = 18,
slice thickness = 1.2 mm, and flip angle = 15°. A 2-point Dixon
fat-water T1-weighted spoiled gradient echo MR sequence was
acquired for MR-based attenuation correction of PET data26 with
acquisition parameters: repetition time/TE1/TE2 = 4.1/1.1/2.2 milli-
seconds, field of view = 50 × 37.5 cm, matrix = 256 × 128, slice
thickness/overlap = 5.2/2.6 mm, 120 images/slab, scan time = 18 seconds.

For calculation of the image-derived input function (IDIF),
dynamic PET frame times of 40 × 1 seconds, 13 × 10 seconds,
and 23 × 2 minutes were reconstructed using time of flight-ordered
subset expectation maximization with 3 iterations and 21 subsets with
corrections for decay, attenuation, scatter, random, and dead time. Time
activity curves of bone PET uptake were determined using dynamic
PET time frames of 8 × 2 seconds and 24 × 2 minutes with the same
corrections. A 3-mL venous blood sample was taken 50 minutes after
injection when arterial and venous blood concentrations have equil-
ibrated and measured in a well counter.27

**IDIF Calculation**

The IDIF was determined from \[^{18}\text{F}]\text{NaF} activity (kBq/mL)
within the popliteal artery of each knee. The artery was segmented
automatically from MR angiography images and a short time-frame
PET angiogram during the arterial phase (0–16 seconds after injec-
tion) when the tracer is predominantly in the arteries.28 In order to
minimize spillover artifacts,28 the voxels centered in the middle of
the artery were determined for each dynamic PET frame and used for
the IDIF. Central voxels were defined by including the voxels in
each axial slice within the highest 10% of arterial NaF activity.

**Bone Segmentations and Regions of Interest**

Using the in-phase, out-of-phase, and water images from the
Dixon scans, masks covering the femur, patella, and tibia were first
created by manually drawing regions of interest (ROIs) (Fig. 2). The
bone tissue was then segmented further to create subchondral/cortical
bone masks using k-means clustering (4 cluster groups minimized to
squared Euclidean distance repeated 4 times with different initial cen-
troids). The long bone of the femur and tibia was identified as the
cortical bone 6 to 8 cm from the center of the joint space. Trabecular bone
ROIs in the tibia and distal end of the femoral bone were drawn for
both legs maintaining a minimum of 3-mm distance from the edge
of the bone to avoid partial voluming. Thereafter, the subchondral bone
of the femur was manually subdivided into 5 regions: trochlea and cen-
tral and posterior regions of the medial and lateral compartments. Simi-
larly, tibial subchondral bone was further separated into lateral and
medial regions. Lastly, cortical bone at the site of a patellar tendon in-
sertion (tibial tuberosity) was identified and excluded from the analysis
of cortical bone.

**SUV and Kinetic Modeling**

The time activity curves and IDIF data were fitted to the 2-tissue
compartment model using the Patlak method and again using the non-
linear regression (NLR) method. Patlak analysis25,29 is a graphical
 technique for estimating \( k_4 \), the total rate of plasma clearance of NaF
to the bone matrix, which assumes that \(^{18}\text{F} \) is irreversibly bound to
bone mineral \( (k_4 = 0) \). Data from 10 to 50 minutes were fit to allow
for equilibration between tracer in plasma and the bone extracellular

![FIGURE 1. Hawkins 2-tissue compartment model of \(^{18}\text{F}\)NaF uptake. The parameter \( k_1 \) represents the rate of transit of the \(^{18}\text{F}\) plasma concentration to the extravascular
compartment. The accumulating fluoride concentration in bone tissue proceeds from the extravascular compartment by binding
to the bone matrix at a rate of \( k_3 \) and diffuses back
to the blood compartment at a rate defined by \( k_3 \). The
eventual rate of dissociation of the fluoride from the bone
matrix is described by \( k_4 \).](Image 24x659 to 270x702)

![FIGURE 2. Example of ROIs created in the lateral knee of 1
subject. Bone ROIs were generated by segmenting the compact
(subchondral and cortical) from trabecular bone tissue using k-means clustering. Regions of interest were then
drawn for the subchondral bone of the patella (teal),
tibia (magenta), and the femoral distal end, which was
subdivided into trochlear (green), central (blue), and
posterior (red) sections. Trabecular bone was segmented in
the femur and tibia by manually drawing ROIs keeping a
minimum distance of 3 mm from subchondral bone (shown
as light gray in the femur and dark gray in the tibia). Regions
of interest were created for cortical bone in the femoral
(yellow) and tibial shaft (orange). Lastly, a separate set of
cortical ROIs was drawn for the cortical bone at the sites of
tendon insertion (purple) on the patella and tibia.](Image 288x209 to 534x444)
fluid. Nonlinear regression fitting included estimation of 3 rate parameters \((K_1, k_2, \text{ and } k_3)\) along with a partial volume fraction, a blood fraction, and an input dispersion estimate and was computed using COMKAT software. The rate constant \(k_4\) was predefined as 0. For both blood fraction and \(K_1\), a parameter range from 0 to 1 was applied, whereas a range of 0.015 to 0.8 was used for \(k_2\) and \(k_3\), and 0 to 2 seconds for the dispersion constant \(\tau\). To avoid local minima, fits were repeated with 3 starting conditions, and results with the lowest residuals were used. The rate of total plasma clearance using the NLR method \((K^\text{NLR}_{\text{CL}})\) was calculated from the \(K_1, k_2, \text{ and } k_3\) values obtained by using the following formula:

\[
K^\text{NLR}_{\text{CL}} = K_1^{-1} \left( k_1^{-1} / (k_2 + k_3) \right)
\]

\(K^\text{NLR}_{\text{CL}}\), like \(K_1\), has units of \(\text{mL/min}\cdot\text{mL}^{-1}\), whereas \(k_2 - k_3\) have units of \(\text{min}^{-1}\).

The \(K^\text{NLR}_{\text{CL}}\) parameter can be separated into 2 parameters of physiological interest. One parameter is \(K_1\), the rate of transit of the \(^{18}\text{F}\) plasma concentration into the extravascular compartment, and reflects flow delivery of the tracer. Perfusion \((P)\) estimates for each ROI were derived from \(K_1\) values using least squares regression to the Renkin-Crone formula:

\[
K_1 = F^* (1 - \exp(-PS/F))
\]

where the product of permeability and surface area \((PS)\) was assumed to be 0.24, as reported by Piert et al.21 The second physiological parameter is the extraction fraction, \(k_3/(k_2 + k_3)\), which represents the fraction of \(^{18}\text{F}\) entering the tissue that binds to the bone matrix as opposed to reentering the bloodstream.

Images for mean SUV and SUVmax were calculated from images obtained by averaging the last 2 frames of the dynamic study (46–50 minutes).

**Statistical Analysis**

Values across the entire patient cohort are reported as median with interquartile range, and \(P\) values are from paired Student 2-tailed \(t\) tests using a threshold for significance of \(P < 0.05\) after Bonferroni correction for multiple comparisons. Correlations between obtained parameters were analyzed using linear regression where goodness of fit was evaluated with a Pearson adjusted \(R^2\) value. Reproducibility between IDIF blood activity and venous blood samples was analyzed by calculating the coefficient of variation, reported in percent. Image coregistration, ROI analysis, calculations, and statistical analysis were performed with software created in MATLAB 2013b (MathWorks, Natick, Mass).

**RESULTS**

Median parametric values along with interquartile range across all subjects are presented in Table 1. Variations in global \(\text{[}^{18}\text{F}\text{]NaF}\) uptake were observed between subjects (Fig. 3) with consequent higher or lower SUV values across all 3 types of bone tissue. Comparisons between the 3 bone tissue types are shown in Figure 4. Cortical bone had highest \(\text{[}^{18}\text{F}\text{]NaF}\) uptake for all measured parameters compared with trabecular bone \((P < 0.01)\), which had the lowest uptake. SUV and \(K_1\) values for subchondral bone were lower than that of cortical bone, but these differences were not significant after correction for multiple comparisons. Subjects had significantly higher SUVmax and \(K_1\) values and a significantly lower extraction fraction in cortical bone compared with subchondral bone. Subchondral bone had significantly higher \(\text{[}^{18}\text{F}\text{]NaF}\) uptake \((\text{SUV, SUVmax, and } K^\text{NLR}_{\text{CL}}; P < 0.01)\) than trabecular bone tissue.

There was a regional variance in distribution of \(K_1\) and extraction fraction values. The distribution ranged from cortical bone

| TABLE 1. Parametric Values for Bone Tissues of the Patellar, Tibia, and Femur |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Patellar - Subchondral | Patellar - Trabecular | Femur-Cortical (Shaft) | Femur-Subchondral | Tibia-Cortical (Shaft) | Tibia-Subchondral |
| SUVmax          | 1.93 (0.97–2.02) | 1.39 (0.71–1.58) | 1.07 (0.92–1.09) | 0.75 (0.63–0.90) | 0.56 (0.47–0.67) |
| SUVmin          | 0.44 (0.32–0.54) | 0.29 (0.13–0.36) | 0.23 (0.16–0.30) | 0.14 (0.09–0.20) | 0.10 (0.06–0.14) |
| Ki              | 0.63 (0.34–0.93) | 0.28 (0.10–0.46) | 0.22 (0.13–0.33) | 0.14 (0.08–0.22) | 0.11 (0.06–0.17) |
| Ki (mg/mL/min/mL) | 10^{-2} | 10^{-2} | 10^{-2} | 10^{-2} | 10^{-2} |
| SUVmean         | 0.71 (0.43–1.00) | 0.52 (0.38–0.94) | 0.52 (0.38–0.94) | 0.52 (0.38–0.94) | 0.52 (0.38–0.94) |
| Ki (ml/min/mL)  | 0.29 (0.14–0.43) | 0.22 (0.13–0.33) | 0.22 (0.13–0.33) | 0.13 (0.07–0.20) | 0.12 (0.06–0.17) |
| Ki (ml/min/mL)  | 0.29 (0.14–0.43) | 0.22 (0.13–0.33) | 0.22 (0.13–0.33) | 0.13 (0.07–0.20) | 0.12 (0.06–0.17) |
| Ki (mg/mL/min/mL) | 10^{-2} | 10^{-2} | 10^{-2} | 10^{-2} | 10^{-2} |
| Blood, %vol     | 0.05 (0.03–0.07) | 0.05 (0.03–0.07) | 0.05 (0.03–0.07) | 0.05 (0.03–0.07) | 0.05 (0.03–0.07) |

Median parametric values from different bone tissue with interquartile range are parenthesized. Measures are further divided into the tibial, femoral, and patellar bones. Comparisons between tissue types are shown in Figure 4.
of the shaft, which had the highest vascularization where $K_1 > K_i$ and $k_3/(k_3 + k_2) < 1$, to the trochlea and patella region of subchondral bone, where $k_3/(k_3 + k_2) ≈ 1$ and $K_1 ≈ K_i$ (Fig. 5). By visual analysis of $K_1$ and $k_3/(k_3 + k_2)$ maps, a negative gradient of $K_1$ values can be seen from the femoral and tibial shafts decreasing toward the joint space. A second gradient can be seen as $K_1$ is higher in subchondral bone and declines toward the center of the trabecular bone of the femur and tibial head (Fig. 6). In the subchondral bone of the femur, $K_1$ and blood volume values were higher in the posterior section, decreasing to the lowest in the trochlea ($P < 0.01$). The

![FIGURE 3. SUV of representative slice from 2 subjects. There was a wide intersubject range of $[^{18}F]$NaF uptake across the joint. The left image is a subject with low uptake in knee, whereas the right image is a subject with high uptake in all bone tissues. In addition to global variations in tracer uptake between subjects, some individual variations in the relative distribution of $[^{18}F]$NaF uptake across bone tissues regions were observed. In this example, the subject on the right has relatively low uptake in the subchondral bone of the femur compared with the subchondral bone of the patella and tibial head, whereas the subject on the left has equally low uptake in all subchondral regions.](image)

![FIGURE 4. Parametric values of $[^{18}F]$NaF uptake for different bone tissue types of the knee. Cortical bone had the highest $[^{18}F]$NaF uptake in all measured parameters when compared with trabecular bone, which had the lowest uptake. Subchondral bone also had higher uptake than trabecular bone with significantly higher SUVmean, SUVmax, and $K_i^{NLR}$ values, yet only slightly elevated $K_1$ and $k_3/(k_3 + k_2)$ values. The relative distribution of $K_i^{NLR}$ values between bone tissues was almost identical to that of SUV. Note that despite having the highest uptake as expressed by SUV and $K_i^{NLR}$, cortical bone has the lowest extraction fraction. $P$ values were corrected for 18 comparisons using a Bonferroni correction ($\dagger P < 0.01$ difference compared with cortical bone, $\ddagger P < 0.01$ difference between trabecular bone and subchondral bone).](image)
opposite gradient was observed in the extraction fraction maps, resulting in total metabolism $K_{3NLR}$ and SUV images that were more spatially uniform.

There were no other significant differences in $[^{18}F]$NaF uptake parameters between the 3 bone tissue types or between subregions of subchondral bone. The sites of tendon insertion had elevated SUV values and significantly higher $K_{3NLR}$ ($P < 0.05$) than remaining cortical bone tissue. The vascularization was lower ($K_1$ 33% less, blood volume 84% lower [both $P > 0.01$]), and the extraction fraction higher ($k_3/(k_3 + k_2)$ 88% higher, $P < 0.01$).

$K_{3NLR}$ values correlated highly with SUV values ($R^2 = 0.90$). $K_{3pat}$ values from the Patlak method had a slightly poorer correlation to SUV ($R^2 = 0.87$) and were 17% lower than those obtained by NLR (Fig. 7). The correlation of $K_{3pat}$ values to $K_{3NLR}$ values was high ($R^2 = 0.97$) despite the 17% bias (Fig. 7). Using $K_1$ values from the NLR fit, flow values were obtained and found to be within a few percentage points of $K_1$ values (Table 1). The $K_1$ values were in the range where $K_1 << PS$, and thereby the condition $F \approx K_1$ applies.

Group average IDIF values at 1, 5, 10, and 50 minutes were 10.2, 6.0, 4.2, and 2 kBq/mL when normalized to a 100-MBq injection. At 50 minutes, mean IDIF values were 6% higher than mean venous blood sample values. Coefficient of variation values between venous blood samples and IDIF values measured at 50 minutes were 8.3%. Repeated injections in 1 subject had mean coefficient of variation values of 9% across time points observed between 1 and 50 minutes (Fig. 8).

FIGURE 5. Components of $K_i$ for different bone tissue type. The SUV/$K_{3NLR}$ relationship did not vary significantly between tissue types or between subjects. Linear regression analysis of SUV and $K_{3NLR}$ gave SUV = 89 x $K_{3NLR}$. The first column compares different ROI values from all subjects to this regression. Given that $K_i = K_1 \times k_3/(k_3 + k_2)$, $K_1$ can be broken into a flow-related $K_1$ component, which is the rate of tracer entering the tissue, and an extraction fraction component, $k_3/(k_3 + k_2)$, which is the fraction of the tracer having entered the tissue that binds to the bone matrix. SUV of the cortical bone (shaft) is correlated to both $K_1$ and $k_3/(k_3 + k_2)$ values, whereas in trabecular bone and the patella, SUV is primarily determined by $K_1$ where $K_i = K_1$ and $k_3/(k_3 + k_2) = 1$ in all subjects.

DISCUSSION

Semiquantitative and quantitative values for $[^{18}F]$NaF uptake in the knee were obtained from healthy subjects using PET/MRI. A large intersubject variation in NaF uptake was observed as there were significant differences in uptake parameters between cortical bone and the subchondral/trabecular bone tissues. Trabecular bone was found to have significantly lower SUV, $K_1$, $K_2$, and blood volume values yet a significantly higher extraction fraction than the cortical bone tissue in the shaft of the femur and tibia. Blood volume was the parameter with the largest discrepancy between bone tissues being significantly higher in the shaft compared with subchondral or trabecular bone of the knee. Subjects had higher vascularization (larger blood volume and higher $K_1$ values) in the shaft of the femur and tibia declining with a negative gradient toward the joint space reaching the lowest values at the center of the trabecular bone near the distal end. This $K_1$ gradient was partially offset by a gradient of increasing extraction efficiency that was significantly lower in the shaft. A similar regional discrepancy was also evident in SUV, SUVmax, and $K_{3NLR}$, although to a lesser degree. These parameters, like $K_1$, were significantly higher in the shaft decreasing in the subchondral bone and trabecular bone of the knee joint in these healthy individuals. Likewise, the sites of tendon insertion of the cortical bone had much lower vascularization ($K_1$ and blood volume), yet a net uptake than regular cortical bone due to a high extraction fraction. A similar observation has been
made between the spine and humeral bone tissues where low $K_1$ values in the humeral bone were partially compensated by a higher $k_3/(k_2 + k_3)$ to give a more comparable, yet still significantly lower, $K_i$ value. Aside from the $K_1$ and $k_3/(k_2 + k_3)$ gradients, all other parametric values within the subchondral bone tissue ROIs of subjects were quite homogenous with no significant differences when comparing subchondral subregions across the patella, femur, and tibia.

$K_i^{Pat}$ values from the Patlak method were 17% lower than those obtained by NLR, which is a larger bias than previously reported by Siddique et al., where $K_i^{Pat}$ was 10% lower than $K_i^{NLR}$ in the lumbar vertebrae. Still, $K_i^{NLR}$ values correlated highly with both $K_i^{Pat}$ values ($R^2 = 0.97$) and SUV ($R^2 = 0.90$) with no regional variations in their correlation. Ultimately, this study gives no evidence of meaningful differences in using Patlak or NLR methods to determine $K_i$ as they could be interchanged with a conversion factor. Studies including mean SUV, $K_i^{Pat}$, and $K_i^{NLR}$ have found these parameters to have similar reproducibility with coefficients of variation ranging between 9% and 15%.

$K_i^{NLR}$ had lower reproducibility when $k_1$ is not limited to 0 when fitting. In this study, $SUV$, $K_i^{Pat}$, and $K_i^{NLR}$ have comparable variance where intersubject SDs are between 43% and 46% of mean values. Despite similar reproducibility, $K_i$ values have been reported to be a more sensitive measure of regional bone metabolism than SUV.7,17,18,27,42 In the limbs, where $F$ uptake is low, Brenner et al.18 and Apostolova and Brenner15 concluded the minimal change of SUV in a patient must be greater than 50% to reliably detect disease or treatment-related changes, whereas the same diagnosis could be made from a change in $K_i$ of 25%. $K_i$ values have also shown to be more sensitive when analyzing alterations in subchondral bone of the femur adjacent to cartilage defects.4 SUVmax values in this study are similar to previously reported mean SUVmax values of 2.44 for the tibia3 and 2.22 in the femur shaft.44 SUVmax has been found to correlate well with adjacent cartilage alterations,24 and although it had the largest intrasubject variation in this study, it had a relatively lower variance between subjects and greater differentiation between bone tissues (Fig. 4). In this study, using NLR was advantageous as obtaining $K_i$ and extraction fraction parameters provided useful information that could not be extracted from $K_i$ alone.

The $K_i$ values obtained in this study were within a flow-dominant regimen where it has, theoretically, a linear correlation to blood flow ($K_i < <$ PS). The flow values obtained in this study compare well with measured blood flow in the femoral shaft,45 but lack a criterion-standard measure to investigate $K_i$ as a surrogate flow measure. To date, the most convincing studies to confirm the relationship between blood flow in bone tissue and $K_i$ for $[^{18}F]$NaF kinetics have been performed in swine vertebrae.15,21 Since then, authors have reported a poor correlation between $K_1$ and bone perfusion in studies of the hip of human surgery patients22 and the forelimbs of healthy rats.46 Obtaining an estimate of flow would be of great clinical value. Bone perfusion is usually linked to metabolic activity and varies greatly between different bones and bone regions in the skeleton where the extremities are among the lowest.15,25,47

FIGURE 6. Example of the distribution of $K_i$, $K_i^{NLR}$ and SUV from 1 subject. Subjects had a negative gradient of $K_i$ values from the shaft toward the joint space and from the subchondral bone toward the center of the distal end of the femoral bone and tibial head. In the sagittal plane, $K_i$ values were highest in the posterior section of subchondral bone and decreased in the anterior direction toward the trochlea and patella. However, the opposite gradient was observed in extraction fraction $k_3/(k_2 + k_3)$ maps, resulting in $K_i^{NLR}$ and SUV images with more localized heterogeneity.
Perfusion studies using microspheres have shown a reduction of blood flow in bones related to age, osteoporosis, and reduced endothelium-dependent vasodilation.

With regard to this study's aim to report key parametric values for $[^{18}F]NaF$ uptake in the healthy knee, there are several limitations to be considered when interpreting the results. First, the number of subjects is small where results can be skewed by relatively few abnormalities. The range of ages (22–44 years) is a period of rather stable bone density in human adults, but factors such as body mass index, varus/valgus alignment, disease, or activity level could alter the kinetics in bone tissue. Second, despite the numerous advantages from combining PET imaging with MRI in knee examinations, there are disadvantages in foregoing the superior information on bone density, which CT provides. Dixon-based methods, as used by the scanner in this study, have been shown to underestimate bone $[^{18}F]NaF$ mean SUV by 10%, ranging between 0% and 20% depending on location. The subchondral bone would be least affected being close to the bone surface, whereas the trabecular bone could have a more pronounced underestimation of SUV due to improper attenuation correction. Likewise, a similar underestimation of $K_I$ and $K_i$ would be expected, although it would be partially offset by a similar underestimation of activity in input function obtained from the popliteal artery. Lastly, the use of an IDIF would best be confirmed by using arterial sampling as a criterion standard. In this study, venous samples confirmed the activity of the later phase of the IDIF but not the earlier phase of high activity.

PET/MRI is an optimal dual-imaging combination offering the advantages of the high soft tissue contrast and resolution of MRI and the sensitivity of PET. In this study, MR angiography added the advantage of segmenting the popliteal artery, making an automated process to obtain the IDIF possible. The input functions obtained correspond well with literature values for $[^{18}F]NaF$ from arterial sampling, and visual inspection of generated ROIs confirmed successful automated segmentation of the popliteal artery. Mean IDIF values 50 minutes after injection were 6% higher than venous blood samples taken on an equilibrium time point, whereas Cook et al found arterial blood samples to be 2% higher than venous blood samples after 24 minutes. With the increased use of NaF in nononcological studies of the skeleton, it has become even more relevant as moderate differences in NaF uptake may be an early indication of bone degradation in diseases such as osteoarthritis. The combination of PET/MRI reduces the radiation dose significantly in 2 ways. First by eliminating CT and, second, because the PET data are acquired for the duration of the MRI protocol (which can be up

**FIGURE 7.** Comparison of $K_{NLR}$ with SUV values and $K_{Pat}$ Patlak. A, Scatterplot of $K_{NLR}$ results from all ROIs of all subjects plotted against ROI mean SUV ($R^2 = 0.90$). The Bland-Altman plot compares SUV values and $K_{NLR}$ values multiplied by the slope determined from the regression (slope = 90). B, A scatterplot with regression fit and Bland-Altman plot of the same ROIs comparing $K_{NLR}$ vs $K_{Pat}$ values ($R^2 = 0.97$). The Patlak method produced $K_i$ values that were 17% lower than those obtained by NLR and had a slightly poorer correlation to SUV ($R^2 = 0.87$).
to an hour), the injected dose of $^{18}$F-fluoride can be decreased from a standard clinical dose of 200 MBq to 90 MBq (used in this study) and still retain the same signal-to-noise ratio in PET SUV maps. The effective dose of this study is estimated to be 2.16 mSv. Quantitative MRI techniques have been widely studied to develop robust biomarkers for the early detection and monitoring of osteoarthritis and semiquantitative evaluation of bone metabolism in the knee at a joint. We have shown the feasibility of using PET/MR to create an image-derived input function of $[{\text{F}}]$NaF repeated in 1 subject. One subject underwent injections on

**CONCLUSIONS**

This study showed significant variations in regional bone perfusion and metabolism between skeletal tissue types in the knee joint. We have shown the feasibility of using PET/MR to create an accurate IDIF from the popliteal artery and to conduct a quantitative and semiquantitative evaluation of bone metabolism in the knee at a low radiation dosage. $^{18}$F-NaF PET/MRI is a noninvasive technique that offers an attractive tool to simultaneously estimate bone perfusion and metabolism at clinically relevant sites of the knee.

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