JASPAR 2020

update of the open-access database of transcription factor binding profiles

Fornes, Oriol; Castro-Mondragon, Jaime A; Khan, Aziz; van der Lee, Robin; Zhang, Xi; Richmond, Phillip A; Modi, Bhavi P; Correard, Solenne; Gheorghe, Marius; Baranaši, Damir; Santana-Garcia, Walter; Tan, Ge; Chèneby, Jeanne; Ballester, Benoit; Parcy, François; Sandelin, Albin; Lenhard, Boris; Wasserman, Wyeth W; Mathelier, Anthony

Published in:
Nucleic Acids Research

DOI:
10.1093/nar/gkz1001

Publication date:
2020

Document version
Publisher's PDF, also known as Version of record

Document license:
CC BY-NC

Citation for published version (APA):

Download date: 13. jun., 2021
JASPAR 2020: update of the open-access database of transcription factor binding profiles

Oriol Fornes1,†, Jaime A. Castro-Mondragon2,‡, Aziz Khan2,†, Robin van der Lee1, Xi Zhang1, Phillip A. Richmond1, Bhavi P. Modi1, Solenne Correard1, Marius Gheorghe2, Damir Baranasič3,4, Walter Santana-Garcia5, Ge Tan6, Jeanne Chèneby7, Benoit Ballester8, François Parcy8, Albin Sandelin9,*, Boris Lenhard3,4,10,*, Wyeth W. Wasserman1,‡ and Anthony Mathelier2,11,*

1Centre for Molecular Medicine and Therapeutics, Department of Medical Genetics, BC Children’s Hospital Research Institute, University of British Columbia, 950 W 28th Ave, Vancouver, BC V5Z 4H4, Canada, 2Centre for Molecular Medicine Norway (NCMM), Nordic EMBL Partnership, University of Oslo, 0318 Oslo, Norway, 3Institute of Clinical Sciences, Faculty of Medicine, Imperial College London, London W12 0NN, UK, 4Computational Regulatory Genomics, MRC London Institute of Medical Sciences, London W120NN, UK, 5Institut de Biologie de l’ENS (IBENS), Département de biologie, École normale supérieure, CNRS, INSERM, Université PSL, 75005 Paris, France, 6Functional Genomics Centre Zurich, ETH Zurich, Zurich, Switzerland, 7Aix Marseille Univ, INSERM, TAGC, Marseille, France, 8CNRS, Univ. Grenoble Alpes, CEA, INRA, IRIG-LPCV, 38000 Grenoble, France, 9The Bioinformatics Centre, Department of Biological and Biotech Research & Innovation Centre, University of Copenhagen, DK2200 Copenhagen N, Denmark, 10Sars International Centre for Marine Molecular Biology, University of Bergen, N-5008 Bergen, Norway and 11Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, 0310 Oslo, Norway

Received September 15, 2019; Revised October 15, 2019; Editorial Decision October 16, 2019; Accepted October 16, 2019

ABSTRACT

JASPAR (http://jaspar.genereg.net) is an open-access database of curated, non-redundant transcription factor (TF)-binding profiles stored as position frequency matrices (PFMs) for TFs across multiple species in six taxonomic groups. In this 8th release of JASPAR, the CORE collection has been expanded with 245 new PFMs (169 for vertebrates, 42 for plants, 17 for nematodes, 10 for insects, and 7 for fungi), and 156 PFMs were updated (125 for vertebrates, 28 for plants and 3 for insects). These new profiles represent an 18% expansion compared to the previous release. JASPAR 2020 comes with a novel collection of unvalidated TF-binding profiles for which our curators did not find orthogonal supporting evidence in the literature. This collection has a dedicated web form to engage the community in the curation of unvalidated TF-binding profiles. Moreover, we created a Q&A forum to ease the communication between the user community and JASPAR curators. Finally, we updated the genomic tracks, inference tool, and TF-binding profile similarity clusters. All the data is available through the JASPAR website, its associated RESTful API, and through the JASPAR2020 R/Bioconductor package.

INTRODUCTION

Transcription factors (TFs) are proteins involved in the regulation of gene expression at the transcriptional level (1). They interact with DNA in a sequence-specific manner through their DNA-binding domains (DBDs), which are used to classify TFs into structural families (2). The genomic locations where TFs bind to DNA are known as TF-binding sites (TFBSs), which are typically short (6–20 bp) and exhibit sequence variability (3). Genome-wide identification of TFBSs is key to understanding transcriptional regulation. As it is not possible to identify all TFBSs for every cell type and cellular condition experimentally, computational modeling of TF-binding specificities has been instrumental to predict TFBSs in the genome. These compu-
Table 1. Overview of the growth of the number of PFMs in the JASPAR 2020 CORE and unvalidated collections compared to the JASPAR 2018 CORE collection

<table>
<thead>
<tr>
<th>Taxonomic Group</th>
<th>Non-redundant PFMs in JASPAR 2018</th>
<th>New non-redundant PFMs in JASPAR 2020</th>
<th>Removed profiles</th>
<th>Updated PFMs in JASPAR 2020</th>
<th>Total PFMs (non-redundant) in JASPAR 2020</th>
<th>Total PFMs (all versions) in JASPAR 2020</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vertebrates</td>
<td>579</td>
<td>169</td>
<td>2</td>
<td>125</td>
<td>746</td>
<td>1011</td>
</tr>
<tr>
<td>Plants</td>
<td>489</td>
<td>42</td>
<td>1</td>
<td>28</td>
<td>530</td>
<td>572</td>
</tr>
<tr>
<td>Insects</td>
<td>133</td>
<td>10</td>
<td>0</td>
<td>3</td>
<td>143</td>
<td>153</td>
</tr>
<tr>
<td>Nematodes</td>
<td>26</td>
<td>17</td>
<td>0</td>
<td>0</td>
<td>43</td>
<td>44</td>
</tr>
<tr>
<td>Fungi</td>
<td>176</td>
<td>7</td>
<td>0</td>
<td>0</td>
<td>183</td>
<td>184</td>
</tr>
<tr>
<td>Urochordata</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td><strong>Total CORE</strong></td>
<td><strong>1404</strong></td>
<td><strong>245</strong></td>
<td><strong>3</strong></td>
<td><strong>156</strong></td>
<td><strong>1646</strong></td>
<td><strong>1964</strong></td>
</tr>
<tr>
<td><strong>unvalidated</strong></td>
<td><strong>337</strong></td>
<td><strong>337</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Figure 1. JASPAR CORE growth. The number of profiles in each taxon and overall (see legend) through all JASPAR releases.

Here, we present the 8th release of JASPAR, which comes with a major expansion and update of its CORE collection. Moreover, we introduce a new collection of unvalidated profiles, which stores quality-controlled PFMs for which our curators could not find orthogonal support. This collection has a dedicated web interface to engage the community of users in the curation of TF-binding profiles. Finally, we have updated the hierarchical clusters of TF-binding profiles, the genomic tracks of predicted TFBSs (now available for 8 genomes), and the profile inference tool.

**EXPANSION AND UPDATE OF THE JASPAR CORE COLLECTION**

For this 8th release of JASPAR, we added to the CORE collection 245 new TF-binding profiles for TFs in the following taxa: vertebrates (169 profiles, corresponding to an expansion of 29% for this taxon), plants (42 profiles, 9% expansion), nematodes (17 profiles, 65% expansion), insects (10 profiles, 8% expansion) and fungi (7 profiles, 4% expansion). We updated 156 profiles (Table 1). The new PFMs were derived from HT-SELEX (30), PBMs (20), ChIP-seq and DAP-seq experiments (data sourced from CistromeDB (31), ReMap (32,33), GTRD (34), ChIP-atlas (35) and ModERN (36), see Supplementary Text for method details). As pre-
Figure 2. Unvalidated TF-binding profile collection. Example with the ZNF793 profile. This high-quality PFM was derived from a ChIP-seq experiment and was built from thousands of potential TFBSs. Further, the TFBSs are enriched around the ChIP-seq peak summits. However, no orthogonal evidence supporting this profile was found by our curators. Users can upload relevant information about the profile in the unvalidated collection through the ‘Community curation’ box.

Previously described, the newly introduced profiles were manually curated to be supported by an orthogonal reference from the literature, which is provided in the metadata of the profiles. Moreover, the TF DBD class and family (following the TFClass classification (2)), the TF UniProt ID (37), and links to the TFBSshape (24,25), ReMap (32,33) and UniBind (38) databases are provided in the profiles metadata (whenever possible). Finally, the profiles previously associated with ID2, ID4 and TRB2 were removed from the CORE collection as these proteins are not TFs (1).

Overall, the JASPAR 2020 CORE collection includes 1646 non-redundant PFMs (746 for vertebrates, 530 for plants, 183 for fungi, 143 for insects, 43 for nematodes and 1 for urochordates) (Table 1; Figure 1). Moreover, we continued with the incorporation of novel transcription factor flexible models (TFFMs), which are hidden Markov-based models capturing dinucleotide dependencies in TF–DNA interactions (11). We introduced new TFFMs for 217 TFs (136 for vertebrates, 38 for plants, 21 for insects, 17 for nematodes, and 5 for fungi) and updated TFFMs for 20 verte-
brates TFs, which represents a 50% increase in the number of TFFMs available. All data is available on the JASPAR website, its associated RESTful API, and through the JASPAR2020 R/Bioconductor package.

A NEW COLLECTION OF UNVALIDATED PROFILES FOR COMMUNITY ENGAGEMENT

We introduced a novel ‘unvalidated’ collection to store high-quality (i.e. passing multiple quality controls, see Supplementary Text) TF-binding profiles for which no independent support was found in the literature by our curators. This collection contains 337 PFM. As these profiles are not yet supported by an orthogonal evidence, we recommend users to use this collection with caution. We encourage the community to engage in the curation of these profiles by providing the JASPAR curators with supporting complementary evidence (from their own work or others) whenever possible. This is facilitated by the availability of an individual submission form for each profile in the ‘unvalidated’ collection (Figure 2).

Further, we started a Q&A forum (https://groups.google.com/forum/#!forum/jaspar) to ease the communication between JASPAR curators and the community; we welcome the community to send us their questions and suggestions, or to report errors in JASPAR.

CLUSTERED PROFILES, GENOMIC TRACKS AND PROFILE INFERENCE TOOL

In the previous releases, we introduced novel features such as hierarchical clustering of TF-binding profiles in the CORE collection to visualize profile similarities, genomics tracks of predicted TFBSs, and an inference tool to predict TF-binding profiles likely recognized by TFs not available in JASPAR CORE. We improved the profile inference tool using our own implementation of a recently described similarity regression method (20). We updated the generation of genomic tracks that are publicly available through the UCSC Genome Browser data hub (39) for 7 organisms: human (hg19, hg38), mouse (mm10), zebrafish (danRer11), Drosophila melanogaster (dm6), Caenorhabditis elegans (ce10), Arabidopsis thaliana (araTha1) and baker’s yeast (sacCer3). For more details on the updated genomic tracks and inference tool, refer to the Supplementary Text. Finally, we generated the hierarchical clusters of available TF-binding profiles for each taxon with RSAT matrix-clustering (40). Users can explore the CORE/unvalidated collection through the trees and access directly the corresponding profiles by clicking on the TF name.

CONCLUSIONS AND PERSPECTIVES

Similar to previous releases, we substantially expanded the CORE collection of the JASPAR database. For this 8th release, we processed more than 18,000 ChIP-seq datasets. As a large number of the obtained high-quality TF-binding profiles were not supported with orthogonal supporting evidence, it motivated us to create the novel ‘unvalidated’ collection of profiles. We expect that upcoming experiments and publications will provide additional supporting evidence to some profiles to be incorporated into the JASPAR CORE collection. Meanwhile, we would like to extend our invitation to the research community to 1) help us curate these unvalidated profiles (e.g. by pointing us to supporting literature), and 2) send us their own novel profiles (e.g. determined experimentally) for incorporation in the next release of JASPAR.

The JASPAR CORE vertebrates collection now contains 746 profiles, 637 of which are associated with human TFs with known DNA-binding profiles (1), which corresponds to a 58% of the 1,107 reported by Lambert et al. (1). While this is an impressive collective achievement by the field (the original JASPAR database only contained 81 profiles, a ~7% coverage for human TFs), it suggests that targeted experimental efforts to find the binding preferences for remaining TFs will be important. Although computational approaches can be used to infer missing TF-binding profiles (20,41), especially for non-model organisms, the JASPAR approach is conservative, including profiles supported by at least two experiments in the literature. This is very important as we stand by the reliability of our data. Since its initial publication in 2004 (23), the JASPAR database has been committed to provide the research community with high-quality, manually curated, non-redundant TF-binding profiles.

Lastly, although PFM have dominated the field of gene regulation for decades, new profile representations have emerged. For example, profiles with expanded alphabets to represent methylated bases (42,43), modelling binding energy (44) or derived from deep learning importance scores (45). Depending on how the field evolves and how popular these profiles become, we will consider them for inclusion in JASPAR in the future.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

ACKNOWLEDGEMENTS

We thank the user community for useful input and the scientific community for performing experimental assays of TF–DNA interactions and for publicly releasing the data. We thank Giovanna Ambrosini for her help with PWM Scan, the UCSC Genome Browser Project Team for their assistance with the genome tracks, WestGrid (https://www.westgrid.ca), Compute Canada (https://www.computecanada.ca), Georgios Magklaras and Georgios Marselis for their IT support, Jacques van Helden and Adam Handel for contacting us to add and validate TF binding profiles, and Dora Pak and Ingrid Kjelsvik for administrative support.

FUNDING

Norwegian Research Council [187615]; Helse Sør-Øst; University of Oslo through the Centre for Molecular Medicine Norway (NCMM) (to A.M., J.A.C.-M., A.K., M.G.); Norwegian Research Council [288404 to J.A.C.-M. and Mathelier group]; The Norwegian Cancer Society [197884 to Mathelier group]; O.F., X.Z., P.A.R., S.C. and W.W.W. were supported by grants from the Canadian Institutes of Health.
This paper is linked to: https://doi.org/10.1093/nar/gkz945.


