Cerebral Plasmodium falciparum malaria
Jensen, Anja Ramstedt; Adams, Yvonne; Hviid, Lars

Published in:
Immunological Reviews

DOI:
10.1111/imr.12807

Publication date:
2020

Document version
Publisher's PDF, also known as Version of record

Document license:
CC BY

Citation for published version (APA):
Cerebral *Plasmodium falciparum* malaria: The role of PfEMP1 in its pathogenesis and immunity, and PfEMP1-based vaccines to prevent it

Anja Ramstedt Jensen¹ | Yvonne Adams¹ | Lars Hviid¹,²

¹Centre for Medical Parasitology at Department of Immunology and Microbiology, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark
²Department of Infectious Diseases, Rigshospitalet, Copenhagen, Denmark

Correspondence
Anja Ramstedt Jensen, Department of Immunology and Microbiology, Faculty of Health and Medical Sciences, University of Copenhagen, Maersk Tower 07-11, Panum Institute, Blegdamsvej 3B, 2200 Copenhagen N, Denmark. Email: atrj@sund.ku.dk

Funding information
Augustinus Fonden; The Consultative Committee for Developmental Research, Grant/Award Number: 17-02-KU; Independent Research Fund Denmark, Grant/Award Number: 4183-00539 and 8020-00034B; Novo Nordisk Fonden, Grant/Award Number: NNF16OC0022298; Hørsholm Fonden; Aase og Ejnar Danielsen Fonden, Grant/Award Number: 10-02129; Læge Sofus Carl Emil Friis og hustru Olga Doris Friis Legat; Svend Andersen Fonden; AP Møller Fonden, Grant/Award Number: 18-L-0040

**Abstract**

Malaria, a mosquito-borne infectious disease caused by parasites of the genus *Plasmodium* continues to be a major health problem worldwide. The unicellular *Plasmodium*-parasites have the unique capacity to infect and replicate within host erythrocytes. By expressing variant surface antigens *Plasmodium falciparum* has evolved to avoid protective immune responses; as a result in endemic areas anti-malaria immunity develops gradually over many years of multiple and repeated infections. We are studying the role of *Plasmodium falciparum* erythrocyte membrane protein 1 (PfEMP1) expressed by asexual stages of *P. falciparum* responsible for the pathogenicity of severe malaria. The immunopathology of *falciparum* malaria has been linked to cyto-adhesion of infected erythrocytes to specific host receptors. A greater appreciation of the PfEMP1 molecules important for the development of protective immunity and immunopathology is a prerequisite for the rational discovery and development of a safe and protective anti-disease malaria vaccine. Here we review the role of ICAM-1 and EPCR receptor adhering *falciparum*-parasites in the development of severe malaria; we discuss our current research to understand the factors involved in the pathogenesis of cerebral malaria and the feasibility of developing a vaccine targeted specifically to prevent this disease.

**KEYWORDS**

antibodies, cerebral malaria, immunity, PfEMP1, *Plasmodium falciparum*, vaccine

1 | INTRODUCTION

Infection with *Plasmodium falciparum* parasites causes the most severe form of malaria that is responsible for essentially all malaria-related deaths. The ability of *P. falciparum*-infected erythrocytes (IEs) to adhere efficiently to host vascular receptors sets this parasite aside from the other malaria parasites infecting humans, and is generally considered an important reason why *P. falciparum* malaria is particularly dangerous.

IE adhesion is called sequestration when the IEs stick to tissue-bound receptors, rosetting when they stick to uninfected erythrocytes, and clumping when the IEs stick to each other. It can lead to circulatory disturbances, vascular occlusion, and inflammation. In all cases, the IEs interact with host receptors via members of...
parasite-encoded antigens displayed on the IE surface. These antigens belong mainly—if not exclusively—to products of several multigene families. Prominent among them—and by far the best studied—is \textit{Plasmodium falciparum} erythrocyte membrane protein 1 (PfEMP1), encoded by the clonally variant \textit{var} gene family with approximately 60 members per parasite genome. This review focuses on PfEMP1, the putative role of this antigen family in the development of one of the most severe forms of malaria called cerebral malaria (CM) and in acquired immunity to CM, and finally on the prospect of a PfEMP1-based vaccine to prevent this often fatal complication. Before discussing each of these aspects it is necessary to recapitulate briefly the parasite life cycle, as it is important for appreciating the sections that follow.

1.1 | The parasite multiplication cycle
\textit{Plasmodium falciparum} has a complex life cycle that involves two hosts (humans and \textit{Anopheles} spp. mosquitoes), and several developmental stages in each host (Figure 1). The human part of the multiplication cycle, which is asexual, is initiated when a \textit{P. falciparum}-infected female mosquito injects sporozoite-stage parasites into the skin while it is feeding for blood. The extracellular sporozoites rapidly transit via the peripheral circulation from the skin to the liver, where they infect hepatocytes. The liver stage is asymptomatic and lasts for approximately 1 week, during which time the intrahepatic parasite multiplies, resulting in a (pre- or extraerythrocytic) schizont that consists of at least 30,000 daughter parasites. These, now called (pre- or extraerythrocytic) merozoites leave the infected hepatocyte and enter the blood circulation. The merozoites rapidly infect erythrocytes, an event that marks the beginning of the intraerythrocytic multiplication cycle. This part of the life cycle continues until the infection is controlled by either immunity or chemotherapy, or until the host dies. Each round of the intraerythrocytic cycle lasts approximately 48 hours. During each, the newly invaded merozoite rapidly transforms to a trophozoite (the early trophozoite is often called a ring-stage parasite, because of the prominent vacuole) that undergoes three to five mitotic divisions, resulting in a schizont. At the end of the intraerythrocytic cycle, the IE ruptures and the released (erythrocytic) merozoites rapidly invade new erythrocytes. Some of the newly invaded merozoites develop into male or female gametocytes rather than continuing the asexual multiplication cycle. The gametocytes do not divide, but remain inside the erythrocyte until taken up by a blood-feeding mosquito, where sexual reproduction and further asexual multiplication steps complete the parasite life cycle.\textsuperscript{1-3}

2 | \textbf{THE P. FALCIPARUM ERYTHROCYTE MEMBRANE PROTEIN 1 ANTIGENS}

The expression of PfEMP1 is largely (but not exclusively) restricted to the intraerythrocytic blood stages of the infection, where these high-molecular weight proteins mediate IE adhesion to a variety of host receptors. Intracellular PfEMP1 can be detected a few hours
after the merozoite has invaded an erythrocyte, whereas IE surface expression starts about 16 hours postinvasion. 5 The PfEMP1 surface expression reaches a plateau about 8 hours later, and starts to decrease during the final hours of the 48-hour cycle, 6 since export of PfEMP1 to the IE surface stops 30-36 hours postinvasion. 7

2.1 | PfEMP1 structure

The members of the PfEMP1 family are high-molecular weight proteins (200-450 kDa), encoded by approximately 60 two-exon var genes per haploid P. falciparum genome. 5 The extracellular part of PfEMP1, which is encoded by exon I, is composed of cysteine-rich interdomain regions (CIDRs) and 2-10 Duffy-binding-like (DBL) domains. These DBL and CIDR domains can be divided into seven (α, β, γ, δ, ε, ζ, x) and three (α, β, γ) main sequence classes, respectively, each with many further subdivisions. 3 While exon I is characterized by extensive intracranial (within single genomes) and intercultural (between genomes) sequence variation, the short transmembrane domain and the acidic intracellular terminal segment (ATS) are encoded by the relatively conserved exon II. 8

The PfEMP1 CIDR domains are characterized by conserved cysteine-rich motifs, 9 while the DBL domains are homologous to P. falciparum EBA-175 adhesive domains and to the Duffy-binding proteins of P. vivax and P. knowlesi. 10 The DBL domains are composed of three structural subdomains (Figure 2A), which have a mixed helix-sheet structure (S1) or consist of helix bundles (S2 and S3). 11 The subdomains are held together by disulphide bonds between conserved cysteine residues, 12 whereas the α-helices of the CIDR and DBL domains are connected by flexible and/or ordered loops. The functional specificity of different PfEMP1 proteins often (but not always) depends on these highly variable loops. 12-14

Despite the sequence variability, PfEMP1 proteins can be grouped according to their chromosomal location, upstream promoter sequence (ups), and direction of transcription of the var genes encoding them. 15 Group A (10 genes in P. falciparum 3D7), Group B (22 genes), and Group B/A var genes (4 genes) are all found in the subtelomeric regions of chromosomes. Group A genes are transcribed toward the telomere, whereas Group B and B/A var genes are transcribed toward the centromere (Figure 2B). Group C (13 genes in P. falciparum 3D7) and Group B/C var genes (9 genes) are typically found in internal regions of chromosome 4, 7, 8 and 12. 5,8,15,16 All PfEMP1 subfamilies except two (Type 3 and VAR2CSA) have a head structure at their N-terminus that is composed of semiconserved DBLx domain and a CIDR domain. 5 This is followed by a second and more diverse DBL-CIDR pair in most PfEMP1 proteins belonging to Group B, B/C, and C. Group A and B/A PfEMP1 proteins are composed of a total of 7-10 extracellular domains, including additional DBL domains upstream and/or downstream of the second DBL-CIDR pair (Figure 2C).

The combination of DBL and CIDR subtypes in different PfEMP1 proteins is non-random, and has led to the identification of 21 domain trains called domain cassettes (DCs). 8 The DCs are defined as var gene sequences encoding two or more DBL or CIDR domains with subclasses that can be predicted from each other, and they often predict the receptor specificity of the encoded PfEMP1 (Figure 2D). 17 DC4 (DBLα1.1/4-CIDRα-6-DBLβ3) 18 DC8 (DBLα2-CIDRα1.1-DBLβ12-DBLγ4/6), and DC13 (DBLα1.7 and CIDRα1.4) 8 are the DCs studied most extensively.

Most PfEMP1 appear to be elongated and are rigid molecules with a zigzag shape and a length of about 30 nm, 19,20 although VAR2CSA-type PfEMP1 assume a more compact and globular shape with a diameter of approximately 20 nm. 21,22

2.2 | PfEMP1 function

The primary function of the PfEMP1 proteins is to mediate adhesion of IEs to host receptors in the vasculature. 17,23 This sequestration is vital to the parasites, as it allows mature IEs (misshapen and rigid because of the parasites growing inside them) to avoid the spleen passage, where they would be filtered and destroyed. 24,25 A range of vascular surface proteins and carbohydrates can serve as IE adhesion receptors, including CD36, 26 intercellular adhesion molecule 1 (ICAM-1), 27 endothelial protein C receptor (EPCR), 28 oncofetal chon‐
droitin sulphate (CSA), 29,30 and ABO blood group antigens. 31 The expression of IE adhesion receptors varies between different vascular beds, and is often regulated by cytokines. 32,33

Many different PfEMP1 proteins appear to have specificity for the same receptor, and this to some extent corresponds to the structurally defined PfEMP1 groups and domain subclasses mentioned above. 8,15,16 Thus, subclasses of DBLβ1 domains found in Group A, B, and C PfEMP1 bind ICAM-1, 34,35,36 CIDRα1 domains from Group A and B/A PfEMP1 bind EPCR, 13,28 whereas Group B and C contain CIDRα2-6 domains that bind CD36 (Figure 2D). 36,37 Finally, Group E (VAR2CSA-type) PfEMP1, which appear responsible for IE accumulation in the placenta, have unique affinity for CSA. 30,38,39 It is not very surprising that several of these large, multidomain proteins contain domains that can simultaneously mediate adhesion to several host receptors. 14,35,40,41 Examples include PfEMP1 variants concurrently interacting with ICAM-1 and EPCR, 14,41 with ICAM-1 and CD36, 35 or variants capable of distinct receptor-ligand interactions with endothelial cells and uninfected erythrocytes. 42

2.3 | Clonal antigenic variation controlling PfEMP1 expression

Aside from an exception of unresolved biological significance, 42 only one PfEMP1 variant is expressed on the surface of a given IE at any given time, 44 but the expressed variant may change from one 48-h cycle to the next. The ability to switch from the expression of one PfEMP1 variant to another (called clonal antigenic variation) acts as a key to the pathogenicity of P. falciparum parasites and is a major de‐
terminant of the characteristic chronicity of untreated infections. 45

The genetic processes governing clonal antigenic variation in P. fal‐
ciparum parasites are complex, and will not be described here, as they have been recently reviewed in detail elsewhere. 46 However, the switching pattern does not appear to be fixed, but rather seems
2.4 | PfEMP1 expression on the infected erythrocyte surface

The expression of PfEMP1 on the IE surface is confined to membrane protrusions called knobs.\textsuperscript{51} Formation of knobs involves multiple host and parasite molecules in addition to PfEMP1, such as the parasite-encoded knob-associated, histidine-rich protein (KAHRP) which multimerizes into a five-molecule spiral cone-like structure linked to erythrocyte cytoskeleton spectrin-ankyrin complexes (Figure 3).\textsuperscript{52-54}

The nascent PfEMP1 molecules are exported to the IE surface via a complex, multistep process that has recently been reviewed in detail elsewhere.\textsuperscript{55} Their ATS domains bind to the cytoskeleton via

---

\[ \text{Figure 2} \quad \text{var genes and PfEMP1 structure. (A), Modeled structure of PFD1235w DBLβ. The structure consists of subdomain 1 (S1, orange) with mixed helix-sheet structure and two helix bundles; subdomain 2 (S2 magenta) and subdomain 3 (S3 green). The ICAM-1 binding site of PFD1235w in S3 is indicated in red.}^{14,18} \ (B), \text{Group A and B var genes are located in subtelomeric regions of all chromosomes but are transcribed in opposite directions, whereas Group C var genes are found in central chromosomal regions.} \ (C), \text{PfEMP1 proteins are composed of different subtypes of DBL and CIDR domains. Groups B and C PfEMP1 predominantly have a four-domain structure, while larger PfEMP1 proteins have additional DBL domains following the first or second DBL-CIDR domains.} \ (D), \text{Plasmodium falciparum genomes encode tandem domain cassettes (DC) that are linked to different known adhesion phenotypes as indicated. DC8 is a chimeric gene between a group A and a group B var gene. Redrawn and modified from Hviid and Jensen}^{17} \]
LyMP (a member of the PHIST [Plasmodium helical interspersed subtelomeric] protein family [Figure 3]).53,56,57 Recent data suggest that the exported PfEMP1 molecules are delivered to the IE membrane away from the knobs, and then moved laterally and assembled into the knobs.54

The knobs appear on the IE surface approximately 16 hours postinvasion, peak in density about 20 hours later, followed by a slight decrease in density toward the end of the intraerythrocytic 48-h cycle.6 Each knob appears to accommodate less than a handful of PfEMP1 molecules, which are expressed in a cluster near the tip of the knob.58 The reason for the clustered and knob-confined display of PfEMP1 on the IE surface is not fully understood. However, it likely includes optimization for adhesion, as the surface knob density appears to vary with the PfEMP1 expressed and may be modulated by immunity, and because knob-less IEs generally have reduced PfEMP1 expression and do not adhere well.8,59-61 Consistent with that, disruption of the gene encoding KAHRP leads to disappearance of knobs, decreased PfEMP1 expression, and reduced IE adhesive-ness.62-64 Disruption of the gene encoding LyMP causes a similar decrease in IE adhesiveness, although expression of both PfEMP1 and knobs remain at wildtype levels.57

Altogether, these findings suggest that the overall IE adhesive-ness is the net result of which PfEMP1 is expressed (quality), how much of it is expressed (quantity), and how it is expressed (topology). This conclusion is supported by studies linking the protective effects of haemoglobinopathies such as HbC, Hbs, and α-thalassemia to an impaired ability of \textit{P. falciparum} parasites to adequately remodel the erythrocyte cytoskeleton and display PfEMP1 in these aberrant erythrocytes.65-68

3 | CEREBRAL MALARIA

Cerebral \textit{P. falciparum} malaria (CM) is defined by the World Health Organization (WHO) as deep and unarousable coma that persists for more than 1 hour after a seizure, irrespective of anticonvulsant medication, in a patient with peripheral \textit{P. falciparum} parasitemia and without another cause of encephalopathy.69 It is estimated that around 1% of children infected with \textit{P. falciparum} develop CM, which has a very high mortality despite treatment, with up to 75% of deaths occurring within the first 24 hours postadmission.70-72 CM is a leading cause of the estimated >400 000 deaths due to malaria each year73 despite the fact that this clinical definition of CM may lead to misclassification in as many as one in four cases.74,75

In sub-Saharan Africa, CM mostly affects children under 5 years of age who have only partial acquired immunity to \textit{P. falciparum} infection. Several studies have shown seasonal and transmission intensity-dependent differences in the frequency of CM,76,77 suggesting that the level of acquired immunity is an important determinant of CM susceptibility.78 This is supported by the finding that CM is mainly seen among older children and adults in Southeast Asia, where malaria transmission is less intense than in sub-Saharan Africa.79

There are significant differences in the pattern of vital organ dysfunction between African children and Southeast Asian adults with CM.70,80-82 In children, CM coincides with a period of rapid brain growth and physiologic changes of the blood-brain-barrier (BBB) that may account for some of these differences.82 Although CM in children generally has lower mortality than in adults under otherwise comparable conditions, seizures are more frequent and rapid, and CM in children is more often associated with anemia and neurocognitive sequelae.83,84 Retinal vessel changes, ring hemorrhages, and accumulation of inflammatory cells in the brain microvasculature are also more frequent, and hypoglycemia is one of the most common concomitant complication in pediatric CM cases.83,85

In the following sections, we will focus on CM as it occurs among children in Africa because this is the population most affected, and also because most of the available knowledge about CM stems from studies of African children (Box 1).

3.1 | Subtypes of CM

Retinopathy, characterized by retinal hemorrhages, papilledema, retinal whitening, and vessel color changes, is the most specific clinical diagnostic sign of CM,86-88 because it directly reflects the cerebral sequestration of IEs and the pathological processes occurring in the brain.86-90 It thus allows distinction between children with coma caused by cerebral IE sequestration and those, whose coma has other causes but fulfil the abovementioned WHO criteria for CM. The number of retinal hemorrhages prior to death correlates with the density of hemorrhages in the brain at mortem.91 Retinopathy has therefore been suggested to reflect the spectrum of CM severity86,92,93 as patients with retinopathy have higher mortality than those without.94,95
Box 1 Animal models of Plasmodium falciparum malaria

A detailed analysis of the intracerebral pathogenesis and pathology in CM comes primarily from postmortem studies. For this reason, infections of various strains of mice with P. berghei ANKA (PbA) is a popular in vivo model to dissect the mechanisms of CM. PbA infection can induce a condition known as experimental cerebral malaria (ECM), which recapitulates some—but certainly not all—features of CM. This has led to considerable scepticism regarding the utility of this murine model.\(^{364,365}\)

In both humans and mice, the clinical signs of CM progress from seizures, ataxia, and paralysis to coma and eventually death.\(^{366}\) However, although cerebral IE sequestration is a prominent feature of CM,\(^{364}\) it is a minor feature of ECM.\(^{367,368}\) Intravascular accumulation of platelets and immune cells has been observed in ECM, and CD8\(^+\) cells appear central to ECM pathogenesis.\(^{369-372}\) The picture is less clear in CM, where some studies have reported infiltration of leukocytes and platelets within the brain microvasculature,\(^{96,173}\) whereas others did not find that.\(^{195,373}\) Intravascular accumulation of monocytes has also been reported, but there is little evidence of high numbers of CD8\(^+\) T-cell accumulation in human CM.

ECM is characterized by a prominent pro-inflammatory cytokine response with high levels of IFN-\(\gamma\) and TNF-\(\alpha\), which results in upregulation of activation markers including ICAM-1, VCAM-1, and E-selectin.\(^{374}\) Although TNF-\(\alpha\)-deficient mice are also susceptible to PbA-induced ECM,\(^{375}\) ICAM-1 must be present for ECM to develop.\(^{376}\) Although inflammatory changes in the brain are lower in CM, elevated concentrations of circulating pro-inflammatory cytokines are characteristic.\(^{377,378}\) This contributes to a marked increase in the expression of endothelial cell adhesion molecules,\(^{106,144}\) and IEs and ICAM-1 co-localize in cerebral vessels postmortem.\(^ {102}\) Of particular relevance to the present text, none of the genomes of rodent malaria parasite and host factors that have been implicated in CM pathology can affect the permeability of the BBB. These include hemozoin-induced matrix metalloproteases (MMP), angiopoietins, sphingosine-1-phosphate, nitric oxide, platelet-activating factor, several cytokines (IL-1\(\alpha\), IL-1\(\beta\), IL-6, TNF\(\alpha\)), and a number of other factors.\(^ {91,111-117}\) As an example, MMP targets structural proteins of the basal lamina (fibronecint, laminin, heparan sulfate) and tight junction proteins (ZO-1, ZO-2, Claudin-5), which is known to cause breakdown of tight junctions, increased paracellular leak, and opening of the BBB during ischemic and inflammatory insults. Another protein, histidine-rich protein-2 (HRP-2) that is released at the time of schizont rupture,\(^ {118}\) can activate the innate immune system via NLRP3 inflammasome activation. The ensuing compromising of tight junction integrity and IL-1\(\beta\)- and MyD88-dependent increased vascular permeability has been proposed to promote CM pathogenesis.\(^ {119}\) In support of this, HRP-2 has been shown to line the endothelial walls of blood vessels,\(^ {120}\) particularly in retinopathy-positive CM patients.\(^ {121}\) Once the BBB is disrupted, leukocytes, cytokines, chemokines, and soluble parasite products may enter the brain parenchyma to activate the microglia and damage astrocytes and neurons, causing neuro-inflammation and coma.\(^ {122}\)
3.3 | Endothelial activation

Endothelial inflammation is a characteristic feature of *P. falciparum* malaria and correlates with disease severity in general and CM in particular. The inflammation may be induced directly by IEs adhering to the endothelium, or indirectly by inflammatory host and parasite products (such as IE membrane components, HRP-2, etc. However, activation may also occur independent of IEs as there is also evidence of generalized endothelial inflammation at sites devoid of IE sequestration.

Endothelial cells derived from Malawian children with CM have been shown to express particularly high levels of parasite and platelet receptors, to produce many endothelial microvesicles, release high levels of pro-inflammatory cytokines (including TNF-α and IFN-γ), and to be highly prone to undergo apoptosis. It seems likely that IEs may be involved, as they can induce apoptosis in primary brain endothelial cells, including cells from the brain, and cellular apoptosis has been suggested to cause increased endothelial permeability.

Activated brain endothelial cells are known to express high levels of a number of potential IE receptors (ie, ICAM-1, VCAM-1, P-selectin, and E-selectin),-exocytose Weibel-Palade bodies, release microvesicles, vascular endothelial growth factor (VEGF), and soluble cell adhesion molecules (ie, sICAM-1), and to show breakdown of tight junctions. Three bioactive molecules are released from the Weibel-Palade bodies, P-selectin (recruiting leukocytes), von Willebrand Factor (vWF; binding platelets), and Angiopoetin (Ang)-2. Ang-1 and Ang-2 are critical soluble regulators of endothelial activation and integrity, and levels of Ang-1 and Ang-2 have been described as reliable biomarkers of CM. Ang-2 is a vessel-stabilizing molecule that increases vascular permeability and facilitates endothelial activation by counteracting the action of Ang-1 by displacing Ang-1 from the receptor.

As mentioned above, activated brain endothelial cells are known to express high levels of Ang-1 and Ang-2, which are critical soluble regulators of endothelial activation and integrity. Levels of Ang-1 and Ang-2 have been described as reliable biomarkers of CM. Ang-2 is a vessel-stabilizing molecule that increases vascular permeability and facilitates endothelial activation by counteracting the action of Ang-1 by displacing Ang-1 from the receptor. It thus acts as a promoter of endothelial cell quiescence and survival. Release of Ang-2 from Weibel-Palade bodies increases the Ang-2/Ang-1 ratio and thus endothelial responsiveness. Increased concentrations of Ang-2 with decreased levels of Ang-1 has been associated with development of severe malaria in several studies, and children with retinopathy-positive CM have higher levels of Ang-2, Ang-2/Ang-1, soluble Tie-2, von Willebrand Factor, VEGF, and sICAM-1, and lower levels of Ang-1, compared to CM patients without retinopathy.

Both Ang-1 and Ang-2 are regulated by nitric oxide (NO), which is produced in the endothelium from L-arginine. NO causes vasorelaxation, downregulation of endothelial receptors, and reduces thrombosis. The bioavailability of NO is reduced in CM and this has been associated with fatal outcome. Low NO stimulates Weibel-Palade body exocytosis and activation of endothelium, with increased Ang-2 release from endothelial cells and expression of ICAM-1 and VCAM-1. Impaired NO production thus disrupts the ang-1/Tie-2-dependent signaling that maintains endothelial cell quiescence and vascular integrity.

This in turn promotes enhanced endothelial cell activation and cytoadhesion of IEs. All this notwithstanding, inhalation of NO was not found to reduce neurological defects or mortality in children with CM.

von Willebrand Factor is synthesized by the endothelial cells, and some of the synthesized vWF is constitutively secreted into plasma, but most is stored within Weibel-Palade bodies and only secreted following activation of the endothelial cell. vWF, particularly as large multimers, show enhanced binding to platelets and efficiently modulates aggregation of platelets. IEs can bind to platelets via P-selectin, C1q receptors, and thrombospondin receptor (CD36), leading to formation of IE/platelet clumps. The significance of this is indicated by the observation that children who died of CM had more platelet build-up in cerebral vessels than those dying of severe malarial anemia or non-malarial encephalopathy. Platelet accumulation was particularly prominent at sites of IE sequestration. Platelet-mediated IE clumping is thus likely to aggravate microvascular obstruction in CM, and release of tumor growth factor β from platelet granules may further cause apoptosis of brain endothelial cells.

In addition, accumulation of platelets may enable transfer of CD36 to endothelial cells, which normally expresses little or no CD36. Overall, the marked increase in plasma vWF levels in patients with severe malaria is likely to contribute to severe malaria pathogenesis. Finally, activation of endothelium leads to increased shedding of microvesicles from the plasma membrane of cells. Endothelial microvesicles have been found in very high concentrations in children with CM, and their levels correlate with disease severity.

It has been proposed that these vesicles may contribute to excessive T-cell activation and the immune pathogenesis of CM, as they express the molecules required for antigen presentation and T-cell stimulation, such as β2-microglobulin, MHC-II, CD40, and ICOSL. In addition, increased concentrations of non-endothelial microvesicles have been observed in CM, where they may also contribute to pathogenesis. Thus, the number of platelet-derived microvesicles correlates with the depth of the coma and thrombocytopenia, and extracellular IE-derived vesicles containing PFEMP1 (see below) can induce pro-inflammatory cytokines in human primary monocytes.

4 | PFEMP1 AND PATHOGENESIS OF CM

As mentioned above, *P. falciparum* parasites display PFEMP1 molecules on the surface of the erythrocytes they infect. From about 16 hours postinvasion, these high-molecular weight variant parasite proteins efficiently mediate adhesion of the IEs to a range of host receptors, and this is the reason why only young, ring-stage IEs are present in the peripheral circulation. It has long been speculated that PFEMP1-mediated IE adhesion to specific
receptors in key tissues and organs is an important determinant of clinical outcomes of *P. falciparum* infection. This hypothesis has been confirmed in the case of placental malaria, where the selective accumulation of IEs in the intervillous space is mediated by VAR2CSA-type PfEMP1 with affinity for placental CSA. It is furthermore well established that protective immunity to placental malaria depends on acquisition of IgG to CSA-adhering and VAR2CSA-expressing IEs. These findings have raised the hope that other specific PfEMP1 variants and host receptors may play similar and decisive roles in other forms of severe *P. falciparum* malaria, not least CM.

### 4.1 Sequestration of infected erythrocytes in the brain

IEs adhere to host endothelial receptors in the postcapillary venules of a number of organs, such as lungs, liver, intestine, brain, and the placenta. This tissue-specific sequestration causes circulatory disturbances and inflammation, and single- and multi-organ pathology such as renal, liver, lung, and placental dysfunction, and CM. Sequestration probably evolved to allow mature IEs, deformed by the parasites inside, to avoid destruction in the spleen. Identification of the parasite ligands, not least the specific PfEMP1 variants, which mediate IE sequestration in particular tissues—and the host receptors they bind to, has thus been a long-standing and important research priority. CM research is no exception.

Many studies have reported links between severe malaria (including CM) in children and infection with parasites transcribing var genes encoding PfEMP1 proteins from Group A and B/A. Other studies have narrowed the list of candidate genes to those having specific sequence signatures and/or encoding PfEMP1 variants with well-defined receptor specificity. ICAM-1 (CD54) was recognized as an endothelial IE receptor early on, and it has long been suspected to be important for the selective accumulation of IEs in the brain of CM patients. In line with this, contact with IEs can lead to endothelial upregulation of ICAM-1. Furthermore, it has been reported that *P. falciparum* parasites isolated from African children with CM bind preferentially to ICAM-1 in vitro. However, the opposite has also been reported, and isolates from Asian adult malaria patients do not appear to show preferential adhesion to ICAM-1. Finally, some studies have failed to find evidence of high ICAM-1 expression in the brains of fatal CM victims. Taken together, a complex picture regarding the role of ICAM-1 is emerging, although most of these data suggest that CM in African children is quite different from CM in Asian adults.

IE adhesion to ICAM-1 is mediated by PfEMP1 variants that can also bind to either EPCR or CD36. The former of these groups, exemplified by the *P. falciparum* 3D7 PfEMP1 PFD1235w, shows a clear association specifically with CM and contains the domain cassette DC4 (Figure 2D). The DC4 family was originally identified by a search for orthologs of the *pfds* gene in parasites from Ghanaian malaria patients, inspired by the link between **PFD1235w** and severe malaria. The search resulted in a panel of genetically distinct parasites binding ICAM-1 via the DBLβ3 domain of the DC4-type PfEMP1 expressed on the IE surface. Sequence analysis of these domains identified a C-terminal ICAM-1-binding motif ([W/L]xx[N/E]GG[P/A]xYx3H). The motif, which is also present in some Group A PfEMP1 proteins outside DC4 (including some DC5- and DC13-containing variants) and in a few Group B/A variants, is restricted to DBLβ domains located immediately downstream of CIDR domains of the EPCR-binding subtype.

Endothelial protein C receptor is the cognate receptor for PfEMP1 proteins containing domain cassette DC8 or DC13. DC13 is found among group A PfEMP1, whereas DC8 is found in Group B PfEMP1, and has evolved by recombination of ancestral Group A and B var genes. PfEMP1 variants containing DC8 or DC13 are common, and bind avidly to endothelial cells of lung, heart, and bone marrow. DC8-containing PfEMP1 proteins tend to be among the first expressed in early childhood infections, indicating that they possess adhesion properties that confer a survival advantage to IEs in malaria-naïve children. In addition, *P. falciparum* parasites obtained from African children and Indian adults with severe malaria—including CM—transcribe DC8- and DC13-encoding var genes at high levels. Their relevance to CM pathogenesis is further indicated by studies showing that IEs selected for adhesion to brain endothelial cells preferentially express these domain cassettes. Finally, expression of EPCR-binding PfEMP1 variants from Group A have been linked to brain swelling, which is a major contributor to mortality in pediatric CM. The available evidence linking the EPCR-adhering IE phenotype to severe malaria in general, and to CM in particular, is not completely unequivocal.

As an example, a study of Kenyan children with CM did not find evidence supporting particular enrichment of DC8- or DC13-containing PfEMP1 variants in children with retinopathy, a well-established indicator of CM, despite finding the expected association between CM and transcription of var genes encoding Group A PfEMP1. As mentioned above, not all PfEMP1 variants capable of binding ICAM-1 also bind EPCR. Indeed, all but one of the ICAM-1-binding DBLβ3 domains identified prior to the discovery of DC4 in Group A were found in Group B and Group C proteins. Those PfEMP1 proteins appear to be under dual selection for adhesion to ICAM-1 and CD36, as they all contain a CD36-binding CIDRβ domain upstream of the ICAM-1-binding DBLβ3 domain. This is not surprising, as CD36 is a very common IE adhesion receptor and most non-placental *P. falciparum* isolates can bind to it. Affinity for CD36 is a feature of the majority of Group B and C PfEMP1 proteins, but is not found in the Group A and B/A PfEMP1 that dominate in severe infections and in individuals with limited malaria immunity. Rather, CD36 binding is associated with uncomplicated malaria, and appears to have evolved to mediate IE sequestration in tissues other than the brain, where CD36 is absent or only sparsely present.

Combining the above evidence, the adhesion phenotype that is most clearly related to CM is IEs expressing Group A PfEMP1.
proteins (including DC4) that allow concomitant binding to both ICAM-1 and EPCR (“double binders”). The association between severe disease (and in particular, CM) and IE affinity for either of these receptors alone is less clear. It is worth noting in that context that the EPCR-binding CIDRα1 domain in DC8, and in some DC13, is not followed by an ICAM-1-binding DBL domain. Finally, although several molecules other than those already mentioned have been implicated as IE adhesion receptors, including some that appear to be expressed on brain endothelium, none of them have been linked to disease severity.

4.2 | Rosetting and clumping

Infected erythrocytes do not only adhere to the endothelium, but also to surrounding uninfected or infected erythrocytes. The former type of such aggregates, called rosettes, were first reported in the *P. falciparum* infected monkeys. The finding was quickly followed by studies demonstrating that the same phenotype was present in *P. falciparum*, and it appears that most species of malarial parasites are capable of inducing rosettes, which is a complex phenotype involving multiple parasite and host molecules. Thus, several erythrocyte molecules, including complement receptor 1, heparan sulphate, and the ABO blood group antigens, appear to be involved as receptors. On the parasite side, several ligands have been implicated, including members of several variant surface antigen families.

Infected erythrocytes isolated from patients with CM have been reported to form rosettes at significantly higher rates than IEs from patients with uncomplicated malaria. This difference might contribute to occlusion of cerebral microvessels by rosettes, to cerebral sequestration of IEs that express PfEMP1 variants that can bind to host receptors shared by cerebral microvessels and erythrocytes, and/or expressing PfEMP1 variants that allow binding to separate receptors on endothelium and on erythrocytes, respectively. With respect to the first of these possibilities, microvascular flow rates appear to be an important determinant for the clinical relationship.

**4.3 | The role of blood flow on IE adhesion**

Erythrocytes normally flow down the central line of a blood vessel, but the deformation and enhanced stiffness of IEs cause them to marginate, thus bringing them into contact with adhesion receptors on the endothelial surface. The distribution of cells, including IEs, in the blood stream is furthermore dependent on variables such as vessel diameter and plasma viscosity and flow rates. Flow not only affects margination of circulating cells, but can also lead to upregulation of endothelial receptors and cytokines in response to changes in shear stress. This is particularly true for ICAM-1, VCAM-1, and IL-1α. Endothelial integrins are also sensitive to changes in blood flow and become activated in response to increased shear stress. Blood flow is thus an important parameter to consider in studies of IE adhesion. A variety of in vitro assays have yielded important insights in this regard. As an example, CD36-specific adhesion of normal and ovalocytic IEs were similar in static assays, but were markedly different in assays conducted under physiologically more plausible flow conditions. With respect to CM-relevant and PfEMP1-specific adhesion, flow-based studies of adhesion of IEs expressing PfEMP1 variants that can mediate binding to both ICAM-1 and EPCR have revealed synergies that could not be discerned in static assays.
4.4 Converging on the protein C pathway

In spite of the very significant morbidity and high mortality of cerebral Plasmodium falciparum malaria, the pathophysiology of the disease is only partly understood.104,191,299 A range of potential and non-exclusive pathogenic mechanisms has been proposed, such as circulatory obstruction by sequestered IEs, imbalanced cytokine responses, and endothelial dysfunction and loss of BBB integrity.105,107,108,300 The available evidence is slowly converging on a scenario where CM is the consequence of the impact of IEs expressing particular PfEMP1 variants on the protein C-dependent maintenance of the integrity of brain endothelium.17,301

The protein C pathway is a crucial anti-coagulant and anti-inflammatory regulator of thrombin production during clot formation.302 Normally (Figure 4), thrombomodulin on the endothelial surface binds thrombin and activates protein C to become activated protein C (APC)

![Image](https://via.placeholder.com/150)

**FIGURE 4** Linking the protein C pathway with EPCR- and ICAM-1-binding IEs in cerebral malaria. (A) Effects of EPCR in the absence of Plasmodium falciparum-IE. Thrombin (Thr) is produced by the interaction between tissue factor (TF) and circulating activated factor VII (VIIa) (1). Thrombin initiates the EPCR- and thrombomodulin (TM)-facilitated activation of protein C (APC) that then inhibits thrombin production (2). APC uses EPCR as a coreceptor for cleavage of proteinase-activated receptor 1 (PAR-1). The EPCR-APC activation of PAR-1 inhibits the nuclear factor-κB pathway and exerts anti-inflammatory and anti-apoptotic activity (3). NF-κB production leads to enhancement of tight junctions and protection of endothelial barrier integrity (4). Angiopoetin-1 (Ang1) produced in response to the APC-PAR-1 interaction decreases Weibel-Palade body (WPB) exocytosis by occupying Tie2 (5). (B) The impact of infected erythrocytes expressing EPCR-bound PfEMP1 on the surface. The IE-EPCR interaction activates endothelial cells to release pro-inflammatory cytokines (IL-1, TNFα) that induce shedding of EPCR and TM from the endothelial surface and increases expression of ICAM-1 (6). The EPCR-IE interaction results in reduced levels of APC and increased thrombin generation with fibrin deposition (7). Increased levels of thrombin shift the PAR-1 response toward activation of the RhoA and NFκB with increased surface expression of ICAM-1 on the endothelial cell (8). The shift in the PAR-1 response inhibits S1P release resulting in loss of tight junctions, and compromises endothelial barrier function by causing localized vascular leaks (9). A reduction in Ang-1 levels increases WPB exocytosis via Tie2 and production of von Willebrand Factor (vWF) and Ang2 (10). Increased levels of Ang-2 further increase WPB exocytosis and contribute to the loss of endothelial barrier integrity and leakage (11). Platelets become activated by thrombin and cytokines, which leads to production of platelet microvesicles (12). Thrombin and activated platelets combine to form thrombi (13). Strings of vWF and activated platelets form complexes, which like thrombi impair the cerebral circulation. (C) The increase in ICAM-1 (in panel B) allows IE expressing PfEMP1 with a shared DBLβ1 ICAM-1 motif to adhere to the brain endothelium. A large proportion of the ICAM-1-adhering IEs might initially bind EPCR via their CIDRα1 domains

in a process that is strongly promoted by EPCR (also known as activated protein C receptor).301 The binding of APC to EPCR inhibits endothelial activation and TNFα-dependent inflammation, thereby limiting the opportunity for IE sequestration mediated by known PfEMP1 receptors such as ICAM-1, VCAM-1, E-selectin, and thrombospondin-1.303 The interaction also activates PAR-1, which has an anti-apoptotic effect that protects the endothelial barrier integrity.302,304 It has been proposed that this delicate system of checks-and-balances may be upset by IEs adhering to EPCR, thereby preventing physiologically appropriate activation of protein C.301,305,306 The PfEMP1 proteins expressed by EPCR-adhering IEs bind EPCR near/at the site where protein C/APC normally binds. The IEs might thereby interfere with binding of the normal ligand and compromise protective APC-dependent maintenance of the BBB via PAR-1.13,307,308 The result would be excessive endothelial inflammation, thrombin activation, fibrin cross-linking, platelet activation, upregulation of ICAM-1
expression, downregulation of EPCR, and endothelial leakage. This corresponds to the petechial lesions, fibrin clots, EPCR denuding, thrombomodulin deficiency, axonal injury, and brain swelling that have been reported in pathology studies of the brains of patients who died of CM.82,301,309-315 The lack of APC may furthermore allow induction of endothelial dysfunction via parasite and host soluble factors, such as histones, heme, and HRP-2, released locally when IEs rupture.316,317

Brain microvascular endothelium may be particularly susceptible to the disruption of the protein C pathway by EPCR-adhering IEs because EPCR is expressed at low levels at this site as opposed to the high expression seen in arteries and veins.318,319 CM is associated with loss of EPCR in the brain, and increased levels of soluble EPCR have been reported and associated with CM mortality.301,320 Human genetic variability affects the level of soluble EPCR, and there is some indication that some variants may be associated with protection from severe malaria, including CM, conceivably by neutralizing IE adhesion to EPCR.322 However, other studies did not detect such associations.233,234 Substantial variation also exists in genetic sequence of the CIDR1α domains in PFEMP1 that mediate binding to EPCR, although the EPCR-binding surface is largely conserved despite this variation.13 The variation nevertheless appears to impart differences in binding affinity that may affect how IE binding impacts normal EPCR function. The CIDR1.1 domains in DC8 thus affected APC and thrombin-induced permeability less than the CIDR1.4 domains of DC13.307,321 It is plausible that such diversity might contribute to the divergent pathophysiology of CM1 and CM2, where CM1 is characterized solely by IE sequestration, whereas CM2 also involves activation of coagulation and formation of fibrin clots and ring hemorrhages.75,82,96

The impact of EPCR-binding IEs on the protein C pathway proposed above is consistent with features of CM pathogenesis, but most studies have found this IE adhesion phenotype to be associated with severe malaria in general, rather than with CM specifically.209,215-217,226,227 However, this lack of specificity can be explained if CM pathogenesis requires parasites that express PFEMP1 variants that bind both to EPCR and ICAM-1 (the above-mentioned "double binders"). Our hypothesis involves a pathogenic cascade (Figure 4), where IEs initially adhere to EPCR on non-inflamed cerebral endothelium.14,17 This activates the endothelial cells as described above, induces their release of pro-inflammatory cytokines, and increases their expression of ICAM-1.110,219,301 "Double binders" may directly exploit this inflammatory response by adhering to ICAM-1, and this has been associated with disruption of the BBB.132 In contrast, erythrocytes infected by parasites expressing PFEMP1 variants that only have affinity for EPCR are likely to dislodge, as EPCR is shed as a part of the inflammatory response.301

5 | PFEMP1 AND IMMUNITY TO CM

In areas with stable transmission of *P. falciparum* parasites, susceptibility to clinical malaria is inversely correlated with age. Antibodies to parasite antigens on the surface of IEs are important, even decisive, determinants of this relationship, including the gradually decreasing risk of developing severe malaria such as CM.322-325 Acquisition of this type of immunity following natural parasite exposure is remarkably slow, incomplete, and temporally unstable, characteristics that all point to variant antigens, and in particular to PFEMP1 as the primary antigenic target.17,326

5.1 Naturally acquired, PFEMP1-specific immunity

The variant-specific, PFEMP1-centric hypothesis of susceptibility to, and acquired immunological protection against, *P. falciparum* malaria hinges on the idea that the infecting parasites adapt to pre-existing and developing immunity by switching to variants that are not recognized by specific antibodies.327,328 Severe disease ensues when those variants enable IEs to adhere to receptors that are widespread, allow strong IE adhesion, are expressed in critical tissues, and/or have vital functions. This is more likely to occur in individuals with little or no acquired immunity, and to involve variants that are relatively conserved among different parasite clones. As immunity to initially virulent variants is acquired, the parasites are steadily forced to express variants that are less virulent (more diverse, less likely to mediate firm IE adhesion, less likely to bind to widespread receptors, and more likely to be expressed in tissues where the consequences of IE sequestration are less serious).

Variant-specific immunity is indeed acquired in the orderly fashion predicted by the above theory. Antibodies to relatively conserved ("common") parasite antigens and protection from severe malaria are acquired first, followed by antibodies to more variant antigens associated with uncomplicated disease, and eventually by antibodies recognizing very diverse ("rare") antigens expressed on the surface of IEs obtained from carriers of asymptomatic/subclinical infections.329-332 Transcription of var genes and acquisition of PFEMP1-specific IgG follows this pattern, and shapes what PFEMP1 variants are compatible with parasite survival in a given semi-immune host.207,238,333 Thus, var gene transcription in children with limited pre-existing immunity and severe disease is skewed toward Group A,202,204,207 supporting the idea that these genes encode PFEMP1 proteins with adhesion specificities that are optimal for multiplication of the asexual parasites in non-immune hosts, and likely include binding properties that predispose to severe malaria. As children acquire Group A anti-PFEMP1 antibodies over successive infections, the proportion of var genes in Group B and C, encoding PFEMP1 variants associated with uncomplicated disease gradually increases.238,333-335 The clearest example of this type of relationship so far is the susceptibility to placental *P. falciparum* infection, and the acquisition of immunity to that particular form of severe malaria, as both depend on a particular type of PFEMP1 called VAR2CSA.187,188,336 However, there is every reason to assume that it applies to all forms of *P. falciparum* malaria, including CM.17

IgG antibodies to the Group A, DC4-containing PFEMP1 that can mediate IE adhesion to both ICAM-1 and EPCR are acquired early in life in areas with stable transmission of *P. falciparum*, and this acquisition is associated with protection from severe malaria, including
The same applies to IgG specific for the CIDR domains that mediate IE adhesion to EPCR,\textsuperscript{214,339,340} although divergent evidence also exists.\textsuperscript{342} A very recent study indicates that IgG to ICAM-1-binding PfEMP1 variants from Group B, which do not share the ICAM-1-binding motif with the Group A PfEMP1s (including DC4 variants) and that do not contain an EPCR-binding CIDR domain are acquired only when Group A ("dual binding")-specific immunity is already in place.\textsuperscript{342} This study thus further underpins the theory of a PfEMP1 hierarchy modulated by host immunity.

The degree of IgG cross-reactivity that transcends clone-specific variability in key antigens is of obvious importance. Encouragingly, naturally acquired antibodies capable of inhibiting the interaction between ICAM-1 and many ICAM-1-binding DBL\(\beta\) domain variants were recently observed.\textsuperscript{338} The prevalence of IgG specific for Group A ICAM-1-binding domains furthermore appears to be higher than those specific for EPCR-binding CIDR\(x1\) domains, suggesting that the former antibody specificity is more cross-reactive than the latter.\textsuperscript{341}

### 5.2 PfEMP1-based vaccination and other PfEMP1-specific interventions against CM

An ideal malaria vaccine would elicit immunity that prevents infection of humans and leads to transmission elimination and global eradication of \textit{P. falciparum}. While this goal is elusive at the present time, vaccines that prevent severe malaria illness, particularly in children and pregnant women, could constitute an alternative or interim strategy.

Current efforts to develop VAR2CSA-based vaccines to prevent placental malaria is the most advanced example of a PfEMP1-based approach to malaria vaccination, as several such vaccines are currently in clinical trials.\textsuperscript{343-347} However, it is conceivable that development of vaccines to prevent severe malaria in children, including CM, might be possible using a similar strategy.\textsuperscript{193} An obvious goal would be vaccines eliciting a broadly reactive antibody response preventing, and ideally reversing, adhesion of IEs to ICAM-1 and EPCR, which appear as key receptors in CM pathogenesis. Vaccines designed to specifically target "double binders" would seem particularly attractive. Blocking of EPCR-specific adhesion of IE by vaccination with relevant CIDR\(x1\) domains would lead to protection not only against CM, but also against severe childhood malaria in general. Unfortunately, EPCR-binding CIDR\(x1\) domains show substantial interclonal sequence variation and experimental antibodies binding distant variants are difficult to generate and are rarely inhibitory.\textsuperscript{348,349} This reduces the likelihood of natural induction of cross-inhibitory IgG responses targeting the critical parts of CIDR\(x1\) domains. In contrast, the highly conserved ICAM-1 binding site of these "dual binding" PfEMP1 variants encourage future efforts to raise broadly cross-reactive IgG antibody responses against such molecules using their ICAM-1 binding DBL\(\beta\) domain as part of a strategy to prevent death due to CM.\textsuperscript{14,338}

However, many questions remain unanswered. Aside from the uncertainty regarding the feasibility of making a vaccine with the desired qualities, it is unclear whether inhibition/reversal of ICAM-1/EPCR-specific IE adhesion would be sufficient to prevent CM. It is similarly unclear to what extent acquired clinical protection from CM requires neutralizing (IE adhesion-blocking) antibodies, as opposed to opsonizing antibodies leading to preferential phagocytic and/or cytotoxic removal of IEs expressing particularly pathogenic PfEMP1 variants.

While efforts to overcome the obstacles in the development of a PfEMP1-specific vaccine to protect against CM continue, other interventions should also be pursued. Unfortunately, adjunctive therapies (eg, glucocorticoids, anti-TNF\(\alpha\)s, iron chelators, osmotic regulators, anti-oxidants, and glycosaminoglycans), aimed at protecting against CM-related brain damage and neuronal injury, have so far been unsuccessful.\textsuperscript{72,350,351} Furthermore, current anti-malarial drugs fail to reverse adhesion of IEs.\textsuperscript{352} This notwithstanding, our improved understanding of molecular and cellular basis of CM pathogenesis might encourage novel adjunctive therapies aimed at dislodging sequestered parasite by interfering with PfEMP1-mediated IE adhesion. Along this line, soluble EPCR has been shown to inhibit adhesion of DC8-expressing IEs and endothelial cells in vitro.\textsuperscript{28} The finding of a soluble EPCR variant that binds PfEMP1 without affecting protein C binding to EPCR supports the in vivo feasibility of this approach,\textsuperscript{13} although the very low off-rates of the PfEMP1-EPCR interaction may not allow release of IEs already bound.\textsuperscript{308} Finally, a monoclonal antibody has been reported to inhibit and reverse adhesion of IEs to ICAM-1, including antibodies that affect "dual binding" PfEMP1 variants.\textsuperscript{353} However, the associated cost of this intervention will probably prevent its use in clinical practice.\textsuperscript{354}

### 6 CONCLUSIONS AND THE WAY FORWARD

Our understanding of the molecular basis of the interactions between \textit{P. falciparum} parasites and the humans they infect is progressing at a rapid pace. This is not least so for the tissue-, organ-, and receptor-specific adhesion of IEs that act as major contributors to the pathogenesis of \textit{P. falciparum} malaria, including the development of severe complications such as CM.

The critical importance of PfEMP1 for parasite survival makes these antigens major immune targets, and acquisition of PfEMP1-specific antibodies indeed appears to be a central component of naturally acquired protection. Emulating and accelerating these responses through vaccination would therefore seem obvious. However, the flipside is that the importance of PfEMP1 to the parasites applies a strong selective pressure on them to evolve mechanisms to evade protective, PfEMP1-specific immunity. The most prominent example of this is clonal antigenic variation, which undoubtedly evolved to delay and frustrate the acquisition of PfEMP1-specific protective antibodies, thereby enabling chronic infections.\textsuperscript{17,326,355-358} It is an important issue to ascertain precisely how, and at what level(s), this undermining operates. While it seems clear that interclonal variation in functionally important
antibody epitopes is involved, it is essentially unknown to what extent clonal antigenic PfEMP1 variation is affecting T-cell-dependent helper functions. Clonal antigenic variation apart, it is becoming increasingly clear that *P. falciparum* parasites have evolved a range of other strategies to evade PfEMP1-specific immunity. Examples are antigen topology and cloaking, interference with antigen presentation, and subversion of soluble host molecules.20,277,359

In conclusion, much has been learnt from years of research on CM pathogenesis and immunity, although a lot remains to be known. Because this malaria complication is such an important part of the intolerable obstacle that *P. falciparum* malaria constitutes to health and to economic progress and equality, we are obliged to continue this research.193,363

**ACKNOWLEDGEMENTS**

Research at the authors’ laboratories is supported by Augustinus Fonden, The Consultative Committee for Developmental Research (17-02-KU), Independent Research Fund Denmark (4183-00539 and 8020-00034B), Novo Nordisk Fonden (NNF16OC0022298), Hørslev Fonden, Aase og Ejnar Danielsen Fond (10-002129), Læge Sofus Carl Emil Friis og hustru Olga Doris Friis Legat, Svend Andersen Fonden and AP Møller Fonden (18-L-0040). Mette U. Madsen and Maiken H. Visti are acknowledged for their excellent technical assistance throughout the years.

**CONFLICTS OF INTEREST**

The authors declare no competing interests.

**ORCID**

Anja Ramstedt Jensen  [https://orcid.org/0000-0002-4004-7554](https://orcid.org/0000-0002-4004-7554)

Yvonne Adams  [https://orcid.org/0000-0001-8041-6580](https://orcid.org/0000-0001-8041-6580)

Lars Hviid  [https://orcid.org/0000-0002-1698-4927](https://orcid.org/0000-0002-1698-4927)

**REFERENCES**

24. Hommel M, David PH, Oligino LD. Surface alterations of erythrocytes in *Plasmodium falciparum* malaria. Antigenic


122. JENSEN ET AL.


183. Smith JD, Chitnis CE, Craig AG, et al. Switches in expression of Plasmodium falciparum var genes correlate with changes in


222. Ockenhouse CF, Ho M, Tandon NN, et al. Molecular basis of sequestration in severe and uncomplicated Plasmodium falciparum


260. McCormick CJ, Craig A, Roberts D, Newbold CI, Berendt AR. Intercellular adhesion molecule-1 and CD36 synergize to


281. Aarts PA, van den Broek SA, Prins GW, Kuiken GD, Datta JC. Blood platelets are concentrated near the wall and red blood cells, in the center in flowing blood. Arteriosclerosis. 1988;8:819-824.


297. Cortes A, Melloombo M, Mgone CS, Beck HP, Reeder JC, Cooke BM. Adhesion of *Plasmodium falciparum*-infected red blood cells to CD36 under flow is enhanced by the cerebral malaria-protective


306. Francischetti IM. Does activation of the blood coagulation cascade have a role in malaria pathogenesis? *Trends Parasitol.* 2008;24:258-263.


335. Duffy MF, Noviyanti R, Tsuboi T, et al. Differences in PfEMP1s recognize human erythrocyte membrane protein 1, VAR4, predict protection


How to cite this article: Jensen AR, Adams Y, Hviid L. Cerebral Plasmodium falciparum malaria: The role of PfEMP1 in its pathogenesis and immunity, and PfEMP1-based vaccines to prevent it. Immunol Rev. 2020;293:230-252. https://doi.org/10.1111/imr.12807