Mutant CEBPA directly drives the expression of the targetable tumor-promoting factor CD73 in AML

Jakobsen, Janus S; Laursen, Linea G; Schuster, Mikkel B; Pundhir, Sachin; Schoof, Erwin; Ge, Ying; d'Altri, Teresa; Vitting-Seerup, Kristoffer; Rapin, Nicolas; Gentil, Coline; Jendholm, Johan; Theilgaard-Mönch, Kim; Reckzeh, Kristian; Bullinger, Lars; Döhner, Konstanze; Hokland, Peter; Fitzgibbon, Jude; Porse, Bo T

Published in:
Science Advances

DOI:
10.1126/sciadv.aaw4304

Publication date:
2019

Document version
Publisher's PDF, also known as Version of record

Document license:
CC BY-NC

Citation for published version (APA):
CANCER

Mutant CEBPA directly drives the expression of the targetable tumor-promoting factor CD73 in AML

Janus S. Jakobsen1,2,3,†, Linea G. Laursen1,2,3,†, Mikkel B. Schuster1,2,3,‡, Sachin Pundhir1,2,3,4, Erwin Schoor1,2,3, Ying Ge1,2,3, Teresa d’Altri1,2,3, Kristoffer Vitting-Seerup4, Nicolas Rapin1,2,3,4,‡, Coline Gentili1,2,3, Johan Jendholm1,2,3, Kim Thelggaard-Mönch1,2,3,5, Kristian Reckzeh1,2,3, Lars Bullinger6, Konstanze Döhner7, Peter Hokland8, Jude Fitzgibbon9, Bo T. Porse1,2,3,§

The key myeloid transcription factor (TF), CEBPA, is frequently mutated in acute myeloid leukemia (AML), but the direct molecular effects of this leukemic driver mutation remain elusive. To investigate CEBPA mutant AML, we performed microscale, in vivo chromatin immunoprecipitation sequencing and identified a set of aberrantly activated enhancers, exclusively occupied by the leukemia-associated CEBPA-p30 isoform. Comparing gene expression changes in human CEBPA mutant AML and the corresponding Cebpa16p30 mouse model, we identified Ntse, encoding CD73, as a cross-species AML gene with an upstream leukemic enhancer physically and functionally linked to the gene. Increased expression of CD73, mediated by the CEBPA-p30 isoform, sustained leukemic growth via the CD73/A2AR axis. Notably, targeting of this pathway enhanced survival of AML-transplanted mice. Our data thus indicate a first-in-class link between a cancer driver mutation in a TF and a druggable, direct transcriptional target.

INTRODUCTION

Recent advances in our ability to sequence cancer genomes have left us with the challenge of translating this knowledge into tailor-made therapies targeting the characteristics of individual tumors, a concept frequently termed precision medicine. To harness the potential of precision medicine, we will need to identify oncogenic driver mutations and understand how they sustain disease development and maintenance.

One of the best characterized cancer types is acute myeloid leukemia (AML), a heterogeneous group of hematopoietic clonal disorders characterized by rapid accumulation of immature myeloid blasts at the expense of normal hematopoiesis. Specific cancer driver mutations, which underlie the distinct molecular AML subtypes, are closely linked to therapy outcome and overall risk stratification (1–3). Frequently found classes of leukemia drivers include fusion proteins and mutations in NPM1 [Nucleophosmin (encoded by NPM1)], epigenetic modifiers, signaling pathways, and the spliceosome, as well as loss-of-function mutations in key myeloid transcription factors (TFs) (1).

Despite our detailed knowledge of AML genetics, treatment has remained largely unchanged for decades and survival rates are low (3). Exceptions are novel therapies exploiting the genetic abnormalities of specific subgroups to target specific molecular events driving malignant transformation. An example is treatment of patients with acute promyelocytic leukemia (APL) with all-trans retinoic acid (ATRA), which overrides the differentiation block facilitated by the t(15:17) encoded PML-RARA fusion (4). Such an advancement emphasizes the importance of functionally characterizing the molecular pathway and ultimately leads to the development of an overt and transplantable AML (11).

CEBPA has both activating and repressive activities. Among the transcriptionally activated targets are the genes encoding the myeloid differentiation block in preleukemic animals. This condition subsequently progresses into myeloid hyperproliferation and ultimately leads to the development of a overt and transplantable AML. (11).

CEBPA has both activating and repressive activities. Among the transcriptionally activated targets are the genes encoding the myeloid-specifying G-CSF, GM-CSF, and MPO (12–14). Conversely, loss of CEBPA causes derepression of Cebpg (15) and Sox4 (16), both of which are crucial for maintaining leukemic cells in a dedifferentiated state. However, it is not understood how the specific loss of full-length p42, and hence, the preferential p30/p30 homodimer formation...
affect the gene regulatory landscape on a global level and ultimately drives leukemic progression.

In this study, we used a combination of gene expression and chromatin immunoprecipitation sequencing (ChIP-seq) analyses to decipher the direct regulatory function of p30 in AML. We compared highly purified populations of leukemic granulocytic monocytic progenitors (L-GMPs; Lin−cKit+Sca1+CD150−CD41−FcyRII/III−) from both leukemic Lp30 mice and patients with AML to their closest healthy counterpart, GMPs. Using this approach, we identified N5ε, encoding the ectoenzyme NT5E/CD73, as specifically up-regulated in biallelic CEBPA mutant leukemia as a consequence of p30 binding to a normally silenced upstream enhancer. CD73 catalyzes the conversion of adenosine monophosphate (AMP) into immune-dampening adenosine and has been shown to provide an immune evasive microenvironment in solid tumors. Correspondingly, an anti-tumor effect has previously been demonstrated upon blockade of the enzyme in cancers with CD73 expression. Here, we validated CD73 as a tumor-promoting factor in biallelic CEBPA mutant AML that can be targeted by dual inhibition of CD73 and adenosinergic signaling through the A2A receptor (A2AR), suggesting a novel precise treatment modality for this AML subtype.

RESULTS

CEBPA isoforms occupy specific genomic regions associated with distinct epigenetic profiles in normal and leukemic GMPs

To investigate the action of the cancer-driving CEBPA-p30 isoform, we performed ChIP-seq for CEBPA on phenotypically defined GMPs from wild-type (WT) and leukemic Lp30 mice isolated using fluorescence-activated cell sorting (FACS) (fig. S1A). In contrast to the Lp30 mouse model, solely expressing p30, WT GMPs almost exclusively express the full-length p42 isoform of CEBPA (11). Thus, for simplicity, we will refer to CEBPA in WT GMPs as p42, although, in addition to the predominant p42/p42 homodimer, these cells retain low levels of p42/p30 heterodimers and p30/p30 homodimers. CEBPA genomic occupancy (Fig. 1A) was used to identify and subdivide stringent sets of regions bound by CEBPA either exclusively in leukemic GMPs (p30 regions), in WT GMPs (p42 regions), or shared between both conditions (common regions) (total of 30,951) (Fig. 1B and data file S1); 87.7% of the CEBPA-bound regions are common, whereas 4.5 and 7.9% are p42 and p30 specific, respectively (Fig. 1, B and C). ChIP-seq of histone 3 lysine 4 monomethyl (H3K4me1) and histone 3 lysine 27 acetyl (H3K27ac) modifications were performed to assess the coverage level of these enhancer-associated marks in the three sets of CEBPA regions (Fig. 1A and fig. S1B). Quantification of H3K4me1 (associated with an open chromatin structure) and H3K27ac (associated with active regions) levels shows substantial decoration with both marks in the common regions and no difference between the Lp30 and WT conditions (Fig. 1, C and D). In comparison, p42-specific regions display an overall lower level of enhancer marks [P (H3K27ac common versus p42) = 2.9 × 10^{-151} and P (H3K4me1 common versus p42) = 4.6 × 10^{-164}] and, moreover, a significant drop in the H3K27ac: P < 0.05 by edgeR, see Materials and Methods) with higher (2342 genes) or lower (2469) expression in Lp30 L-GMPs compared to WT GMPs (Fig. 1F and data file S2). In line with a loss of transactivation activity as a consequence of the truncated trans-activation domain (TAD) in p30 (fig. S1C) (17, 18), genes associated with common regions are more frequently down-regulated (1600) than up-regulated (1191) when comparing the Lp30 L-GMPs to their GMP WT counterparts. For the p42 and p30 regions, gene expression changes are in accordance with the differences in overall enhancer histone mark levels at these regions. Hence, p30 regions are preferentially associated with down-regulated genes (308 versus 102 up-regulated), while p30 regions are more associated with up-regulated genes (359 versus 253 down-regulated), comparing the Lp30 L-GMPs to WT GMPs (Fig. 1F). Together, gene expression and histone mark pattern changes indicate that CEBPA predominantly acts as a transcriptional activator, both in the Lp30 and in the WT cells, in line with most previous studies on CEBPA in late granulopoiesis.

In conclusion, we identify genomic regions bound specifically by p30 in the leukemic condition. Binding of p30 at these regions leads to increases in enhancer mark levels and changes in gene expression, suggesting that p30 is initiating a unique transcriptional program, potentially influencing leukemogenesis.

Leukemia-specific CEBPA target enhancers are enriched for E-twenty-six (ETS) family motifs and low-affinity CEBPA sites

To elucidate the differential CEBPA binding and enhancer mark activity of the p30 and p42 isoforms, we examined the relative enrichment of known TF binding motifs at p42- and p30-specific regions using the common regions as a reference. TF motifs enriched in p30 regions compared to common regions are binding sites recognized by TFs belonging to either the CEBP or the ETS family (e.g., ELKs, ERG, ETSs, ETVs, GABP1, FEV, and FLI1) (Fig. 2A) (see data file S1 for the full list of motifs). Among the ETS factors, ERG, FLI1, and PU.1 belong to a core set of TFs previously shown to bind to a wide range of hematopoietic enhancers in the murine HPC-7 stem/progenitor cell line (19). Using these data, we demonstrated that binding of the ETS factors was selectively depleted at p42-specific regions, and ERG in particular displayed a high degree of overlap with common and p30-specific regions (Fig. 2B). Moreover, and in contrast to FLI1 and PU.1, global quantitative proteomics analysis demonstrated that ERG was selectively up-regulated in Lp30 L-GMPs compared to WT GMPs, hinting at a role for ERG in binding of p30 to the p30-specific regions (Fig. 2C and data file S3). Conversely, the sequences enriched at the p42 regions are binding sites for the basic region leucine zipper (bZip) domain–containing CEBPs, as well as the PAR bZip family members DBP, HLF, and TEF (Fig. 2A). Of these, several are expressed in WT GMPs, e.g., CEBPE, CEBPG, and
CEBPZ (protein), as well as Tef and Hlf (mRNA). Tef is up-regulated, while Hlf is strongly down-regulated in Lp30 L-GMPs (Fig. 2D and data file S2). Thus, some of the loss of CEBPA binding at p42-specific regions might be explained by the loss of cobinding with HLF, as this factor has some occupancy overlap with the p42-specific regions (fig. S2A). Of the CEBP family members, CEBPA itself was strongly up-regulated (Fig. 2, C and D, and data file S3), which is consistent with previous findings in both human and murine CEBPA mutant AML (11, 20). We validated these findings using quantitative reverse transcription polymerase chain reaction (RT-qPCR) (Cebpa; Fig. 2E) and Western blotting (CEBPA; Fig. 2F and fig. S2B).

To further probe the differential binding of the p30 and p42 CEBPA isoforms, we next compared the CEBPA motif scores (i.e., focusing only on bona fide CEBPA motifs) in each individual bound region with August 27, 2019 http://advances.sciencemag.org/ Downloaded from
and found that, compared to the common regions, p42 regions, on average, harbor better matching motifs, while p30 regions display a lower average match score (Fig. 2G). Similarly, when we assess the CEBPA motif score distribution, CEBPA motifs associated with p30-specific regions are shifted toward low-scoring CEBPA motifs, compared to common and p42-specific regions (Fig. 2H), with both high- and low-scoring motifs conforming to CEBPA motifs (fig. S2C). These findings suggest that the increased CEBPA levels in Lp30 L-GMPs allow the p30 isoform to associate with low-affinity CEBPA sites.

Overall, our data indicate that CEBPA-p30–specific binding in Lp30 L-GMPs occurs at already active (H3K27ac marked) or at least open (H3K4me1 marked) enhancers. These enhancers are normally not bound by CEBPA in GMPs but are associated with other TFs such as ERG. We propose that increased levels of CEBPA in the Lp30 cells promote the binding of p30 to the low-affinity CEBPA motifs, leading to increased expression of a subset of associated genes.

**Comparative analyses of murine and human CEBPA mutant AML pinpoints conserved transcriptional changes**

We next set out to identify deregulated genes shared between human AML and the corresponding Lp30 mouse model, i.e., the shared core p30 leukemic transcriptional program, to identify gene expression...
changes of potential clinical relevance. To this end, leukemic and normal GMPs from four patients with N/C-biallelic CEBPA mutant AML and two healthy volunteers, respectively, were isolated by FACs and subjected to RNA-seq (fig. S3A). Overall, fewer genes are deregulated (log2 fold change > 0.58, \( P < 0.05 \), edgeR, see Materials and Methods) in the human leukemic cells as compared to those of mice (data file S2), likely reflecting a higher level of biological variation in the human samples. More genes are down-regulated (2064) than up-regulated (1491) when comparing L-GMPs to normal GMPs (Fig. 3A).

Gene Ontology (GO) analyses revealed enrichment of many shared categories between human and mouse AML-associated gene expression changes (data file S4). Examples of categories significantly associated with up-regulated genes shared between the two species include “nucleosome core,” “focal adhesion,” “immunity,” “apoptosis,” and “negative regulation of cell proliferation.” Similarly, many categories were shared between human and mouse down-regulated genes, including “cytoskeleton,” “cell cycle,” “cell junction,” “focal adhesion,” “transcription,” and “DNA damage” (Fig. 3B and fig. S3B).

To pinpoint conserved changes of individual genes, we generated a stringently filtered shortlist of genes for which a direct human or mouse ortholog could be identified and counted genes with aligned up- or down-regulation [using a cutoff of false discovery rate (FDR) < 0.01 for both edgeR and DEseq] (data file S2). Of the 415 (human) and 1778 (mouse) shortlisted genes, 102 were found to be regulated in the same direction, which corresponds to threefold more than expected by random distribution (\( P < 0.0005 \), Fisher’s exact test) (Fig. 3C). Hence, these 102 genes constitute the core transcriptional program of CEBPA mutant AML and further demonstrate that the L30 mouse model mirrors key features of the corresponding human disease.

We next sorted the 102 genes associated with the CEBPA mutant transcriptional program based on expression fold changes in both species (Fig. 3D, left) and found that most were down-regulated (82 genes), while only 20 genes were up-regulated in Lp30 L-GMPs compared to normal GMPs. Notably, Gene Set Enrichment Analysis (GSEA) showed a strong overrepresentation of the 20 up-regulated genes and underrepresentation of the 82 down-regulated genes in biallelic CEBPA AML versus all other AMLs in the The Cancer Genome Atlas (TCGA) dataset (fig. S3C), supporting the notion of a subtype-specific core genetic program in human AML.

Normalizing expression changes individually for human and mouse samples (Fig. 3D, right), we found that only the top three genes, ARPP21/Arpp1, NT5E/Ni5e, and ITGAX/Itgax, were up-regulated from almost undetectable levels in healthy cells to high expression in leukemic cells derived from both species (data file S2). Of these, only NT5e was significantly up-regulated at the protein level (NT5E/CD73) (Fig. 3E), while ARPP21 was not found by the proteomics analysis and ITGAX was not up-regulated (Fig. 3E and data file S3).

In summary, we have defined cross-species conserved core expression changes induced by the CEBPA-p30 AML driver. Up-regulated genes, encoding potentially druggable proteins, were limited to a narrow set, in which NT5e is an interesting candidate.

**A leukemia-specific CEBPA-p30-bound enhancer controls NT5e transcription**

NT5e encodes NT5E, ecto-5’-nucleotidase (aka CD73), an ectoenzyme that catalyzes the rate-limiting step of AMP to adenosine conversion. CD73 facilitates progression in several cancer types via (i) evasion of anti-tumor immune responses (21, 22) and (ii) adenosine-mediated inhibition of apoptosis and promotion of proliferation of the tumor cells (23, 24). Our demonstration of the conserved up-regulation of CD73 in p30-driven AML is therefore intriguing, and we sought to explore the connection further.

First, we validated the AML-specific expression of NT5e mRNA and CD73 protein by independent methods in the Lp30 mouse model (Fig. 4, A and B). Having established a leukemia-specific expression of NT5e, we investigated whether the aberrant expression of CEBPA-p30 in leukemic cells directly affects NT5e levels. Scrutinizing the ChIP-seq data, we found that the p30-specific regions included a position 40 kb upstream of the NT5e transcription start site (TSS), displaying clear CEBPA binding and enhancer-associated marks (H3K4me1 and H3K27ac) exclusively in the Lp30 cells, indicative of a leukemia-specific enhancer (Fig. 4C). Notably, this putative regulatory region is not decorated by H3K4me1 or H3K27ac histone marks at the granulocyte stage, suggesting that the enhancer mark is not due to premature differentiation of Lp30 L-GMPs (fig. S4A).

To investigate whether the putative −40-kb enhancer could regulate NT5e expression, we first examined whether this region physically interacted with the NT5e TSS using chromosome conformation capture (3C) with qPCR. Of seven tested regions spanning 50 kb upstream of NT5e, we found two bordering the enhancer region interacting significantly with the TSS when comparing Lp30 L-GMPs to a c-kit-enriched bone marrow (BM) control population, consistent with a direct physical contact between the −40-kb region and the NT5e TSS (Fig. 4C, middle). We then applied CRISPR interference (CRISPRi) to assess the functional role of the −40-kb enhancer (25). To this end, we targeted two control positions (−112 and −15 kb, located distally from and in between the enhancer and TSS, respectively) and two enhancer positions, as well as the TSS with guide RNAs (gRNAs). This resulted in a significant reduction of NT5e expression for enhancer-directed gRNAs equivalent to TSS targeting, compared to the controls (Fig. 4C, bottom). In contrast, no neighboring genes were significantly affected (fig. S4B). Last, we cloned the −40-kb region to a luciferase reporter vector and observed a small but significant induction of expression upon cotransfection with either CEBPA-p42 or the mutant p30 isoform (Fig. 4D). This supports the region’s potential as a CEBPA targeted enhancer, activatable by either isoform when present at sufficient levels.

In combination, these data indicate that, in the Lp30 mouse model of CEBPA mutant AML, elevated levels of the p30 isoform directly drive expression of NT5e. This is accomplished via a 40-kb upstream enhancer active exclusively in the malignant cells.

**NT5e is overexpressed in CEBPA mutant AML patient samples and controlled by a CEBPA targeted enhancer**

To test the potential clinical relevance of our findings in the Lp30 mouse model, we next wanted to extend them to human AML. To this end, we first carried out RT-qPCR and confirmed the high expression of NT5E found in our human RNA-seq data along with elevated levels of CEBPA (Fig. 5, A and B). Moreover, using flow cytometric analysis, we observed increased frequencies of GMP-resembling cells within the CD3+ CD19− fraction (devoid of residual normal lymphocytes) of the CD73+ population in patients with AML with biallelic CEBPA mutations (fig. S5A). These findings were further corroborated by analysis of three publicly available AML gene expression datasets, all demonstrating the high expression of CEBPA and NT5E in CEBPA mutant AML compared to other normal karyotype subtypes (Fig. 5C). The difference between CEBPA mutant and
Fig. 3. Gene expression analysis of murine and human CEBPA mutant AML. (A) RNA-seq assessment of total up- and down-regulated transcripts in L-GMPs sorted from human AML samples with biallelic CEBPA mutations versus WT GMPs (log2 fold change > 0.58, \( P < 0.05 \) by edgeR, see Materials and Methods). (B) Overrepresented GO categories (David online) of deregulated genes in L-GMPs versus normal GMPs. Included are representative, nonredundant categories found among the top 12 (up) or 64 (down) enriched in either species. "Negative regulation of cell proliferation" was included in AML-up for comparison to the down-regulated genes "cell cycle" category enrichment. "Transcription" was included for AML-down as it was enriched above the shared "DNA damage" in humans. Shared categories are colored; species specific or 64 (down) enriched in either species. "Negative regulation of cell proliferation" was included in AML-up for comparison to the down-regulated genes "cell cycle" category.

(C) Stringently assessed deregulated mRNAs (FDR \( \leq 0.01 \)) in leukemic GMPs versus normal GMPs from human (teal) and murine (orange) BM. Lists of deregulated mRNAs were reduced to genes for which orthologs could be found in both species. Observed overlap (orthologs with shared directionality) (top) and expected random overlap (bottom) between the two species are indicated. (D) Quantified differential expression of the 102 overlapping genes shown in (C). Left: log2 fold change for each species. Right: Relative expression levels normalized for each species individually. (E) CD73 and ITGAX protein levels assessed by mass spectrometry (MS) (Limma multifactorial analysis, Benjamini-Hochberg corrected \( P \) value. *\( P \leq 0.05 \)).
Fig. 4. CEBPA-p30 mediated activation of Nt5e expression. (A) RT-qPCR quantification of Nt5e expression normalized to Actg1 (WT GMPs, n = 3; Lp30 L-GMPs, n = 4; mean ± SD, t test). (B) CD73 expression on the surface of WT GMPs and Lp30 L-GMPs as determined by flow cytometry (left) and the associated quantification CD73 expression GMPs (right; WT GMPs, n = 2; Lp30 L-GMPs, n = 3; mean ± SD, t test). (C) Top: ChIP-seq analysis on WT GMPs and Lp30 L-GMPs, showing a putative p30-specific enhancer located 40 kb upstream of the murine Nt5e gene. Middle: Assessment of direct interaction between the −40-kb enhancer and the Nt5e TSS by 3C-qPCR. Primer localization is depicted by open circles. qPCR (n = 3 to 5) was normalized to cKit–enriched BM cells (multiple t test, Holm-Sidak). Bottom: CRISPRi-mediated knockdown of Nt5e in Lp30 L-GMPs using gRNAs targeting the positions indicated by open circles. Nt5e expression was quantified by RT-qPCR, and expression levels were normalized to Actg1 and to cells transduced with gRNA targeting position −112 kb [n = 3, mean ± SD, one-way analysis of variance (ANOVA) with Dunnett’s correction, testing expression reductions between targeting enhancer or TSS positions versus the −112-kb control position]. (D) Dual luciferase assay assessing p42- or p30-mediated trans-activation of the −40-kb enhancer cloned upstream of a minimal promoter compared to an empty vector control (n = 3, mean ± SEM, t test). For all panels, *P ≤ 0.05; **P ≤ 0.01; ***P ≤ 0.001; ****P ≤ 0.0001.
other normal karyotype samples was most pronounced in the TCGA data, likely due to the lack of distinction between monoallelic and biallelic CEBPA mutant AMLs in the two older datasets (26, 27). Last, we used RT-qPCR to test the expression of NT5E in a panel of defined AML subtype and healthy volunteer samples. Notably, we found that, apart from biallelic CEBPA mutant AML, NT5E expression was only consistently up-regulated in INV(16) AML (Fig. 5B). Analysis of a previously generated ChIP-seq dataset demonstrated that the CBFB-MYH11 fusion protein, associated with INV(16) AML, is located right at the NT5E promoter (fig. S5B) (28). These findings strongly suggest that NT5E/CD73 is specifically up-regulated by two leukemic driver mutations in their respective AML subtypes.

To elucidate the regulation of NT5E expression in human biallelic CEBPA mutant AML, we performed ChIP-seq for the H3K27ac histone mark on sorted GMP samples from either bi- or monoallelic CEBPA mutant AML patients or from healthy volunteers. Notably, a region 48 kb upstream of the human NT5E TSS displayed K27 acetylation in biallelic CEBPA mutant cells but neither in the monoallelic AML sample nor in healthy cells (Fig. 5D). Comparison with an external H3K4me1 and H3K27ac dataset from CD34+ hematopoietic stem and progenitor cells (29) revealed enrichment for H3K4me1, but not for H3K27ac. This indicates that the −48-kb region is in an open but inactive state at the earliest phase of hematopoietic differentiation. Inspection of a comprehensive, human pan-hematopoiesis HiC study revealed that this region physically associates with the NT5E TSS in CD8+ T cells (Fig. 5D, black and red bars) (30). Another publicly available ChIP-seq study showed the region to be marked by both H3K4me1 and H3K27ac in a similar T cell subset (Fig. 5D) (31), suggesting that, in these NT5E-expressing T cells, the region functions as an active enhancer driving NT5E transcription. Last, the −48-kb region gave rise to a small but significant increase in reporter activity in response to either p42 or p30 expression (Fig. 5E), indicating that CEBPA is able to function as a trans-activator at this region.

In short, our data show that both CEBPA and NT5E are specifically overexpressed in human biallelic CEBPA mutant AML. Our identification of an AML-specific −48-kb enhancer that can be activated by CEBPA suggests that CEBPA directly drives overexpression of NT5E in human CEBPA mutant leukemia, in line with the mechanism observed in the mouse model.

**Down-regulation of Nt5e delays leukemia development in vivo**

We next wanted to assess whether Nt5e played a tumor-promoting role in CEBPA mutant AML, and to this end, we applied short hairpin RNA (shRNA)-mediated knockdown of the gene (Fig. 6A). Lp30 cells transduced with retroviruses encoding either a scrambled (control) shRNA or one of two individual shRNAs targeting Nt5e along with a green fluorescent protein (GFP) marker were used for a competitive BM transplantation (BMT) assay (Fig. 6B). BM cells were harvested 4 weeks after BMT, and the GFP/YFP (yellow fluorescent protein) ratio of the leukemic cells was analyzed by flow cytometry analysis (Fig. 6C). Nt5e knockdown leukemic cells were efficiently outcompeted by competitor cells in comparison to the scrambled mix, thus establishing a tumor-promoting role of Nt5e in leukemia progression in vivo (Fig. 6D).

To further assess the effect of Nt5e knockdown on disease progression, we carried out a survival analysis on recipients transplanted with AML cells transduced with shNt5e or scrambled control. In accordance with the competitive assays, we found that shRNA-mediated Nt5e knockdown led to a significantly increased latency time compared to the scrambled control, with a median survival of 51.5 days for the scrambled control and 66.5 and 71 days for each of the shRNAs (Fig. 6E).

To rule out the possibility of off-target effects of the shRNAs, and to validate the ability of the enhancer to regulate Nt5e expression in vivo, we applied CRISPRi using gRNA constructs targeting either the Nt5e TSS or the −40-kb enhancer for transcriptional repression (Fig. 4C, bottom). Sublethally irradiated mice were transplanted with sorted Lp30 cells double positive for the KRAB-dCas9-mCherry and sgRNA-GFP expression vectors. Targeting either the Nt5e TSS or the −40-kb enhancer for transcriptional repression significantly increased survival of recipient mice compared to targeting the −112-kb control region (Fig. 6F).

Collectively, these results establish Nt5e as a critical factor for tumor progression in Lp30 AML. Importantly, our data also provide functional validation of the −40-kb Nt5e enhancer in vivo.

**CD73 promotes tumor-protective adenosinergic autocrine signaling and is a potential therapeutic target in AML**

Up-regulation of CD73 can cause an increased generation of extracellular adenosine. Accumulation of adenosine in the extracellular space signals through one or more of the four adenosine receptors: A1R, A2AR, A2BR, or A3R encoded by Adora1, Adora2a, Adora2b, and Adora3 (32). RT-qPCR analysis of Lp30 and WT cell samples showed a specific and selective high expression of Adora2a, in comparison to Adora1, Adora2b, and Adora3 in the Lp30 cells (Fig. 6G).

Human biallelic CEBPA mutant AML also displays ADORA2A, ADORA2B, and ADORA3 expression, again with ADORA2A being the highest, which is parallelled in human INV(16) AML (fig. S6A). In a cancer setting, adenosine signaling, in particular, via the A2AR, may contribute to tumor progression by modulating inflammatory processes, promoting proliferation, and inhibiting apoptosis of the cancer cells, mostly via activation of the intracellular cyclic adenosine monophosphate (cAMP) pathway (33).

To examine expression changes caused by knockdown of Nt5e in the Lp30 cells, we performed RNA-seq analysis on Lp30 leukemic cells transduced with shNt5e or scrambled control (data file S5). To test the hypothesis that adenosine generated by CD73 can have an autocrine or paracrine effect on the leukemic cells through activation of A2AR, we curated a negative A2AR signature of downstream targets directly suppressed by A2AR signaling and calculated the signature enrichment alongside GO signatures identified using GSEA (data file S7). The negative A2AR signaling signature had the highest fold change between control and Nt5e knockdown Lp30 cells, followed by signatures associated with immune activation, responses that are affected by adenosine/cAMP signaling as well (Fig. 6H) (34). This suggests that targets normally repressed in Lp30 cell by A2AR signaling are indeed up-regulated by Nt5e knockdown, which supports the notion of an adenosinergic autocrine loop of signaling, promoted by CD73 in the Lp30 cells. Further, we queried the TCGA AML dataset by GSEA and found the A2A signature genes to be repressed in the biallelic CEBPA-mutated subtype versus all other subtypes (Fig. 6I), supporting the existence of subtype-specific signaling.

To directly assess the functional effect of A2AR signaling on cell growth, we measured the proliferation rates of Lp30 cells subjected to a specific A2AR inhibitor, SCH-58261. Cells treated with increasing concentrations of the inhibitor showed a dose-dependent decrease
in growth rates (fig. S6, B and C). In comparison, murine leukemic Inv(16) cells with low A2AR expression were not affected, demonstrating the requirement for A2AR (fig. S6, D and E). Analyses of A2AR inhibitor–treated Lp30 cells revealed a significant >2-fold decrease of cells in the S-G2-M phase (fig. S6, F, G, and H) and a marked increase of apoptotic cells (sub-G1 or annexin V marked) compared to the control (fig. S6, G and I).

Studies on solid cancer models have demonstrated anti-tumor effects of inhibiting CD73 activity with specific blocking monoclonal Abs (mAb) or using genetic knockdown (35–39). Expression of A2AR
A

B

C

D

E

F

G

H

I

J

K

Fig. 6. Effect of Nt5e KD on in vivo AML progression and identification of CD73 tumor-protective adenosinergic signaling as a potential therapeutic target.

(A) Nt5e knockdown efficiency; RT-qPCR quantification of Nt5e normalized to Actf. GFP+ sorted Lp30 cells transduced with shNt5e or scrambled control (n = 3, mean ± SD).

(B) Schematic outline of competitive BM transplant experiment. (C) Representative FACs of Lp30 cells transduced with shRNA targeting Nt5e or scrambled control at the input and output time points. (D) Target-to-competitor (GFP/YFP) ratio of Lp30 cells transduced with shRNA targeting Nt5e or scrambled control after BM and 4 weeks of leukemia progression. Ratio was normalized to input cells. (n = 4, mean ± SD). (E) Survival of recipient mice transplanted with 2.5 × 10^5 GFP+ sorted Lp30 cells transduced with shNt5e or scrambled control (n = 8). (F) Survival of recipient mice transplanted with 5 × 10^4 cells double positive for KRAB-dCas9-mCherry and sgRNA-GFP targeting Nt5e TSS, a putative enhancer (−40 kb), or a negative upstream region (−100 kb) (n = 7; control-Ig, n = 6; A2ARi + PBS, n = 7; Adora1, n = 5; Adora2a, n = 5; Adora2b, n = 5; Adora3, n = 5; CD73, n = 5; DAB6861, n = 5; DAB7461, n = 5). (G) Signatures enriched in both Nt5e knockdown groups in vivo compared to scrambled control based on RNA-seq (Scr, n = 3; shNt5e #1, n = 3; shNt5e #2, n = 2). Color indicates log2 fold change (red) and P value (purple) for each signature. See data file S6 for full list. (H) A2A suppressed direct targets GO humoral immune response GO regulation of leukocyte degranulation Neutrophil degranulation Associated with adenosine_Harmonizome

J

K

Statistics were determined by Student’s two-tailed t test (two groups) or one-way ANOVA corrected for multiple comparisons (Dunnett’s correction) between control and treatment groups (three or more groups) or log-rank test (survival). *P < 0.05; **P < 0.01; ***P < 0.001; ****P < 0.0001.
regions in WT GMPs displayed a relatively low openness as defined by H3K4me1 coverage, suggesting that these regions depend on the “pioneering” activity of full-length CEBPA (6, 41) and that p30 does not retain this activity. This is emphasized by the fact that p42 binding overlaps relatively more with late enhancers, which depend on the pioneering capacity of this isoform (6). As for the correlation between CEBPA binding and gene expression changes, the common regions were generally associated with genes down-regulated in L-GMPs. This is probably accounted for by the absence of the N-terminal TAD1 in the p30 isoform, which has previously been shown to have the major trans-activation potential of the protein. Conversely, p30-specific binding is associated with leukemia-specific increases in gene expression, underscoring that the TAD2 is sufficient (albeit less efficient) for gene activation, potentially also by cooperating with other TFs, like ERG. Collectively, our analyses demonstrate that CEBPA-p30 controls an AML-specific transcriptional program through novel binding at already established enhancers, induced, in part, by the increased expression of the p30 isoform, and that modulation of CEBPA binding may potentially be influenced by other TFs such as ERG and HLF.

Previous work has identified Cebpg and Sox4 as critical downstream targets of CEBPA in CEBPA silenced/CEBPA mutant AML, respectively (15, 16). Both proteins are normally repressed by CEBPA and up-regulated in its absence and, in the case of SOX4, up-regulated by the expression of the p30 isoform as well. In line with this, we found that both p42 and p30 bind the promoters of both genes and that SOX4 is up-regulated in Lp30 L-GMPs (data not shown). One possible explanation for the derepression of these genes following the p42 to p30 isoform switch could be that repression depends on the previously reported selective association of p42 with HDAC1 and HDAC2 (42).

Overall, our data, together with previous studies, suggest that the isoform switch from p42 to p30 expression drives an AML-specific transcriptional program via several mechanisms, including (i) loss of CEBPA-driven gene expression at p42-specific loci, (ii) loss of p42-mediated repression at common sites, (iii) loss of transcriptional activity at common regions due to reduced transcriptional activity of the p30 isoform, and (iv) activation of already established enhancers via p30 binding to low-affinity sites mediated by its increased expression and potentially also by ETS factors such as ERG.

Although targeting of, SOX4 and CEBPG may have a potential benefit in CEBPA mutant AML, TFs are generally not considered as good drug targets as cell-surface proteins or proteins harboring enzymatic activities such as kinases and epigenetic regulators. Here, we identified Nt5e as belonging to a transcriptional program driven by the p42 to p30 isoform switch in CEBPA mutant AML. We found Nt5e expression to be regulated by a p30-bound leukemic-specific enhancer in a manner that appears to be conserved across human and murine AML. Apart from CEBPA mutant AML, deregulated expression of NT5E was only consistently observed in INV16 AML, where the CBFβ-MYH11 fusion protein locates directly to the promoter of NT5E and likely drives its expression. This demonstrates that NT5E up-regulation is tightly associated with specific oncogenic drivers. Down-regulation of Nt5e either by shRNAs or through CRISPRi using gRNAs targeting the TSS or the p30-bound ~40-kb enhancer demonstrated a key role for CD73 as a tumor-promoting factor in CEBPA mutant AML.

CD73, together with CD39, catalyzes the formation of free adenosine on the surface of lymphocytes, endothelial cells, and a large
subset of primary tumors (22). By shifting the balance between the immune-activating ATP and immune-suppressing adenosine, CD73 has been proposed to protect cancer cells against the host immune system (21, 22). We tested this concept for the Lp30 AML model by comparing the effect of Nt5e knockdown in immunocompromised NSG mice versus C57BL/6 mice or following T cell depletion. However, in neither case did a reduction in the host immune system interfere with the tumor-promoting effect of CD73 (data not shown), suggesting that CD73, in CEBPA mutant AML, protects tumors under another mechanism. Free adenosine may also act on tumor cells in a paracrine/autocrine manner via the adenosine receptors (33). In line with this, we found that treatment of Lp30 cells in vitro with an A2AR receptor antagonist halted cell cycle progression and induced apoptosis. Moreover, we found that down-regulation of Nt5e promoted the derepression of genes, which are normally inhibited by the stimulatory action of A2AR on the adenylyl cyclase, one of its downstream targets. Thus, our observations are consistent with a mechanism where CD73-dependent adenosine acts on A2AR to sustain leukemic cell growth and protect against apoptosis, paralleling earlier reports using a model of chronic lymphocytic leukemia (24).

To test the clinical potential of targeting the CD73–A2AR adenosinergic pathway, we used a combined CD73 Ab-mediated blocking/A2AR inhibition strategy and observed a significant increase in survival of mice transplanted with Lp30 AML cells. The lack of pronounced efficacy of CD73 or A2AR targeted monotherapies has also been observed in solid tumor models and may involve pharmacokinetics issues of the CD73 Ab, which is optimized for human use (39, 43). We also note that the Ab isotype is an important parameter in our setting, as only the IgG2A and not the IgG1 isotype displayed therapeutic benefit (data not shown), perhaps reflecting the higher FcγR binding capacity of the former. Similar findings have been made in the context of solid cancers (39).

In conclusion, our work provides novel insights on how an isoform switch in a key myeloid TF drives a leukemic-specific transcriptional program, ultimately resulting in AML. We furthermore identify CD73 and A2AR as potential targets in biallelic CEBPA mutant AML via their ability to stimulate autocrine adenosinergic signaling. This contrasts observations in solid cancers, where CD73-mediated suppression of the adaptive immune system seems to be the main function of CD73 as a tumor-promoting factor (35, 36, 44), perhaps reflecting different physical properties of leukemic versus solid cancer niches. Currently, a number of agents targeting CD73 and the adenosinergic pathway are being evaluated for their efficacy against a variety of solid cancers in early clinical trials, either as single agents or in combination with immune modulatory therapies such as anti-PD1 or anti-PDL1 (45). Another intriguing possibility is the combination with Janus kinase inhibitors, shown recently to be a potential CEBPA mutant targeted therapy (46). On the basis of the findings in the present work, we suggest that such emerging treatment strategies should also be evaluated in the context of CEBPA mutant AML.

**MATERIALS AND METHODS**

**Experimental design**

To identify CEBPA-bound putative enhancers specific for CEBPA mutant AML, we performed microscan, in vivo ChIP-seq on sorted leukemic GMPs and their WT counterparts. RNA-seq was carried out to identify gene expression changes in human CEBPA mutant AML and the corresponding Cebpa<sup>Δ<sub>30</sub></sup> mouse model, leading to the identification of Nt5e, encoding CD73, as a cross-species AML gene. Functional validation of CD73 as a druggable tumor-promoting factor was carried out by shRNA or dCas9-KRAB–mediated KD or pharmacological inhibition followed by BMT.

**BMT, mouse**

All BMTs were carried out on sublethally irradiated 10- to 15-week-old female B6-SJL recipients by intravenous tail vein injections. For a detailed description of different experimental setups, see Supplementary Materials and Methods. The Cebpa<sup>Lp30</sup> mice were described in (11). All mouse experiments were conducted according to protocols approved by the Danish Animal Ethical Committee (permission #2013-15-2934-00780), with regard to the three R’s (refine, reduce, and replace) of animal experiments. Animals were housed in individually ventilated cages, and all experiments were carried out under supervision of veterinarians of the Department of Molecular Medicine, University of Copenhagen.

**Patients with AML and healthy controls, BM collection**

BM cells were aspirated from the posterior iliac crest of healthy subjects and patients with de novo AML before treatment. This study was performed in accordance with the Declaration of Helsinki under the following ethical approvals: Copenhagen, H-15004577; Aarhus, M-20070171; London, 10/H0704/65-006650QM; and Ulm, 148/10.

**Primary cultures and cell lines**

Lp30 cells were generated by serial BMT/expansion of BM cells from leukemic primary Cebpa<sup>Lp30</sup> mice. A pool of BM cells from tertiary expansions were used for both in vitro and in vivo experiments. BM cells were harvested from femur, tibia, and iliac bones; crushed; washed; pooled; and frozen in vials.

For in vitro proliferation, cell cycle, and apoptosis assays, Lp30 cells (female) were grown in R20/20 media: RPMI 1640 medium (Life Technologies) supplemented with 20% WEHI conditioned medium, 20% fetal bovine serum (FBS), 1% penicillin/streptomycin (Pen/Strep), and cytokines (PeproTech): stem cell factor (SCF) (20 ng/ml), IL-3 (10 ng/ml), and GM-CSF (20 ng/ml). A2AR inhibitor (SCH58264, Sigma-Aldrich) was added from stock (5 mg/ml) in dimethyl sulfoxide (DMSO) to 30, 10, or 3 μM, and DMSO volume was normalized/added as mock.

BM cells from recipients transplanted with donor BM cells from Chifb<sup>MYH11</sup> and KIT<sup>D816Y</sup> mutant mice [Chifb<sup>Δ16;50m</sup>; Tg(Mx1-Cre)/<sup>D816Y<sub>KIT</sub></sup>] [referred to as Inv(16) cells] (47) were grown in x-vivo complete media: X-VIVO 15 with gentamicin (Lonza), supplemented with 10% bovine serum albumin (BSA) (Stem Cell Technologies), 0.1 mM β-mercaptoethanol (Sigma-Aldrich), 1% L-glutamine (Gibco), 1% Pen/Strep, and cytokines (PeproTech): mSCF (50 ng/ml), hIL-6 (50 ng/ml), mIL-3 (10 ng/ml), and GM-CSF (10 ng/ml). Phoenix and HEK293T (human embryonic kidney line, female) cells were cultured in DMEM (Life Technologies) + 10% FBS and 1% Pen/Strep.

All cell cultures were mycoplasma-tested before freezing and never kept in culture for more than 3 months after rethawing. Phoenix and HEK293T cells were the only commercially available cell lines used, and virus production served as authentication of these cell lines.

**Proliferation assays [Lp30 and Inv(16) cells]**

Cells were plated at equal concentrations with varying concentrations of the inhibitor and in triplicate wells. Cells were counted, the concentration was calculated every second/third day, and the cells...
were replated at even cell concentrations. A growth curve was plotted for cumulative growth according to the dilution at each time point. Doubling time was calculated from fitted semi-logarithmic slope (Nonlin). Fit using least squares in GraphPad Prism software by log/slope and Student’s two-tailed test was used to test for significance. For cell cycle assays (Lp30 cells), cells were plated with 30 μM inhibitor or DMSO for 24 hours. Cells were harvested, and an equal number of cells were fixed in 2% paraformaldehyde, permeabilized in 0.1% saponin, stained with 4',6-diamidino-2-phenylindole (DAPI), and analyzed by flow cytometry (LSR-II, BD Biosciences). For apoptosis assay (Lp30 cells), cells were plated with 30 μM inhibitor or DMSO for 72 hours, harvested, washed and stained using a PE-annexin V kit (BD Biosciences), and analyzed by flow cytometry (Accuri, BD Biosciences). All flow cytometry analyses were run with FlowJo software V.10.

Retro- and lentiviral transductions

sgRNA nucleotides were cloned into pLKO5-sgRNA-EFS-GFP (Addgene) as described in (48) with BamBI digestion. pLKO5-sgRNA-EFS-GFP and pHV-SFFV-KRAB-dCas9-p2A-mCherry (Addgene) were used for lentiviral supernatant production by transfection of HEK cells.

shRNA-mir oligonucleotides were cloned into the pMSCV-LTRmiR30-5V-40-GFP vector using EcoRI restriction digest. Retroviral supernatant was produced by transfection of Phoenix-ECO cells.

Expanded, tertiary Lp30 cell vials were thawed and cultured in x-vivo complete media. One day after thawing, cells were transduced twice on two consecutive days by 50-min retro- or lentivirus spin-inhibitor or DMSO for 24 hours. Cells were harvested, and an equal number of cells were fixed in 2% paraformaldehyde, permeabilized in 0.1% saponin, stained with 4',6-diamidino-2-phenylindole (DAPI), and analyzed by flow cytometry (LSR-II, BD Biosciences). For apoptosis assay (Lp30 cells), cells were plated with 30 μM inhibitor or DMSO for 72 hours, harvested, washed and stained using a PE-annexin V kit (BD Biosciences), and analyzed by flow cytometry (Accuri, BD Biosciences). All flow cytometry analyses were run with FlowJo software V.10.

Flow cytometry

Analytical stainings were assessed on an LSRII, whereas cell sorting was carried out on either an Aria I or an Aria III (all instruments were from Becton Dickinson). Detailed descriptions of all stainings and gating strategies are provided in Supplementary Materials and Methods.

Western blotting

Flow cytometry–sorted GMPs (see above) from leukemic Lp30 and WT mice, corresponding to approximately 10,000 cells for each sample, were boiled in SDS-loading buffer for 5 min, subjected to nucleic acid degradation by Benzonase (E1014-5KU, Sigma-Aldrich) for 20 min on ice, and spun for 20 min at 20,000g. Material was size-separated using NuPAGE precast 4 to 12% Bis-tris gels (Invitrogen). The Cell Signaling protocol (www.cellsignal.com/support/protocols/western.html) was used for blotting. ImageJ was used for quantifications using program guidelines (http://rsb.info.nih.gov/ij/). CEBPA Ab [at 1:1000 dilution in 5% (w/v) BSA, 1 hour at room temperature] corresponds to the one used for ChIP (clone 14AA, sc-61, Santa Cruz Biotechnology), and loading control was anti-Histone H3 [1:5000 in 5% (w/v) nonfat milk blocking buffer, 2 hours at room temperature; Ab10799, Abcam]. A high-sensitivity chemiluminescence HRP detection kit was used (Amersham ECL Prime Western Blotting Detection Reagent, RPN2232).

Transient transfection/dual luciferase assay

Luciferase assays were performed according to the kit manufacturer protocol (cat# E1910, Promega). Briefly, 15,000 HEK293 cells were plated in a white-bottom 96-well tissue culture plate. The following day, 0.2 ng of Renilla control vector (pRL-CMV, cat# E2261, Promega), 20 ng of empty or enhancer-containing promoter-pGL4 (pGL4:23, cat# E841A, Promega), and 20 ng of either empty or CEBPA-construct containing pcDNA3 (MSC-154) (Thermo Fisher Scientific) vector were transfected using a 3:1 μl/μg ratio of TransIT-2020 (MIR 5404, Mirus) reagent. After a 24-hour incubation, readouts were obtained using a standard luminometer. Constructs used include pcDNA3-Cebpa(WT) and pcDNA3-Cebpa(p30) expression constructs, and pGL4:23 containing human and mouse putative NT5E/NT5e enhancers cloned to the multiple cloning site with the primers listed in data file S7. The human enhancer was first cloned to the TOPO vector (pCR-2.1-ToPO, K450002, Thermo Fisher Scientific) and then cloned using Kpn I and Xho I restriction enzymes. The mouse enhancer was cloned directly from PCR with the same restriction enzymes.

mRNA–RT-qPCR expression analysis

For expression analysis, cells were sorted as described above, RNA-extracted using the RNeasy Micro Kit (QIAGEN) or NucleoSpin RNA XS (MN), and reverse-transcribed to cDNA using the ProtoScript First Strand cDNA Synthesis Kit (M-MuLV) and oligo (dT) primers (NEB). RT-qPCR was performed using LightCycler 480 SYBR Green I Master (Roche Life Sciences). Data were normalized to Actg1 (mouse) or POL2AR and H6PD (human). Primers used are listed in data file S7.

Preparation of human cDNA libraries and sequencing

Total RNA was extracted from GMPs using the RNeasy Micro Kit (QIAGEN). Thirty nanograms of RNA was processed for double-stranded cDNA synthesis with the Ovation RNA-Seq System V2 (NuGEN Technologies). Subsequently, 1 μg of the sheared cDNA [fragment size, 200 to 600 base pairs (bp)] was subjected to library preparation according to the Illumina TruSeq DNA Sample Preparation Kit (ref #15012999) protocol. The indexed libraries were pooled in equimolar ratios and subjected to 100-bp paired-end sequencing on the Illumina HiSeq 2000 system.

Preparation of Lp30 cDNA libraries and sequencing

Recipient mice were transplanted with shRNA-transduced Lp30 cells (1.5 × 106 for each Nt5e knockdown construct and 0.5 × 106 for scrambled control). Three weeks after BMT, 5 × 105 GFP-sorted cells from two or three individual mice per construct were used for mRNA extraction (RNeasy mini kit, QIAGEN), and cDNA library was generated using 600 ng of input RNA (TruSeq Library RNA Prep Kit v2, Illumina). Libraries were diluted and sequenced (NextSeq, Illumina).

Mapping and quantification of RNA-seq data

See Supplementary Materials and Methods for detailed description.

Mass spectrometry

Detailed description of the procedure can be found in Supplementary Materials and Methods. Briefly, FACS-sorted cells were washed twice with ice-cold PBS and lysed in 6 M guanidinium hydrochloride, 10 mM tris(2-carboxyethyl)phosphine, 40 mM 2 -chloroacetamide, and 100 mM tris (pH 8.5). Proteins were digested first with LysC and subsequently with trypsin at 37°C. Enzyme activity was quenched by trifluoroacetic acid (TFA) at a final concentration of 1%. Before mass spectrometry (MS) analysis, the peptides were desalted on in-house packed C18 StageTips. Samples were analyzed in a label-free manner, where each sample was loaded onto a 2-cm C18 trap column,
connected in line to a 50-cm C18 reverse-phase analytical column (Easy-Spray ES803, Thermo Fisher Scientific). Peptides were eluted over a 200-min gradient, ranging from 5 to 48% of acetonitrile at 250 nl/min, and analyzed in a ddMS2-IT-HCD top speed method on an Orbitrap Fusion (Thermo Fisher Scientific). MS performance was verified for consistency by running complex cell lysate quality control standards, and chromatography was monitored to check for reproducibility.

**Chromatin preparation and immunoprecipitation**

ChiP on both mouse and human samples was performed as described previously (49, 50), with the exception that amplification for the human ChiP samples was done with the New England Biolabs Ultra Amplification Kit (cat# E7370, NEB) and no Escherichia coli carrier DNA. Specific washing conditions not mentioned previously include four times low-salt (140 mM NaCl) radioimmunoprecipitation assay (RIPA) buffer washes for both H3K4me1 and H3K27ac precipitations, replacing all high-salt (500 mM) washes [see (50) for detailed protocol]. Abs used were CEBPA (clone 14AA, sc-61, Santa Cruz Biotechnology), H3K4me1 (ab8895, Abcam), and H3K27ac (ab4729, Abcam). See primer table (data file S7) for primer sets used for assessing the quality of each individual precipitation.

**Mapping and quantification of ChiP-seq data**

See Supplementary Materials and Methods for detailed description.

**3C-qPCR**

3C was done essentially as described previously (51) with some modifications. Specifically, mouse Lp30 leukemic GMP cells (Lin− cKit+ Sca1− CD150+ CD41+ FcγRII/III) were isolated by FACS, while cKit+ WT BM cells were isolated by magnetic-activated cell sorting (MACS) (CD117 magnetic microbeads, Miltenyi Biotech) for comparison. Approximately 2 million cells were used per reaction, fixed in 1.5% formaldehyde in 10 ml of cold PBS/RPMI medium 50%/50% rotating at room temperature for 10 min, and quenched for 2 min in 187 mM of added glycine. Lysis was done by douncing (glass) 10 strokes on ice after 15 min of incubation on ice. Cell fragments/nuclei were recovered by centrifugation at 2200g for 5 min at 4°C and washed once in ice-cold restriction buffer and resuspended in 0.5 ml of restriction buffer. Chromatin was exposed for digestion by a 1-hour incubation with 0.1% of SDS at 37°C, followed by a 1-hour incubation with 1% Triton X-100 at 37°C. Overnight digestion with 200 U of Xba I (R0145T, NEB) with the addition of 100 U of enzyme for the last 4 hours was followed by 30 min of heat inactivation at 65°C. Ligation was performed with 20,000 U of T4 ligase (M0202T, NEB) for 3 hours at 20°C. DNA was isolated by RNase A (19101, QIAGEN) exposure for 30 min, followed by addition of 1 mg of proteinase K (P6556, Sigma-Aldrich) and incubation in 0.4% SDS for 2 hours and phenol-chloroform extraction using phase-lock tubes (713-2536, 5-prime). R1-qPCR was performed as described above, with quadruplicates for each data point and a high-stringency, 50-cycle program. All primers are listed in data file S7.

**Statistical analysis**

The statistical analyses used, the definition of what n represents, and the meaning of numbers of asterisks are indicated for each experiment in the relevant figure legends. For BMT experiments, recipient mice were randomized to receive control and test leukemia cells, respectively. For survival analysis, individual mice were shuffled between cages. See Supplementary Materials and Methods. No blinding of experimental groups was performed. No statistical method was applied to predetermine sample sizes, but sample sizes are indicated in relevant figures.

**Supplementary Materials**

Supplementary material for this article is available at http://advances.sciencemag.org/cgi/content/full/5/7/eaaw4304/DC1

**Supplementary Materials and Methods**

See Supplementary Materials and Methods for detailed description.

**Data File 51.** CEBPA peak coordinates, enriched motifs in CEBPA regions, related to Fig. 1.

Data File 52. RNA-seq data, related to Figs. 1 and 2.

Data File 53. MS data, related to Figs. 1 and 2.

Data File 54. Gene signatures, related to Fig. 3.

Data File 55. RNA-seq data, related to Fig. 6.

Data File 56. Gene signatures, related to Fig. 6.

Data File 57. Materials lists, related to Materials and Methods.

References (52–73).

**downloaded from http://advances.sciencemag.org on August 27, 2019.**


Acknowledgments: We thank A. Fossom for help with cell sorting and members of the Porse lab for discussions. MedImmune provided the anti-CD73 Ab used for in vivo treatment experiments. Funding: This study was supported by the Danish Cancer Society, the Danish Association for Cancer Research, and the Novo Nordisk Foundation and through a center grant from the Novo Nordisk Foundation (Novo Nordisk Foundation Center for Stem Cell Biology, DanStem; grant number NRF17CC002785). The AML tissue bank at Queen Mary University was supported by the Medical College of St. Bartholomew’s Hospital Trust. Author contributions: J.S.J., L.G.L., M.B.S., Y.G., E.S., C.G., T.D., and J.J. performed the experiments. J.S.J., L.G.L., M.B.S., P.P., E.S., K.V.-S., J.F., K.R., K.T.-M., and B.T.P. analyzed the data. J.S.J., L.G.L., M.B.S., Y.G., E.S., C.G., T.D., and J.J. performed the experiments. The remaining authors declare that they have no competing financial interests. J.S.J. is presently an employee of Symphogen A/S. The remaining authors declare that they have no competing interests. Data and materials availability: All data needed to evaluate the conclusions in the paper are present in the paper and/or the Supplementary Materials. All sequencing data can be retrieved at GEO (GSE118963). The MS data have been deposited to the ProteomeXchange Consortium (http://proteomcentral.proteomeexchange.org) via the PRIDE partner repository with the dataset identifier PXD011359. Additional data related to this paper may be requested from the authors.

Submitted 19 December 2018 Accepted 31 May 2019 Published 10 July 2019 10.1126/sciadv.aaw4304

Mutant CEBPA directly drives the expression of the targetable tumor-promoting factor CD73 in AML
Janus S. Jakobsen, Linea G. Laursen, Mikkel B. Schuster, Sachin Pundhir, Erwin Schoof, Ying Ge, Teresa d'Altri, Kristoffer Vitting-Seerup, Nicolas Rapin, Coline Gentil, Johan Jendholm, Kim Theilgaard-Mönch, Kristian Reckzeh, Lars Bullinger, Konstanze Döhner, Peter Hokland, Jude Fitzgibbon and Bo T. Porse

Sci Adv 5 (7), eaaw4304.
DOI: 10.1126/sciadv.aaw4304