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Serine-Threonine Kinases Encoded by Split hipA Homologs Inhibit Tryptophanyl-tRNA Synthetase

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ABSTRACT  Type II toxin-antitoxin (TA) modules encode a stable toxin that inhibits cell growth and an unstable protein antitoxin that neutralizes the toxin by direct protein-protein contact. hipBA of Escherichia coli strain K-12 codes for HipA, a serine-threonine kinase that phosphorylates and inhibits glutamyl-tRNA synthetase. Induction of hipA inhibits charging of glutamyl-tRNA that, in turn, inhibits translation and induces RelA-dependent (p)ppGpp synthesis and multidrug tolerance. Here, we describe the discovery of a three-component TA gene family that encodes toxin HipT, which exhibits sequence similarity with the C-terminal part of HipA. A genetic screening revealed that trpS in high copy numbers suppresses HipT-mediated growth inhibition. We show that HipT of E. coli O127 is a kinase that phosphorylates tryptophanyl-tRNA synthetase in vitro at a conserved serine residue. Consistently, induction of hipT inhibits cell growth and stimulates production of (p)ppGpp. The gene immediately upstream from hipT, called hipS, encodes a small protein that exhibits sequence similarity with the N terminus of HipA. HipT kinase was neutralized by cognate HipS in vivo, whereas the third component, HipB, encoded by the first gene of the operon, did not counteract HipT kinase activity. However, HipB augmented the ability of HipS to neutralize HipT. Analysis of two additional hipBST-homologous modules showed that, indeed, HipS functions as an antitoxin in these cases also. Thus, hipBST constitutes a novel family of tricomponent TA modules where hipA has been split into two genes, hipS and hipT, that function as a novel type of TA pair.

IMPORTANCE  Bacterial toxin-antitoxin (TA) modules confer multidrug tolerance (persistence) that may contribute to the recalcitrance of chronic and recurrent infections. The first high-persister gene identified was hipA of Escherichia coli strain K-12, which encodes a kinase that inhibits glutamyl-tRNA synthetase. The hipA gene encodes the toxin of the hipBA TA module, while hipB encodes an antitoxin that counteracts HipA. Here, we describe a novel, widespread TA gene family, hipBST, that encodes HipT, which exhibits sequence similarity with the C terminus of HipA. HipT is a kinase that phosphorylates tryptophanyl-tRNA synthetase and thereby inhibits translation and induces the stringent response. Thus, this new TA gene family may contribute to the survival and spread of bacterial pathogens.

KEYWORDS  persistence, ppGpp, toxin/antitoxin systems, translation, tRNA synthetase

Prokaryotic toxin-antitoxin (TA) modules are usually composed of two elements, a toxin that can inhibit cell growth and an antitoxin that counteracts the inhibitory effect of the toxin (1, 2). Based on the molecular modes of antitoxin activity, TA modules have been divided into different types (3). The abundant type II modules are charac-


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characterized by protein antitoxins that bind directly to and inhibit their cognate toxins by tight molecular interaction. Type II antitoxins usually contain a DNA-binding motif used to regulate TA operon transcription via binding to operators in the promoter region and a separate domain that interacts with and neutralizes the cognate toxin. Moreover, antitoxins are degraded by cellular proteases, such as Lon and/or Clp, and the cellular activity and amount synthesized of a given toxin are thus determined by the concentration of cognate antitoxin (4).

Type II modules are highly abundant; that is, most prokaryotic chromosomes encode at least one and some chromosomes encode cohorts of them. For example, *Mycobacterium tuberculosis* has 88 known, well-conserved type II TAs, while the insect pathogen *Photorhabdus luminescens* has a similarly large cohort (5). Toxin gene similarities were used to divide type II modules into superfamilies (6, 7). Thus, in general, toxins that exhibit sequence similarity inhibit cell growth by identical or related molecular mechanisms and can be grouped into the same family. Type II toxins belonging to the RelE, MazF, VapC, HipA, and TacT families curtail cell growth by inhibiting translation, CcdB and ParE inhibit DNA replication, Zeta toxins inhibit cell wall synthesis, and RES toxins inhibit cell growth by depleting NAD$^+$ (8–18).

The biological functions of TAs have been debated. For type II modules, many studies now point to a function in survival during stress, including tolerance of multiple antibiotics (1). Stochastic or stress-induced activation of TA modules can protect bacteria from unfavorable environmental conditions by inducing persister formation (19, 20), a transient, slow-growing state in which the bacteria are tolerant of antibiotics and various other forms of stress (21). The stochastic formation of persisters is due to phenotypic heterogeneity in clonal populations of cells and can be viewed as a bet-hedging strategy that increases the survival rate in rapidly changing environments (22). Moreover, sublethal concentrations of antibiotics and other stresses have been found to stimulate the formation of persisters (23, 24).

The first gene associated with persistence was *hipA* (high persister gene A) of *Escherichia coli* strain K-12, identified as a gain-of-function allele, *hipA7* (25). This allele, found also in clinical isolates of uropathogenic *E. coli* (26), showed a 100- to 1,000-fold increase in persistence due to two amino acid changes in HipA (changes of G to S at position 22 [G22S] and D to A at positions 291 [D291A]) (27). The *hipA* toxin gene and the upstream *hipB* antitoxin gene together constitute a type II TA module (28). Modest ectopic expression of wild-type HipA causes severe growth inhibition that can be countered by the HipB antitoxin, which interacts directly with HipA (28). HipA and HipB form a complex that represses *hipBA* transcription via binding to operators in the promoter region (26). HipA is a Hanks serine-threonine kinase (29, 30) and was found to specifically phosphorylate and inhibit glutamyl-tRNA synthetase (GltX or GltRS), causing strong inhibition of translation and induction of guanosine tetra- and pentaphosphate ([p]ppGpp) synthesis and persistence (11, 31, 32). HipA-mediated phosphorylation of the conserved residue Ser239 inhibits the activity of GltX (11), thereby preventing charging of tRNA$^{Glu}$. As a consequence, the ratio of charged to uncharged tRNA$^{Glu}$ decreases, which in turn stimulates binding of RelA-tRNA complexes to the ribosome, leading to activation of RelA (33). The resulting increase in the cellular [p]ppGpp level triggers the stringent response (11, 27, 32).

Here, we describe a novel family of three-component TA modules encoding toxins exhibiting sequence similarity to HipA. We discovered that HipT of the enteropathogenic *E. coli* O127:H6 strain E2348/69 (HipTO127) is a toxin that can be counteracted by overproduction of tryptophanyl-tRNA synthetase (TrpS or TrpRS). Consistently, our *in vitro* data show that HipTO127 is a serine-threonine kinase that inhibits translation by phosphorylating TrpS. HipTO127 aligns colinearly with HipA but lacks $\sim$100 amino acids (aa) at its N terminus (Fig. 1A). Interestingly, *hipTO127* is preceded by *hipS0127*, encoding HipS$_{O127}$ (103 aa), which exhibits sequence similarity with the N-terminal part of HipA that is missing from HipTO127 (Fig. 1A). Finally, *hipS0127* is preceded by a gene encoding a HipB homolog containing a helix-turn-helix (HTH) DNA-binding motif. HipB, HipS, and HipT form a complex *in vivo* and *in vitro*, and HipS$_{O127}$ alone counteracts HipTO127.
FIG 1  hipBST of E. coli O127 encodes a three-component toxin-antitoxin module. (A) Schematic showing a comparison of the hipBA and hipBST operons of E. coli K-12 and O127, respectively. Bent arrows pointing right indicate promoters. The hipBA operon contains two genes, hipB and hipA, while hipBSTO127 contains three genes, hipB\textsubscript{O127}, hipS\textsubscript{O127}, and hipT\textsubscript{O127}. The region of hipA between the dashed lines exhibits sequence similarity to hipS\textsubscript{O127}. The 8 amino acid residues of the P loop in HipA (150-VAGAQEKT-158) that binds phosphates of ATP are shown; the autophosphorylated S150 residue is shown in green (35). The homologous P loop and autophosphorylated serine in HipT\textsubscript{O127} were inferred by sequence similarity. (B) Overnight cultures of E. coli MG1655 harboring pSVN1 (pBAD33::hipTO127) or the empty pBAD33 vector combined with pSVN111 (pNDM220::hipBO127), pSVN109 (pNDM220::hipSO127), pSVN110 (pNDM220::hipBSO127), or the empty low-copy-number pNDM220 vector, as indicated, were diluted to obtain the same values of OD\textsubscript{600}, centrifuged at 5,000 rpm for 5 min, washed in phosphate-buffered saline (PBS), and serially diluted before being spotted onto LB nutrient agar plates containing 0.2% glucose (to repress hipTO127), or 0.2% arabinose plus 200 μM IPTG (to induce hipTO127, hipSO127, or hipBSO127). (C) The strains used in the experiment whose results are shown in panel B were grown in LB medium plus appropriate antibiotics. Overnight cultures were diluted, cells were grown exponentially for at least 3 h until the doubling time appeared constant, and at an OD\textsubscript{600} of ~0.3, arabinose (0.2%) was added to induce hipT\textsubscript{O127} (red arrow). After a further 1.5 h, IPTG (200 μM) was added to induce hipS\textsubscript{O127}, hipB\textsubscript{O127}, or hipBS\textsubscript{O127} (green arrow). (D and E) Viable counts of strains from the experiment whose results are shown in panels B and C were grown in PBS before a 10-times dilution series was spotted on agar plates with glucose (0.2%) to repress hipTO127 expression (D) or with glucose (0.2%) to repress hipTO127 expression and IPTG (200 μM) to induce hipBS\textsubscript{O127}, hipS\textsubscript{O127}, or hipBS\textsubscript{O127} (E). Plates were incubated for 16 h at 37°C before counting. Data points in panels C, D, and E represent mean values of results from at least three independent experiments, and error bars indicate standard deviations.
activity in vivo. The HipB homolog (called HipBO127) does not counteract HipTO127 but instead augments the ability of HipSO127 to counteract HipTO127. Analysis of the hipBST modules of *Haemophilus influenzae* and *Tolumonas auensis* revealed that the HipT proteins of these organisms also are counteracted by overproduction of TrpS. Moreover, cognate HipS neutralizes HipT in both these cases. In summary, we describe here a family of novel three-component TA modules that potentially can increase the stress resilience and spread of bacterial pathogens.

**RESULTS**

Homologs of HipA are encoded by three-gene operons. Using similarity searching with HipA (440 amino acids [aa]) of *E. coli* K-12 as the query sequence, we identified a number of genes encoding HipA homologs that aligned colinearly with the C terminus of HipA but were shortened by ~100 aa at their N termini (Fig. 1A, and see Fig. S1A in the supplemental material) (34). The HipA homologs contain P-loop motifs matching the experimentally validated P loop of HipA, as well as conserved catalytic domains and Mg$^{2+}$ binding motifs, suggesting that, like HipA, HipT proteins are kinases (Fig. S1A) (35). A phylogenetic analysis showed that HipA and HipT group monophyletically in a cladogram based on 8 HipA and 40 HipT sequences (Fig. S1D) (36). The majority of the *hipT* genes were from gammaproteobacteria, but two HipT homologs deeply embedded in the phylogenetic tree were from a *deltaproteobacterium* and a *firmicute* (*Streptococcus pneumoniae*) (Fig. S1D). The HipT homolog from *S. pneumoniae* is identical to the HipT homolog of *H. influenzae* strain 10810 (Fig. S1D). These two organisms separated more than a billion years ago, and both are highly competent for DNA uptake and live in the same biological habitat (the upper respiratory tract). These observations raise the possibility that *hipBST* loci undergo lateral gene transfer between distantly related organisms.

In all the *hipT*-containing organisms examined, we discovered short open reading frames adjacent to and upstream from *hipT*, which are herein called *hipS*, encoding proteins of ~100 aa that exhibit sequence similarity to the missing N-terminal part of HipA (Fig. S1B). In all these cases, open reading frames upstream from *hipS* encode putative proteins of ~100 aa containing HTH DNA-binding motifs (Fig. S1C). These putative HipB homologs may thus autoregulate the *hipBST* operons. We chose the *hipBST* module of *E. coli* O127:H6 strain E2348/69 as our primary model system for functional analysis (Fig. 1A). The *hipBST* module encodes HipBO127 (107 aa), HipSO127 (103 aa), and HipTO127 (335 aa). Gene pair *hipBO127* and *hipSO127* overlaps by 16 nucleotides (nt), and gene pair *hipSO127* and *hipTO127* overlaps by 1 nt, suggesting that the genes may be translationally coupled.

HipTO127 inhibits cell growth and is counteracted by HipSO127. We validated the components encoded by *hipBST* experimentally by inserting *hipTO127* into plasmid vector pBAD33 (carrying the arabinose-inducible pBAD promoter) and *hipSO127*, *hipBO127*, and *hipBSO127* into the low-copy-number R1 vector pNDM220 (carrying the synthetic, isopropyl-$\beta$-D-thiogalactopyranoside [IPTG]-inducible pA1/O4/O3 promoter) and subjected the standard *E. coli* K-12 strain MG1655 carrying combinations of these plasmids to growth assays and viable-count measurements. Induction of *hipTO127* resulted in strong inhibition of cell growth, both on plates and in liquid medium, supporting the hypothesis that HipTO127 can function as a toxin (Fig. 1B and C). Growth was rescued by induction of *hipSO127* alone but not by *hipBO127* alone, suggesting that HipSO127 functions as the antitoxin (Fig. 1B and C). Coinduction of *hipBO127* and *hipSO127* provided a consistent, yet mild growth rescue advantage compared to the results for *hipSO127* alone, suggesting that HipBO127 augments the antitoxin activity of HipSO127 (Fig. 1C) Thus, HipBO127 does not function as a classical antitoxin.

Upon induction of *hipTO127*, CFU decreased dramatically (Fig. 1D). However, later induction of *hipSO127*, or *hipSO127* plus *hipBO127* (by adding IPTG and glucose to the agar plates to induce $P_{\text{A1/O4/O3}}$-*hipBSO127* and repress pBAD-*hipTO127*, respectively) fully rescued cell viability (Fig. 1E). This result showed that ectopic production of HipTO127 induces a bacteriostatic condition from which the cells can be resuscitated. In support
of this conclusion, strains in which \( hipt_{O127} \) was induced recovered viability after prolonged incubation times (~40 h), even in the absence of \( hips_{O127} \) or \( hips_{B127} \) (Fig. S2).

We were puzzled by the observation that HipSO127 but not HipBO127 exhibited antitoxin activity and therefore decided to analyze the hipBST modules of two additional gammaproteobacteria, \( Haemophilus influenzae \) Rd KW20 (hipBSTHi) and \( Tolumonas auensis \) DSM 9187 (hipBSTTa) (Fig. S1). Induction of \( hipTHi \) or \( hipTTa \) inhibited cell growth of \( E. coli \) MG1655 in liquid medium in both cases, and induction of cognate hipS genes was sufficient to neutralize the two HipT toxins (Fig. S3A and B). Like HipBO127, HipBSTHi augmented the ability of HipSHi to neutralize HipTHi, as the presence of the HipBSTHi-encoding plasmid almost entirely prevented growth inhibition after induction of hipTHi (Fig. S3A). HipBSTTa did not detectably augment the antitoxin effect of HipSTa in this experimental setup (Fig. S3B). We also tested \( hipT \) genes from strains of \( Vibrio cholerae \) and \( Vibrio halioticoli \), but their induction was, for unknown reasons, not toxic in \( E. coli \) K-12 and the corresponding hipBST modules were not analyzed further.

**HipB, HipS, and HipT form a ternary complex in vivo.** The above-described observations suggest that HipBO127, HipSO127, and HipTO127 might form a protein complex in vivo, as seen for other type I TA modules. To test this, we constructed a plasmid (pSVN94) encoding N-terminally His6-tobacco etch virus (TEV)-tagged HipBO127, HipSO127, and the enzymatically inactive HipTO127\(_{D233Q} \) mutant protein in which all three genes had optimized translation signals (Shine-Dalgarno [SD] sequences and ATG start codons) to increase translation rates. His6-TEV-HipBO127 was purified under native conditions and analyzed by denaturing polyacrylamide gel electrophoresis (SDS-PAGE). Indeed, three proteins of the expected molecular weights (MWs) copurified (Fig. S4A), indicating that the HipBSTO127 proteins form a complex in vivo. Further separation of the protein complex using a heparin column allowed isolation of three samples containing HipTO127, HipBTO127, and HipBSTO127 (Fig. S4B, top). Gel filtration chromatography further confirmed that HipT and HipBST are monodispersed in solution, suggesting that HipBSTO127 is a heterotrimer (Fig. S4B, bottom).

**Multicopy suppression of HipT by trpS.** Previously, we showed that overproduction of GltX suppresses HipA-mediated growth inhibition and that HipA phosphorylates GltX in vitro (11). Unexpectedly, overproduction of GltX did not suppress HipT-mediated growth inhibition (Fig. S5A). Therefore, we performed a second multicopy gene library screening in an attempt to identify genes that in high copy numbers could suppress the effect of HipTO127 (see Materials and Methods). Using a pBR322-based Sau3A-derived gene library of \( E. coli \) MG1655ΔydeA, 8,300 colonies with an average insert size of ~3,300 bp were screened, resulting in a coverage of roughly 5.8 times. In this screening, 105 hits were obtained, of which 19 were retransformed. Six of these plasmids exhibited a stable phenotype and were sent for sequencing. Thereby, we identified a DNA fragment containing \( rpe \), \( gph \), and \( trpS \) that suppressed HipTO127. Of these genes, only conditional induction of \( trpS \), which encodes tryptophanyl-tRNA synthetase (TrpS), suppressed HipTO127-mediated growth inhibition, both on solid medium (Fig. S5B) and in liquid culture (Fig. 2A). TrpS also suppressed HipTHi and HipTTa (Fig. 2B and C and Fig. S5B), whereas GltX had no such effect (Fig. S5A).

**HipT phosphorylates TrpS at a conserved sequence motif.** The above-described results suggested that HipT phosphorylates TrpS. To analyze HipT kinase activity directly, we purified HipTO127 and its presumed target, TrpS. For comparison, we included HipA and its known target GltX in the analysis. Indeed, HipTO127 phosphorylated TrpS in vitro (Fig. 3A, lanes 5 and 8) in a reaction that did not require tRNA (Fig. S6). We showed previously that HipA phosphorylates GltX in vitro in a reaction that requires the addition of tRNA\(_{Glu} \) (11). Here, we were able to reproduce the results showing that HipA phosphorylates GltX in the presence but not in the absence of tRNA (Fig. 3B, lanes 6 and 9). Thus, HipT and HipA kinases differ not only with respect to their specific target but also by whether there is a requirement for the presence of tRNA in the in vitro reaction mixtures (see Discussion).
The best-conserved stretch of amino acids between GltX and TrpS are the highly conserved KLS239KR/KMS197KS flexible-loop motifs (Fig. S7). Lys237 and Lys195 participate in the catalytic reaction by stabilizing the transition state of ATP, and intact loop motifs are required for catalysis (37). The observation that HipA phosphorylates GltX at S239 (11) raised the possibility that HipT phosphorylates TrpS at the homologous S197. To test this, we introduced two amino acid changes, S197A and S197D, into TrpS, the latter to mimic a phosphorylated serine. Both changes abolished phosphorylation of HipTO127, consistent with the proposal that HipT phosphorylates TrpS at S197 (Fig. 3C). Finally, mass-spectrometric analysis revealed that, indeed, HipTO127 phosphorylates TrpS at S197 in vitro (Table 1). We also note that HipA did not phosphorylate TrpS (Fig. 3A, lanes 6 and 9), while HipTO127 did not phosphorylate GltX (Fig. 3B, lanes 5 and 8). This lack of cross-reactivity in the in vitro reactions is consistent with the specificity of the multicopy suppression data.

![Diagram](http://mbio.asm.org/)
HipA is known to inactivate itself by trans autophosphorylation at Ser150 (35, 38). In the reaction mixture containing only HipTO127, a faint radioactive band corresponding to the MW of HipTO127 was observed (Fig. 3A, lane 2). Since HipTO127 was the only protein in the reaction mixture, we infer that HipTO127 phosphorylates itself. Consistently, the weak HipTO127 band also appeared when HipTO127 was mixed with the noncognate target Gltx (Fig. 3B, lanes 5 and 8). Accordingly, the analysis of the products of the in vitro reaction between HipTO127 and TrpS by mass spectrometry showed that HipTO127 autophosphorylates either on S57 or S59 (Table 1).

**HipTO127 stimulates production of (p)pGpp.** We and others showed previously that HipA activates RelA to synthesize (p)pGpp (11, 31, 32). Here, we measured...
whether induction of hipT$_{O127}$ induces (p)ppGpp synthesis and compared it to the effect of induction of hipA or relE of E. coli K-12, the latter of which inhibits translation by ribosome-dependent mRNA cleavage (8). Indeed, induction of both hipT$_{O127}$ and hipA resulted in increased levels of (p)ppGpp, albeit at a somewhat lower level in the case of hipT$_{O127}$ (Fig. 4A and B). The latter observation is consistent with the fact that tryptophan is encoded by one codon only, compared to two in the case of glutamate, and the fraction of tryptophanyl-tRNA is less than 2% of total tRNA, whereas that of glutamyl-tRNA is more than 7% (39). Thus, deficiency of charged tRNA$^{Glu}$ leads to a higher level of hungry ribosomal A sites and, therefore, a higher number of activated RelA molecules and a higher level of (p)ppGpp. Consistent with previous results (40),

<table>
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<th>Protein</th>
<th>Amino acid</th>
<th>Andromeda score</th>
<th>Localization probability</th>
<th>Mass error (ppm)</th>
<th>Phosphopeptide sequence of the best localized MS/MS spectrum</th>
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<td>HipT</td>
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<tr>
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<td>S197</td>
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<td>0.999003</td>
<td>0.17001</td>
<td>KMS(1)KSDDNRNNGILLEDPK</td>
</tr>
<tr>
<td>TrpS</td>
<td>S199</td>
<td>309.72</td>
<td>0.864936</td>
<td>0.094932</td>
<td>SGARVMSLLEPTKKS(0.135)KS(0.865)DDNRNNGILLEDPK</td>
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**Fig 4** HipT$_{O127}$ induces (p)ppGpp accumulation in vivo. Levels of (p)ppGpp of E. coli MG1655 containing pNDM220 (vector), pAH1 (pNDM220::hipA), pSVN116 (pNDM220::hipT$_{O127}$), or pAH2 (pNDM220::relE). The toxin-encoding genes were located downstream from the IPTG-inducible P_{A1-03/04} promoter (56, 57). (A) Cells were grown exponentially at 37°C in low-phosphate MOPS (morpholinopropanesulfonic acid) minimal medium containing radiolabeled $^{32}$P$_{i}$ (see Materials and Methods). Samples were withdrawn before and 10, 30, and 60 min after the addition of IPTG (1 mM) and analyzed by thin-layer chromatography (TLC) and phosphor imaging. (B) Quantification of the results of experiment shown in panel A and of repetitions of the experiment shown in Fig. S8. Materials and Methods gives additional experimental details. Error bars indicate standard deviations of three independent experiments.
induction of relE did not stimulate (p)ppGpp synthesis, showing that inhibition of translation per se is not sufficient to stimulate (p)ppGpp accumulation (Fig. 4A and B).

**DISCUSSION**

In this paper, we describe the discovery of a novel family of bacterial serine/threonine kinases, HipT kinases, that exhibit sequence similarity with HipA of *E. coli* K-12. HipA inhibits GltX (glutamyl-tRNA synthetase) by phosphorylation and thereby triggers RelA-dependent synthesis of (p)ppGpp (11, 31, 32). We found that HipT of *E. coli* O127 phosphorylates and inhibits TrpS (tryptophanyl-tRNA synthetase) and thereby, similarly to HipA, stimulates synthesis of (p)ppGpp (Fig. 4). Even though TrpS and GltX belong to the same class of tRNA synthetases (41), HipT<sub>O127</sub> and HipA do not exhibit cross-phosphorylation of TrpS and GltX *in vitro*, implying that the two kinases exhibit substrate specificity (Fig. 3A and B). We showed previously that HipA phosphorylates S239 of the conserved KLS<sup>239</sup>KR motif in GltX (11). A variant of this motif (KMS<sup>197</sup>KS) is present in TrpS. Even though there are two amino acid differences between the two motifs, they represent the overall highest degree of sequence similarity between the two synthetases, suggesting that HipT phosphorylates S197 of TrpS. Indeed, this proposal was confirmed by our mass spectrometric and mutational analysis of TrpS (Table 1 and Fig. 3C).

We showed previously that phosphorylation of the conserved S239 of GltX by HipA requires the presence of tRNA<sub>Glu</sub> in the *in vitro* reaction mixture (11). We proposed that the binding of tRNA<sub>Glu</sub> to GltX would induce a conformational change of the motif KLS<sup>239</sup>KR that would make S239 accessible to phosphorylation (11). In contrast, even though GltX and TrpS belong to the same class of tRNA synthetases and the structural organization of their active sites is similar (42), phosphorylation of TrpS by HipT<sub>O127</sub> does not require the addition of tRNA (Fig. S6 in the supplemental material). We believe that this difference is consistent with the requirement of GltX for the presence of cognate tRNA to activate glutamate to glutamyl-adenylate (41), a property shared with only two other type I tRNA synthetases (GlnRS and ArgRS). Thus, TrpS does not require the presence of tRNA<sub>Trp</sub> to activate tryptophan to tryptophanyl-adenylate and does not require tRNA<sub>Trp</sub> to be phosphorylated by HipT (Fig. S6).

HipA inactivates itself by autophosphorylation at the fully conserved, essential S150 located adjacent to the P loop of the kinase (35). Structural analysis revealed that autophosphorylation stabilizes a conformation of HipA that disrupts the ATP-binding pocket. It was proposed that autophosphorylation of HipA functions in the resuscitation of cells inhibited by HipA by preventing further activity of available toxins. This explanation is plausible, because cells inhibited by HipA somehow must revert the inhibition of GltX before the cells can resuscitate. We observed that HipT<sub>O127</sub> is autophosphorylated *in vitro* (Fig. 3A and B) at the fully conserved S57 adjacent to the P-loop motif in HipT and, to a minor extent, at S59 in the P-loop motif, both of which are likely to inactivate the enzyme (Table 1).

The *hipT* gene is the third gene of the *hipBST* operon, and HipS and HipT exhibit sequence similarity with either end of HipA. The most parsimonious explanation as to how this arrangement appeared seems to be that *hipA* was duplicated during evolution and split into *hipS* and *hipT*, shifting the kinase specificity during this evolutionary trajectory. Analysis of the structure of HipBA reveals that HipS likely corresponds to the N-terminal subdomain of HipA, which was found to be involved in dimerization during DNA binding, as well as to harbor several mutations associated with high-persister phenotypes (Fig. S9A and B, blue) (26). A more detailed look at the N-terminal subdomain of HipA shows that residues involved in forming the hydrophobic core of the domain are well conserved in HipS, suggesting that HipS and the N-terminal subdomain of HipA share structure, while residues that are involved in HipA-HipA dimerization appear to differ in HipS while being conserved between HipS orthologs. This could suggest that the higher-order structure of HipBST differs from that of HipBA. We also note that several of the known high-persister mutations found in the N-terminal subdomain of HipA (including one of the mutations responsible for the
hipA7 genotype) are naturally present in HipS, which raises the possibility that HipS is HipA7-like (Fig. S9C). Finally, the structural analysis also reveals that HipB (of HipBST) closely matches the corresponding antitoxin HipB in HipBA and likely harbors a DNA-binding domain (Fig. S9A and B). Of the three proteins, the function of HipS as the “third TA component” is clearly the most intriguing. We found that all three HipS orthologs investigated are able to counteract cognate HipT toxins on their own, while the HipB proteins do not have such an effect (Fig. 1). However, in two cases, we observed that the HipB proteins augmented HipS-mediated neutralization of HipT, suggesting that HipB somehow increases the activity of HipS, for example, by increasing HipS metabolic stability or by stabilizing the HipS-HipT interaction. The latter proposal is consistent with the observation that HipBST form a stable complex in vivo (Fig. S4). A summary of our findings is presented in Fig. 5 and described further in the legend to the figure.

Although two-component TA modules are by far the most common, a number of other three-component TA modules have been identified (1). In many of these cases, two adjacent genes exhibit sequence similarity with known type II TA modules, while the function of the third component often remains unclear. However, in a few cases, the function of the third component is known. For example, M. tuberculosis contains a three-gene TA module that encodes a RelE-homologous HigB toxin and the HigA antitoxin. The third gene encodes a SecB-like chaperone that controls the stability of HigA such that the antitoxin becomes metabolically unstable under environmental stress, thereby leading to activation of HigB and inhibition of translation by mRNA cleavage (43). Thus, in this TA module, the third component provides a link between cellular physiology and activation of the TA module. \(\omega-e-\zeta\) of Streptococcus pyogenes is a three-component TA module in which \(\omega\) is a DNA-binding autorepressor of the operon and \(e\) is an antitoxin that neutralizes the \(\zeta\) toxin by direct protein-
protein contact (44). In the paaA-paaA-parA modules of E. coli O157:H7, the first gene encodes a transcriptional regulator of the module operon and the second a type II antitoxin that counteracts the activity of the ParE toxin (45). Thus, our work presented here reveals a novel type of three-component TA modules with unknown regulator properties that will be important and exciting to study. We hope that future biochemical and structural studies will be helpful in revealing the mechanisms of HipBST activation and regulation, as well as why this locus is configured as a three-component TA module.

MATERIALS AND METHODS

Strains, plasmids, media, and growth conditions. Strains and plasmids are listed in Table 2, and DNA oligonucleotides in Table 3. Cultures were grown at 37°C with shaking at 160 rpm in Luria-Bertani (LB) medium. When required, the medium was supplemented with 25 μg/ml chloramphenicol, 30 μg/ml or 100 μg/ml ampicillin, and 25 μg/ml kanamycin. Gene expression from plasmids carrying the pBAD promoter was induced by 0.2% arabinose and repressed by 0.2% glucose. Gene expression from plasmids carrying the synthetic P10::parA promoter was induced by 200 μM isopropyl β-D-1-thiogalactopyranoside (IPTG). The solid medium used to grow cells was Luria-Bertani agar (LB agar) medium supplemented with the appropriate antibiotics and incubated at 37°C for approximately 16 h unless otherwise stated.

Gene knockout by P1 transduction. To construct strain E. coli MG1655ΔydeA, gene knockout was obtained by phage P1 transduction using a strain of the Keio collection as donor according to standard procedure (46, 47).

Multicopy suppression screening. Genomic DNA (gDNA) of E. coli MG1655ΔydeA was purified according to the manufacturer’s instructions (EdgeBio). The gDNA was then partially digested with Sau3AI (Bsp143I) and fragments inserted into pBR322, which had been digested with BamHI and dephosphorylated. The gene library was transformed into a strain harboring the pBAD33:HipTO127 plasmid and plated on agar plates containing arabinose.

Site-directed mutagenesis. Amino acid changes TrpSS197A, TrpSS197D, and HipTO127D were constructed by PCR mutagenesis (Table 3). The PCR products were digested with DpnI, and the resulting plasmids were transformed into E. coli strain DH5α.

Protein purification. HipTO127 (produced by pSVN42) was purified from E. coli strain BL21 that also produced HipO127, and HipO127, pSVN44. Overnight cultures were diluted 100-fold into 350 ml fresh LB medium. At an optical density at 600 nm (OD600) of ~0.3, the toxin gene was induced by the addition of 1 mM IPTG for 4 h, and cells were harvested by centrifugation. Pellets were resuspended in 25 ml cold buffer A (50 mM NaH2PO4 [pH 8], 0.3 M NaCl, 10 mM imidazole, 5 mM β-mercaptoethanol (BME)) with the addition of half a protease inhibitor cocktail each. Cells were carefully sonicated for 5 min at 60% amplification (2 s on and 2 s off) while still kept cold. The cell lysate was spun at 16,000 rpm for 30 min at 4°C, and the cleared lysate was incubated at 4°C for 1 h with 1 ml Ni beads that had been freshly equilibrated in the same buffer for 1 h. Protein-bound beads were then applied to gravity flow columns and washed with 50 ml of buffer B (50 mM NaH2PO4 [pH 8], 0.3 M NaCl, 35 mM imidazole, 1 mM BME).

As described previously, the toxin and antitoxins were separated with urea washes to leave the His-tagged protein on the affinity column (48). His-tagged proteins were purified according to the manufacturer’s protocol, further purified using an Äkta Pure (GE Healthcare) fast protein liquid chromatography (FPLC) instrument, and stored in 200 mM NaCl, 50 mM Tris-HCl, and 5% glycerol. All proteins purified with His tags were tested and compared to wild-type proteins in vivo prior to purification in order to assess their functionality.

Phosphorylation in vitro. Phosphorylation reactions were performed in the presence of 0.05 μl [γ-32P]ATP (6,000 Ci/mmol; Perkin Elmer) per microliter of reaction mixture, 66.6 μM ATP (nonradioactive), and aminoacylation buffer (1 mM dithiothreitol [DTT], 10 mM KCl, 16 μM ZnSO4, and 20 mM MgCl2) for 45 min as described previously (11). Each reaction was stopped by the addition of 1 volume Laemmli loading buffer, the reaction mixture was incubated for 10 min at 95°C, and proteins were resolved by SDS-PAGE and exposed using phosphorimaging (GE Healthcare) overnight.

Phosphorylation in vitro measured by LC-MS. The phosphorylation reaction was performed with 13.5 μM TrpS and 6.7 μM HipTO127 in the presence of 5 mM ATP and aminoacylation buffer for 45 min at 37°C. The reaction was stopped by the addition of 4 volumes of denaturation buffer (6 M urea, 2 M thiourea, 1 mM DTT, and 10 mM Tris-HCl, pH 8.0), and the reaction mixture incubated for 30 min at room temperature, followed by incubation with 5.5 mM iodoacetamide for 45 min at room temperature. Denatured proteins were digested overnight either with endoproteinase Lys-C (1:100 [wt/wt]; Wako) in 20 mM bicine, pH 8.0, or with endoproteinase Arg-C (1:100 [wt/wt]; Roche) in 90 mM Tris-HCl, pH 7.6, 8.5 mM CaCl2, 5 mM DTT, 0.5 mM EDTA. Digested peptides were purified via Pierce C18 Spin Tips, and 0.5 μg of each sample was measured by liquid chromatography-tandem mass spectrometry (LC-MS/MS) as described previously (49). Brieﬂy, peptides were separated by an Easy-nL 1200 ultra-high-performance liquid chromatography (UHPLC) instrument (Thermo Fisher Scientiﬁc) and transferred into an online coupled Q Exactive HF mass spectrometer (Thermo Fisher Scientiﬁc) by nanoelectrospray ionization. Peptides were eluted from a 20-cm-long analytical column packed with 1.9-μm reverse-phase particles using a 33-min segmented gradient of 5% to 50% solvent B (80% [vol/vol] acetonitrile, 0.1% [vol/vol] formic acid) at a constant flow rate of 300 nl/min. Full-scan MS spectra were acquired in a mass range from 300 to 1,650 m/z with a maximum injection time of 45 ms and a resolution of 60,000. Higher-energy collisional dissociation MS/MS scans of the 7 (Top7 data-dependent method) most abundant peaks were...
recorded with a maximum injection time of 220 ms at a resolution of 60,000. Acquired raw data were processed with MaxQuant software (version 1.5.2.8) (50) using default settings if not stated otherwise. The derived peak list was searched against a reference E. coli database (51). Methionine oxidation, protein N-terminal acetylation, and Ser/Thr/Tyr phosphorylation were defined as variable modifications, and carbamidomethylation of cysteines was set as a fixed modification. The maximum number of missed cleavages allowed was set to 3 for the endoproteinase Lys-C and to 2 for Arg-C. Only phosphopeptides with an Andromeda score of >0.75 were considered, and their MS/MS spectra were inspected manually (Fig. S10).

**Measurement of cellular (p)ppGpp levels.** Measurement of cellular (p)ppGpp levels was performed as described previously (52, 53).

### TABLE 2 Bacterial strains and plasmids

<table>
<thead>
<tr>
<th>Strain or plasmid</th>
<th>Description*</th>
<th>Reference or source</th>
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<tr>
<td>E. coli strains</td>
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<td>Wild-type K-12</td>
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<td>MG1655ΔydeA</td>
<td>K-12 MG1655ΔydeA−FRT</td>
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<td>BL21</td>
<td>F−ompT hsdSb (r− m−) gal dcm</td>
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<tr>
<td>C41 (DE3)</td>
<td>Derived from BL21(DE3): F−ompT hsdSb (r− m−) gal dcm</td>
<td>(DE3)</td>
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<td>EG235</td>
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<td>JW239S</td>
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<table>
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<td>pNDM220</td>
<td>Mini-R1 lacZ P_A1/d04/d03, Amp'</td>
<td>56</td>
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<tr>
<td>pCP20</td>
<td>pSC101 rep(Ts), Amp' Cm'</td>
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<tr>
<td>pBR322</td>
<td>pMB1 rop, Amp' Tet'</td>
<td>62</td>
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<td>pMG25</td>
<td>pUC lacZ P_A1/d04/d03, Amp'</td>
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<td>pEG25</td>
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<td>pSVN124</td>
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<td>pSVN135</td>
<td>pBAD33::hipT_0127, optimized SD</td>
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<td>pSVN138</td>
<td>pNMDM20::hipB_0127, optimized SDs</td>
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*SD, Shine-Dalgalno sequence.*
Mass spectrometry. The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE (54) partner repository with the data set identifier PXD012023.

Construction of plasmids. Construction of plasmids is summarized below.

**pKG127.** The region of the *E. coli* O127 E2348/69 genome (accession number NC_011601.1) containing the hipBST locus ([H11001](https://www.ncbi.nlm.nih.gov/nuccore/H11001) 3,948,403 to [H11001](https://www.ncbi.nlm.nih.gov/nuccore/H11001) 3,950,320) was synthesized (GeneScript) and inserted into the SalI restriction site of pUC57.

**pSVN1.** *hipTO127* with start codon GTG was amplified from pKG127 using primers FP1(GTG) and RP1. The resulting PCR product was digested with SalI and SphI and ligated with pBAD33.

**pSVN37.** *trpS* was amplified from pCA24N::*trpS* from the ASKA collection (55) using primers trpS Fw and trpS Rv. The resulting PCR product was digested with BamHI and HindIII and ligated into pEG25.

**pSVN42.** *hipTO127*::His6 was amplified from pKG127 using primers FP13 and RP5. The resulting PCR product was digested with BamHI and HindIII and ligated into pEG25.

**pSVN44.** *hipBSO127* was amplified from pKG127 using primers FP15 and RP6. The resulting PCR product was digested with SalI and SphI and ligated into pBAD33.

**pSVN46.** *trpS* His6 was amplified from pSVN37 using primers trpS Fw and trpS RP3. The resulting PCR product was digested with BamHI and HindIII and ligated into pEG25.

**pSVN49.** The mutation in *trpSS197D* His6 was created using pSVN46 and primers *trpS S197D Fw* and *trpS S197D Rv* in a site-directed plasmid mutagenesis PCR. The fragment was digested with DpnI before being transformed into *E. coli* DH5α.

**pSVN52.** The mutation in *trpSS197A* His6 was created using pSVN46 and primers *trpS S197A Fw* and *trpS S197ARv* in site-directed plasmid mutagenesis PCR. The fragment was digested with DpnI before being transformed into *E. coli* DH5α.

**pSVN94.** His6-tev*hipBO127*::*hipSO127*::*hipTD233Q* with optimized SDs for all three genes was subcloned from pSVN60 by digesting with XbaI and XhoI, purifying the DNA fragment from a 1% agarose gel, and ligating into pET-15b. The mutation in His6-tev*hipBO127*::*hipSO127*::*hipTD233Q* was created using primers hipX D233Q Fw and hipX D233Q Rv in a site-directed plasmid mutagenesis PCR. The fragment was digested with DpnI before being transformed into *E. coli* DH5α.

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### TABLE 3 Oligonucleotides

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<tr>
<th>Oligonucleotide Sequence</th>
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<tr>
<td>FP1(GTG) CCCCCCGTCGACGGATCATGCTCACAGCAGCCGAGCAG</td>
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<td>FP2 CCCCCCGATCGGAAATCCTCAGTACAGGCGCTCAAAAC</td>
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<td>RP15 GGGGGAATTCTTACGGCTTCGACAAAACC</td>
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<td>FP39 CCCCCGGGGATCCTTAAAGGAGGAAAAAAATAGGCCATAGTTGAGGOTCG</td>
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<td>RP6 GGGGGAATTCTTACGGCTTCGACAAAACC</td>
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<td>FP14 GGGGGAATTCTTACGGCTTCGACAAAACC</td>
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<tr>
<td>RP13 GGGGGAATTCTTACGGCTTCGACAAAACC</td>
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May/June 2019 Volume 10 Issue 3 e01138-19 mbio.asm.org

Downloaded from http://mbio.asm.org on August 20, 2019 at Copenhagen University Library
and hipX D233Q Rv in a site-directed plasmid mutagenesis PCR. The fragment was digested with DpnI before transformation.

**pSVN103.** trpS was amplified from pCA24N::trpS from the ASKA collection (55) using primers trpS Fw and trpS RP4. The resulting PCR product was digested with BamHI and EcoRI and ligated into pNDM220.

**pSVN109.** hipSO127 was amplified from pKG127 using primers FP22 and RP14. The resulting PCR product was digested with XhoI and EcoRI and ligated into pNDM220.

**pSVN110.** hipBSO127 was amplified from pSVN61 using primers FP25 and RP14. The resulting PCR product was digested with XhoI and EcoRI and ligated into pNDM220.

**pSVN111.** hipBO127 was amplified from pSVN61 using primers FP25 and RP15. The resulting PCR product was digested with XhoI and EcoRI and ligated into pNDM220.

**pSVN113.** The region of the H. influenzae Rd KW20 genome (NC_000907.1) containing the hipBST locus (H11001710,585 to H11001712,589) was synthesized and inserted into the SalI site of pUC57 (GeneScript).

**pSVN114.** The region of the Tolumonas auensis DSM 9187 genome (NC_012691.1) containing the hipBST locus (H110012,117,168 to H110012,119,170) was synthesized and inserted into the SalI site of pUC57 (GeneScript).

**pSVN116.** hipT0127 was amplified using primers FP5 and RP1 from pSVN1. The fragment was then cloned into cut pNDM220 using BamHI and EcoRI, resulting in pSVN116 (pNDM220::hipT0127).

**pSVN122.** hipBm was amplified from pSVN113 using primers FP29 and RP19. The resulting PCR product was digested with BamHI and KpnI and ligated into pNDM220.

**pSVN123.** hipSm was amplified from pSVN113 using primers FP30 and RP20. The resulting PCR product was digested with BamHI and KpnI and ligated into pNDM220.

**pSVN124.** hipBSm was amplified from pSVN113 using primers FP29 and RP20. The resulting PCR product was digested with BamHI and KpnI and ligated into pNDM220.

**pSVN126.** hipBm was amplified from pSVN114 using primers FP32 and RP22. The resulting PCR product was digested with BamHI and KpnI and ligated into pNDM220.

**pSVN127.** hipSm was amplified from pSVN114 using primers FP33 and RP23. The resulting PCR product was digested with BamHI and KpnI and ligated into pNDM220.

**pSVN128.** hipBSm was amplified from pSVN114 using primers FP32 and RP23. The resulting PCR product was digested with BamHI and KpnI and ligated into pNDM220.

**pSVN129.** hipTm was amplified from pSVN114 using primers FP34 and RP24. The resulting PCR product was digested with XmaI and PstI and ligated into pBAD33.

**pSVN135.** hipTm was amplified from pSVN113 using primers FP39 and RP21. The resulting PCR product was digested with XmaI and PstI and ligated into pBAD33.

**pSVN138.** The optimized SD inserted between hipBm and hipSm was created using pSVN128 and primers FP41 and RP29 in a site-directed plasmid mutagenesis PCR. Eight reactions were carried out at different temperatures with a gradient PCR. The samples were pooled and digested with DpnI to digest the parental plasmid before being transformed into E. coli DH5α.

**pSVN139.** The optimized SD inserted between hipBm and hipSm was created using pSVN124 and primers FP42 and RP30 in a site-directed plasmid mutagenesis PCR. The fragment was digested with DpnI before transformation.

**SUPPLEMENTAL MATERIAL**

Supplemental material for this article may be found at https://doi.org/10.1128/mBio.01138-19.

**FIG S1** , PDF file, 2.2 MB.

**FIG S2** , PDF file, 0.4 MB.

**FIG S3** , PDF file, 0.4 MB.

**FIG S4** , PDF file, 2.9 MB.

**FIG S5** , PDF file, 0.2 MB.

**FIG S6** , PDF file, 1.1 MB.

**FIG S7** , PDF file, 0.4 MB.

**FIG S8** , PDF file, 2.7 MB.

**FIG S9** , PDF file, 0.2 MB.

**FIG S10** , PDF file, 0.2 MB.

**ACKNOWLEDGMENTS**

We thank Alexander Harmes, Biozentrum, Center for Molecular Life Sciences, University of Basel, Switzerland, and Elsa Germain, Laboratoire de Chimie Bactérienne for donation of plasmids, CNRS-Aix Marseille University, Institut de Microbiologie de la Méditerranée, Marseille, France, for the donation of plasmids.

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REFERENCES


