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Acclimation of Biogenic Volatile Organic Compound Emission From Subarctic Heath Under Long-Term Moderate Warming

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Abstract

Biogenic volatile organic compounds (BVOCs) are a diverse group of carbon-based compounds released not only from plants but also from soil (Peñuelas et al., 2014). Many of them (e.g., isoprenoids) are very reactive in the atmosphere, and thus, they play a significant role in tropospheric chemistry (Kulmala et al., 2004; Paasonen et al., 2013). In the atmosphere, BVOCs get rapidly photooxidized with NOx, forming harmful tropospheric ozone (Atkinson, 2000; Laathoworkittukul et al., 2009). In areas where NOx levels are low due to the limited anthropogenic influence, for example, remote Arctic areas, BVOCs react with hydroxyl radicals and therefore deplete the oxidation capacity in the atmosphere, which leads to increase in lifetime of methane, an important greenhouse gas (Di Carlo et al., 2004). Furthermore, some BVOCs can act as precursors for secondary organic aerosols (SOAs) that scatter solar radiation and can function as cloud condensation nuclei, increasing cloudiness (Claeys et al., 2004; Scott et al., 2014). The effects of BVOCs on SOA formation and negative climatic feedback (cooling) are strongest in the northern, remote areas (Paasonen et al., 2013).

The latest estimate of annual global BVOC emissions is 760 Tg C, of which isoprene (C5H8) accounts for 70% and monoterpenes for 11% (Sindelarova et al., 2014). Production and emission of isoprene is light and temperature dependent (Sharkey & Yeh, 2001). Monoterpenes show light and temperature dependency for species lacking storage structures (Staudt & Bertin, 1998), while BVOCs in general follow an exponential temperature dependency (Kesselmeier & Staudt, 1999). With the conservative prediction of global temperature increase of 2°C by 2100 (IPCC, 2013), the annual load of C to the atmosphere due to BVOCs is thus expected to rise. In subarctic and Arctic regions, the rate of increase in air temperature is predicted to be as much as twice the global average (IPCC, 2013), and this highlights the importance of gaining knowledge about BVOCs emitted from plant communities in these regions. Northern ecosystems from subarctic/Arctic wetlands (Faubert, Tiiva, Rinnan, Rasänen, et al., 2010; Holst et al., 2010) to heath tundra (Faubert, Tiiva, Rinnan, Michelsen, et al., 2010; Potosnak et al., 2013; Schollert et al., 2014; Tiiva et al., 2008) have shown significant BVOC emissions. The field experiments mimicking warmer climate have shown a great potential of increase in BVOC emissions from these ecosystems in response to different time spans of moderate warming.
warming (Faubert, Tiiva, Rinnan, Michelsen, et al., 2010; Kramsøj et al., 2016; Lindwall, Schollert, et al., 2016; Lindwall, Svendsen, et al., 2016; Tiiva et al., 2008; Valolahti et al., 2015).

Previous studies have shown that several plant physiological responses can acclimate to higher temperatures. The temperature optimum of photosynthesis was shown to increase under warming by 2–4°C, but photosynthesis at the temperature optimum did not increase (Gunderson et al., 2010). Leaves can acclimate to warming by downregulating respiration rate (Slot & Kitajima, 2015). The emission of isoprene, directly dependent on carbon availability, was lower at higher temperature, when photosynthesis and respiration showed downward acclimation (Fares et al., 2011).

Acclimation also takes place at ecosystem level: respiration is stimulated by warming after the first few years, but it stabilizes after a longer time in the new temperature regime (Rustad et al., 2001). The same acclimation of ecosystem CO$_2$ fluxes to decadal warmer and drier conditions has also been observed in Alaskan Arctic (Oechel et al., 2000). However, it is not known yet if BVOC emissions from Arctic ecosystems can also stabilize in warmer climate since the first measurements in most long-term warming experiments have been done 6–7 years after the setup (Faubert, Tiiva, Rinnan, Michelsen, et al., 2010; Kramsøj et al., 2016; Lindwall et al., 2015; Tiiva et al., 2008). However, results from a leaf-level study of warming effects on BVOC emission from the subarctic and Arctic plants by Schollert et al. (2015) suggest that acclimation may take place: they found that the unaltered emissions in the warming treatment could be partly explained by leaf anatomical acclimations.

In addition to affecting plant metabolism and physiology, warming can also contribute to changes of vegetation composition. In the Arctic, warming increases total plant biomass by promoting especially the growth of taller and deciduous shrub species (Wahren et al., 2005; Walker et al., 2006), while the shading caused by these taller plants diminishes the coverage of ground-level plants, such as lichens and mosses (Elmendorf et al., 2012; Walker et al., 2006). Additionally, studies conducted in tundra and alpine ecosystems (Chapin et al., 2005; Gottfried et al., 2012; Henry & Molau, 1997; Truong, Palme, & Felber, 2007) have demonstrated that changes in species abundance can occur already after a few years of elevated temperature. Our earlier study by Valolahti et al. (2015) showed that vegetation change in response to 13 years of warming partially explained the increased BVOC emissions.

The aims of this study were to reveal if (1) experimental warming for only 3 years increases BVOC emission from a subarctic heath; (2) if shifts in vegetation composition, shown to affect BVOC emissions after 13 years of warming (Valolahti et al., 2015), already occur after warming for a few growing seasons; and (3) whether warming still continues to increase BVOC emissions rates over decadal time span or if the emission of BVOCs stabilizes or diminishes over time. In order to assess the impacts of warming on general ecosystem functioning, we also measured net ecosystem exchange (NEE) and ecosystem respiration ($E_R$). That also allowed us to evaluate if these processes stabilize under long-term warming, as reported for ecosystem respiration (Rustad et al., 2001) and summertime CO$_2$ fluxes (Oechel et al., 2000).

## 2. Materials and Methods

### 2.1. Study and Experimental Design

The experimental site was located on a wet subarctic heath in Abisko, Northern Sweden (68°21′N, 18°49′E, 385 m above sea level), where the long-term warming treatment has been maintained since 1999. Mean annual temperature in the area is around 0°C, and precipitation is 332 mm (2002–2011; Callaghan et al., 2013). Vegetation at the experimental site is dominated by different low shrub species, graminoids, horse-tails, and mosses (Tiiva et al., 2008; Valolahti et al., 2015). Soil characteristics have been presented in detail by Rinnan, Michelsen, and Jonasson (2008). The new short-term warming treatment was established in 2009. The new plots were selected randomly within each block of the experiment. The aim was to assess differences between unmanipulated control (C), open-top chamber summer warming for 3 years (OTC3), and OTC warming for 13 years (OTC13) treatments. Both warming treatments covered an area of 1.2 × 1.2 m and used open-top warming tents made of transparent 0.05 mm thick polythene sheet, while the control plots were of the same size but without tents. Each treatment was replicated in six blocks yielding all in all 18 plots within an area of 1,000 m$^2$. The responses of the OTC13 warming and interactions with litter addition have been earlier presented by Valolahti et al. (2015).
Canopy-level air temperature inside the BVOC measurement chamber was recorded (Hygrochron DS 1923-F5 iButton, Maxim Integrated Products Inc., CA, USA) once per minute during the sampling. Photosynthetically active radiation (PAR) was monitored using PAR sensors (S-LIA-M003, Onset Computer Corporation, Bourne, MA, USA) coupled to a Hobo Micro Station (Onset Computer Corporation). During the BVOC sampling, air temperature inside the flux measurement chambers did not differ more than 0.4°C between control (growing season average 25.4 ± 1.4°C), OTC3 (25.8 ± 1.4°C), and OTC13 (25.6 ± 1.3°C). In the long term, the OTCs increase the canopy air temperature by 0.8°C and the soil temperature at 2 cm depth by 0.7°C (Ravn et al., 2017). The OTCs decreased PAR, on average by 16% (data not shown). PAR, canopy-level air temperature, and soil temperature during the measurement period are shown in Figure 1.

2.2. Vegetation Analysis

Vegetation coverage and species composition were analyzed once during the season, on 10–11 August 2012. The point intercept method described by Jonasson (1988) was used, and the vegetation analyses were made for 0.22 × 0.22 m plots subjected to gas exchange measurements. Plant species were grouped as graminoids, forbs, deciduous shrubs, evergreen shrubs, vascular cryptogams, mosses, and lichens. In addition, the coverage of litter and standing dead biomass was recorded.

2.3. Sampling of BVOCs

Sampling was done 4 times during the growing season in 2012 (Figure 1) using transparent polycarbonate chambers (thickness 0.0015 m, 0.22 × 0.22 m, height 0.2 m; Vink Finland, Kerava, Finland). Chambers were placed on aluminum collars, which had been permanently installed in the plots in the beginning of the experiment. To create an airtight headspace, collar grooves were filled with water before placing the chamber. The air was circulated through the chambers using battery-operated pumps (12 V; Rietschle Thomas, Puchheim, Germany) at 200 mL min⁻¹ for both inflow and outflow, and the chambers were equipped with fans to ensure well-mixed headspace. The incoming air was purified using a charcoal filter (Wilkerson F03-C2–100) to remove particles and volatile impurities and a MnO₂ scrubber to remove ozone (Ortega & Helmig, 2008).

During each sampling time, two blank samples were collected to identify BVOCs emitted from the sampling and analysis equipment following Lindwall, Svendsen, et al. (2016), and the low amount of VOCs in blanks was removed from the data set.

Before sampling, the ambient air inside the chamber was replaced with filtered air by flushing for 10 min with a flow rate of 1,000 mL min⁻¹. After that the BVOCs released from the plots were trapped in stainless steel adsorbent tubes (150 mg Tenax TA, 200 mg Carbograph 1TD, Markes International Limited, Llantrisant, UK) for 30 min. The tubes were sealed with Teflon-coated brass caps and stored at 5°C to prevent fragmentation of collected BVOCs before analysis within 2 weeks from sampling.

2.4. GC-MS Analysis of BVOCs

BVOCs collected in adsorbent tubes were analyzed using a Unity 2 thermal desorber (Markes International Limited, Llantrisant, UK) coupled with an Ultra autosampler and a gas chromatograph-mass spectrometer (7890A Series GC, 5975C inert MSD/DS Performance Turbo El, Agilent Technologies, Santa Clara, CA, USA). After thermodesorption at 250°C for 10 min and cryofocusing at −10°C, the samples were immediately injected, with helium as a carrier gas, into an HP-5 capillary column for separation (length 50 m × diameter 0.2 mm × 0.33 μm film thickness). The oven temperature during the first minute was 40°C, then it was raised to 210°C at a rate of 5°C min⁻¹ and then further to 250°C at a rate of 20°C min⁻¹.
Table 1
Coverage of Plant Functional Groups (% Cover) on 10–11 August (Mean ± SEM, n = 6) in Control, Open Top Chamber Warming for 3 (OTC3) and 13 Years (OTC13) and the Statistical Significance for Differences Among Treatments (ANOVA)

<table>
<thead>
<tr>
<th>Functional Group</th>
<th>Control</th>
<th>OTC3</th>
<th>OTC13</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Graminoids</td>
<td>24.7 ± 4.8</td>
<td>15.3 ± 4.6</td>
<td>24.7 ± 6.4</td>
<td>0.361</td>
</tr>
<tr>
<td>Forbs</td>
<td>5.2 ± 1.4</td>
<td>10.7 ± 4.2</td>
<td>14.5 ± 8.4</td>
<td>0.604</td>
</tr>
<tr>
<td>Deciduous shrubs</td>
<td>26.2 ± 4.7</td>
<td>32.0 ± 8.2</td>
<td>32.7 ± 13.7</td>
<td>0.943</td>
</tr>
<tr>
<td>Evergreen shrubs</td>
<td>47.0 ± 10.1</td>
<td>53.3 ± 10.0</td>
<td>57.2 ± 9.3</td>
<td>0.650</td>
</tr>
<tr>
<td>Vascular cryptogams</td>
<td>4.5 ± 2.6</td>
<td>16.8 ± 3.9</td>
<td>6.3 ± 3.3</td>
<td>0.042</td>
</tr>
<tr>
<td>Total vascular plants</td>
<td>107.5 ± 12.5</td>
<td>128.2 ± 16.1</td>
<td>135.3 ± 20.1</td>
<td>0.061</td>
</tr>
<tr>
<td>Mosses</td>
<td>8.3 ± 4.6</td>
<td>18.7 ± 3.7</td>
<td>10.3 ± 4.3</td>
<td>0.218</td>
</tr>
<tr>
<td>Lichens</td>
<td>7.5 ± 4.5</td>
<td>0.7 ± 0.7</td>
<td>3.5 ± 2.6</td>
<td>0.183</td>
</tr>
<tr>
<td>Litter</td>
<td>15.5 ± 1.9</td>
<td>13.8 ± 2.9</td>
<td>16.0 ± 3.5</td>
<td>0.832</td>
</tr>
<tr>
<td>Standing dead</td>
<td>9.7 ± 1.5</td>
<td>11.8 ± 2.2</td>
<td>10.3 ± 2.8</td>
<td>0.787</td>
</tr>
</tbody>
</table>

Note. Values followed by different superscript letters within a functional group significantly differ from each other based on the pairwise multiple comparisons.

BVOCs were identified using pure standards and according to their mass spectra compared with the NIST library. They were quantified by pure standard solutions for isoprene, α-pinene, camphene, sabinene, 3-carene, limonene, 1,8-cineole, γ-terpinene, copaene, δ-cadinene, aromadendrene, and cis-3-hexenyl acetate (Fluka, Buchs, Switzerland). When quantifying compounds for which no pure standard was available, α-pinene was used for quantification of monoterpenes, copaene for sesquiterpenes, and cis-3-hexenyl acetate for other volatile organic compounds detected. Chromatograms were analyzed using the software Enhanced ChemStation (Agilent Technologies, Santa Clara, CA, USA). Compounds that had an identification quality above 90% in the NIST data library and were present at least in 10% of the samples were accepted in the data set (Faubert, Tiiva, Rinnan, Michelsen, et al., 2010).

BVOC emission rates were expressed on ground area basis (μg m⁻² h⁻¹) (see Faubert et al. (2012) for description of calculations) with chamber headspace volume corrected for microtopographical differences.

2.5. Measurement of CO₂ Exchange

Ecosystem respiration (Eₐ) and net ecosystem exchange (NEE) were measured with an EGM-4 Environmental Gas Monitor (PP Systems, Hitching, UK) connected to an Environmental Monitor Sensor Probe Type 3 (PP Systems, Hitching, UK). A transparent polycarbonate chamber with a volume of 13.5 L and coupling to the EGM4 was placed in the aluminum collar groove, which was filled with water to create an airtight headspace. A fan was used to ensure the mixing of the headspace. NEE was measured continuously for 5 min, then the chamber was lifted to return to ambient CO₂ concentration and Eₐ was measured. For the Eₐ measurement, the chamber was darkened and the fluxes were recorded for 5 min. Gross ecosystem production (GEP) was calculated by summing NEE and Eₐ. Positive fluxes indicate carbon gain by the ecosystem. After each measurement the soil temperature in each plot was measured from 3 points at 2 and 5 cm depth (T-Handle Lab Thermometer DT520TH), and soil moisture was measured using Theta Probe ML2x (Delta T-Devices Ltd, Cambridge, UK) (see Table S2).

2.6. Statistics

Mixed models analysis of variance (ANOVA) with Bonferroni post hoc test was used to analyze for treatment effects on emissions of isoprene, total monoterpenes (MTs), total sesquiterpenes (SQTs), and total other VOCs as well as for Eₐ, GEP, and NEE (IBM SPSS Statistics 23, Armonk, NY, USA). The model included treatment (with three levels: control, OTC3, and OTC13) and date as fixed factors, their interactions, and block as a random factor. To test for the normality of the data and the model residuals, Shapiro-Wilk normality test was used and data were inspected for potential inhomogeneity of variance. Logarithmic or square root transformation was used if needed. P-values < 0.05 were considered as statistically significant and those < 0.1 to indicate marginal significance. Vegetation coverage data were tested for treatment effects using ANOVA when the presumptions of parametric tests were fulfilled and using the nonparametric Kruskall-Wallis one-way ANOVA followed by pairwise multiple comparisons, when needed.

To assess how the treatments had affected the ecosystem, data on CO₂ exchange (NEE, GEP, and Eₐ), VOC emissions (the growing season averages), and the cover of functional groups of vegetation were subjected to a principal component analysis (PCA). The data were mean-centered and standardized to unit-variance. Partial least squares regression (PLSR) was used to analyze the influence of coverage of individual plant species on the emissions of the 10 most emitted individual BVOCs. One-component PLSR models, which were cross-validated using seven cross-validation groups, were extracted separately for each BVOC. The PCA and PLSR analyses were conducted using Simca 13.0.3 (Umetrics, Umeå, Sweden).

3. Results

3.1. Vegetation Coverage

Warming in general increased the coverage of total vascular plants by 19% after 3 years and by 26% after 13 years of exposure, although this was only marginally significant (Table 1). For functional groups
(graminoids, forbs, deciduous shrubs, evergreen shrubs, vascular cryptogams, mosses, and lichens), there were no significant treatment effects except for the coverage of vascular cryptogams, which was a factor of 2.7 higher in the OTC warming for 3 years as compared to the control. At species level, the most abundant vascular plant species in the experimental plots were *Carex vaginata*, *Vaccinium uliginosum*, *Andromeda polifolia*, and *Empetrum hermaphroditum* (Table S1 in the supporting information). The coverage of *A. polifolia* was a factor of 2 higher in the OTC13 treatment compared to the control, while OTC3 treatment did not significantly differ from either of the other treatments. The coverage of *Equisetum arvense* was significantly higher in the OTC3 treatment than in the OTC13 treatment.

### 3.2. BVOC Emission Rates

A total of 31 compounds, isoprene, 7 MTs, 15 SQTs, and 8 other VOCs were detected. Isoprene was the most emitted single compound with emission rates varying from 0 to 335.7 μg m⁻² h⁻¹. The most emitted MT detected in this study was 1,8-cineole (maximum at 6.6 μg m⁻² h⁻¹ and the mean 0.9 ± 0.2 μg m⁻² h⁻¹). The most emitted SQT was β-selinene (maximum at 105.6 μg m⁻² h⁻¹ and the mean 2.9 ± 1.6 μg m⁻² h⁻¹), and for other VOCs, it was toluene (maximum at 7.5 μg m⁻² h⁻¹ and the mean 1.2 ± 0.2 μg m⁻² h⁻¹). Emission rates for the individual compounds emitted on each date and averaged over the season can be found in Table S3.

Warming increased the emission rates of isoprene, MTs, and other VOCs and tended to increase SQT emission (Figure 2). For isoprene emission, the increase compared to the control in the growing season average was by a factor 6.9 for OTC3 and by a factor of 5.6 for OTC13. In the growing season average, there was no significant difference between OTC3 and OTC13 (Figure 2a). There was a significant treatment × date interaction, as the isoprene emission rates varied differently among the treatments throughout the season. MT emissions, averaged over the season, increased under warming, and this difference was significant between control and OTC13 (Figure 2b). Emissions from OTC13 increased by a factor of 3.8 compared to control. No significant difference was found between OTC3 and OTC13. SQT emissions in general had high variations within dates and treatments. There was a marginally significant treatment effect on SQT emission rates (Figure 2c). Emissions from OTC13 increased by factor of 12.7, and those from OTC3 increased by a factor of 11, compared to control. Overall, warming tended to increase the emission rates of the other VOCs in both OTC3 and OTC13 compared to the control, but this factor 1.5- to 1.7-fold increase was not statistically significant. A significant increase in OTC13 compared to control was detected on 28 June (Figure 2d). Overall lower
emission levels were noticeable in MTs, SQTs, and other VOCs on 28 June, when temperature was lower than in other collection dates.

3.3. CO2 Exchange
Both GEP and ER were significantly increased by warming. The two OTC treatments did not differ from each other (Figures 3a and 3b). The difference in GEP was significant between control and OTC13, where fluxes were increased by on average 34% compared to the control. For ER, both OTC3 and OTC13 increased the fluxes significantly compared to control, by 38% and 46%, respectively. Warming did not significantly alter NEE. When averaged over the growing season, the control plots were sinks of CO2, whereas OTC3 was a net source of CO2 and the NEE in OTC13 was close to zero (Figure 3c), although these differences were not statistically significant.

3.4. Correlations Between CO2 Exchange, VOC Emissions, and the Vegetation Coverage
The PCA showed that based on the CO2 exchange, VOC emissions, and the vegetation coverage, the control plots differed from the OTC3 and OTC13 plots, which were similar to each other, and the warmed plots showed more between-plot variation than the controls (Figure 4a). The first two principal components explained 44% of the variance in the data. According to the loading plot, GEP, ER, and NEE correlated positively with the emission of MT and SQT as well as with the coverage of deciduous and evergreen shrubs, vascular cryptogams, and forbs (Figure 4b). The CO2 flux variables correlated negatively with the coverage of litter, standing dead biomass and graminoids. Isoprene emission was negatively correlated with the coverage of evergreen and deciduous shrubs, forbs, and lichens and showed positive correlation with the cover of mosses and graminoids. The emission of other (nonterpenoid) VOCs was also positively correlated with isoprene emission and the coverage of mosses.

The PLSR analysis on the influence of species coverage showed that isoprene emission was positively related to the coverage of *Betula nana*, *Carex pararallela*, and *Salix reticulata* but negatively related to the coverage of *V. uliginosum*, *Astragalus alpinus*, lichens, and *E. hermaphroditum* (Figure 5). The coverage of *B. nana* also positively correlated with the emissions of 1,8-cineole, α-caryophyllene, germacrene, and δ-cadinene (Figures S1 and S2 in the supporting information). The coverage of *Rhododendron lapponicum* positively correlated with the emission of β-selinene (Figure S2), and that of graminoids, such as *C. pararallela* and *Festucia ovina*, correlated with toluene emission (Figure S3).

4. Discussion
Our results show that already after 3 years of air temperature increase, the OTC treatment clearly increased the BVOC emissions from the subarctic heath. The increase in BVOC emissions in response to warming would be expected because both the biosynthesis of the compounds and the diffusion of compounds from storage pools are temperature-dependent (Loreto & Schnitzler, 2010). However, we also show that there were no differences in BVOC emissions between OTC treatments carried out for 3 and 13 years. This suggests that a relatively short-term warming exposure is enough to cause an increase and that this increase remains high over a continuous exposure of 10 additional years, without a further significant increase.
The increase in BVOC emissions, including isoprene, MTs, SQTs, and other VOCs, was well in agreement with earlier reports from the same location (Faubert, Tiiva, Rinnan, Michelsen, et al., 2010; Tiiva et al., 2008; Valolahti et al., 2015), and with the suggested high temperature dependency of subarctic BVOC emissions (Holst et al., 2010; Potosnak et al., 2013; Tang et al., 2016). In general, direct and physiological responses of BVOC emissions can occur rapidly after the plant experiences an increase in temperature, although isoprene and monoterpene production are known to acclimate to warmer conditions within days (Niinemets et al., 2010). In field experiments this effect is not as straightforward as plants can have delayed responses to warming, and/or interactions with other biotic or abiotic factors, which also influence BVOC emissions (Niinemets et al., 2010; Sharkey et al., 1999).

In the present study, the increased BVOC emissions were most likely partly a result of increased vegetation coverage and a shift in vegetation composition. The total vascular plant coverage showed an increasing trend from the control to 3 years and further to 13 years of OTC warming. An increase in the coverage of vascular cryptogams had already manifested after 3 years of OTC warming. These kinds of rapid composition shifts in Arctic plant communities caused by warming over a few growing seasons have also been observed at 11 locations across the tundra biome (Walker et al., 2006).

An increased abundance of deciduous and evergreen shrubs and forbs as observed in response to recent warming in the Arctic (Elmendorf et al., 2012) could contribute to an increase in BVOC emissions. According to the PCA on our data, the coverage of vascular cryptogams, shrubs, and forbs positively correlated with MT and SQT emissions. The drastically increased MT (three- to fourfold higher season averages compared to control) and SQT (11- to 13-fold higher season averages compared to control) emissions were probably related to increased abundances of many terpenoid emitters in both the OTC treatments. The PLSR analyses of the dependence of emission of the most emitted individual BVOCs on the plant species coverage showed more fine-scaled dependencies. Of the dominant shrubs, *B. nana* coverage was positively correlated with emissions of isoprene, and several of the most emitted mono- and sesquiterpenes, and *B. nana* is often found to increase in abundance and growth with warmer climate (Hollesen et al., 2015; Sistla et al., 2013). *B. nana* is not an isoprene emitter itself (Vedel-Petersen et al., 2015), and the correlation between the *B. nana* coverage and isoprene emission is likely due to the presence of co-occurring isoprene-emitting species or another factor correlating with the *B. nana* coverage. The coverage of the dwarf shrub *A. polifolia* was significantly increased in the OTC13 treatment, and this species is known as monoterpene and sesquiterpene emitter (Rinnan et al., 2005). The coverages of both *A. polifolia* and *B. nana* were positively related to the emission of SQTs such as δ-cadinene. As both plant coverage and compound emission are likely to increase under warming, an altered climate will modify the blend of volatiles from tundra in a warmer climate.

The emissions of nonterpenoid VOCs were also increased by warming although not as drastically as those of terpenoids. The compounds of this group, for example, toluene, benzene, and 2-methylfuran, have been earlier reported in emissions from soil and litter (Gray et al., 2010; Huber et al., 2010; Leff & Fierer, 2008). In our experimental setup and ecosystem plot-level measurements, we are unable to precisely identify the source of the emitted BVOCs, but the nature of these compounds and the lower temperature-dependency suggests that they could originate from soil and litter.

We also measured ecosystem CO$_2$ fluxes in order to assess whether the impacts of warming on these follow the same trend as for the BVOC emissions. GEP and $E_P$ increased under OTC warming, which is in agreement
with results from several North American warming experiments (Hobbie & Chapin, 1998; Oberbauer et al., 2007; Welker et al., 2004) and a Greenlandic study (Marchand et al., 2004). GEP represents total photosynthetic carbon uptake in light by vegetation present in the measured ecosystem plots, while $E_R$ includes both dark respiration of vegetation and CO2 released from microbial decomposition in soils. According to the PCA, GEP and $E_R$ appear to be linked to the increased coverage of shrubs, vascular cryptogams, and forbs. Thus, similarly to BVOC emissions, CO2 exchange appears to be largely controlled by vegetation changes in the studied ecosystem. We suggest that part of the increased BVOC emissions may be due to the higher photosynthesis and the concomitant increased carbon allocation to BVOC synthesis under warming. The ongoing vegetation changes also seem to override any sign of treatment effects decreasing over time, a response acclimation pattern that has been earlier shown for soil respiration in Alaskan moist/wet tundra (Oechel et al., 2000). No significant change was observed in NEE, because both GEP and $E_R$ increased under warming and canceled out the effects on NEE. However, the magnitude of the increase in response to the moderate warming was much less for GEP and $E_R$ than for the emissions of most BVOC groups, only up to 46% compared to an increase by a factor of 3–13. This highlights the increasing importance of accounting for the emissions of BVOCs in the Arctic.

No significant changes were observed in BVOC emissions, GEP, or $E_R$ between 3 and 13 year long exposures to OTC warming on this subarctic heath. This suggests that after the relatively rapidly developing initial responses, which had already taken place after 3 years, the ecosystem acclimates to the new temperature regime, leading to a slower rate of change. This observed acclimation could be related not only to the physiological acclimation to the changes in growth conditions, like nutrient status or neighboring species (Oechel et al., 2000), but also to leaf anatomical adaption (Schollert et al., 2015). Our study does not allow for identification of other acclimation or adaptation responses, such as plant physiological responses to new environmental conditions or leaf anatomical changes.

In this study, the passive warming by OTCs only produced a 0.8°C long-term temperature increase. The predicted increase by 2°C by the year 2100 (IPCC, 2013) is likely to cause even larger alterations in the vegetation.
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