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A Generative Angular Model of Protein Structure Evolution

Michael Golden,*1 Eduardo García-Portugués,2,3,5 Michael Sørensen,3 Kanti V. Mardia,1,4 Thomas Hamelryck,5,6 and Jotun Hein1

1Department of Statistics, University of Oxford, Oxford, United Kingdom
2Department of Statistics, Carlos III University of Madrid, Madrid, Spain
3Department of Mathematical Sciences, University of Copenhagen, Copenhagen, Denmark
4Department of Mathematics, University of Leeds, Leeds, United Kingdom
5Bioinformatics Centre, Section for Computational and RNA Biology, Department of Biology, University of Copenhagen, Copenhagen, Denmark
6Image Section, Department of Computer Science, University of Copenhagen, Copenhagen, Denmark

*Corresponding author: E-mail: golden@stats.ox.ac.uk.
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Protein families were obtained from the HOMSTRAD database.

Abstract

Recently described stochastic models of protein evolution have demonstrated that the inclusion of structural information in addition to amino acid sequences leads to a more reliable estimation of evolutionary parameters. We present a generative, evolutionary model of protein structure and sequence that is valid on a local length scale. The model concerns the local dependencies between sequence and structure evolution in a pair of homologous proteins. The evolutionary trajectory between the two structures in the protein pair is treated as a random walk in dihedral angle space, which is modeled using a novel angular diffusion process on the two-dimensional torus. Coupling sequence and structure evolution in our model allows for modeling both “smooth” conformational changes and “catastrophic” conformational jumps, conditioned on the amino acid changes. The model has interpretable parameters and is comparatively more realistic than previous stochastic models, providing new insights into the relationship between sequence and structure evolution. For example, using the trained model we were able to identify an apparent sequence–structure evolutionary motif present in a large number of homologous protein pairs. The generative nature of our model enables us to evaluate its validity and its ability to simulate aspects of protein evolution conditioned on an amino acid sequence, a related amino acid sequence, a related structure or any combination thereof.

Key words: evolution, protein structure, probabilistic model, directional statistics.

Introduction

Recently, several studies (Challis and Schmidler 2012; Herman et al. 2014) have proposed joint stochastic models of evolution which take into account simultaneous alignment of protein sequence and structure. These studies point out the limitations of earlier non-probabilistic methods, which often rely on heuristic procedures to infer parameters of interest. A major disadvantage of using heuristic procedures is that they typically fail to account for sources of uncertainty. For example, relying on a single fixed alignment, which is highly unlikely to be the true underlying alignment, may bias the inference of the posterior distribution over evolutionary trees.

We present a generative evolutionary model, ETDBN (Evolutionary Torus Dynamic Bayesian Network) for pairs of homologous proteins. ETDBN captures dependencies between sequence and structure evolution, accounts for alignment uncertainty, and models the local dependencies between aligned sites.

A key step in modeling protein structure evolution is selecting a suitable structural representation and corresponding evolutionary model. Early works by Gutin and Badredtinov (1994) and Grishin (1997) represented protein structure using three-dimensional Cartesian coordinates of protein backbone atoms and used diffusion processes to model the relationship between structural distance (measured using RMSD) and sequence similarity. More recent publications by Challis and Schmidler (2012) and Herman et al. (2014) likewise used the three-dimensional Cartesian coordinates of amino acid Cα atoms to represent protein structure and additionally used Ornstein-Uhlenbeck (OU) processes to construct Bayesian probabilistic models of protein structure evolution. These models emphasize estimation of evolutionary parameters such as the evolutionary time between species, tree topologies and alignment, and attempt to fully account for sources of uncertainty. For the sake of computational tractability, the aforementioned approaches treat the Cartesian coordinates associated with atoms as evolving independently of another. A non-probabilistic approach by Echave (2008) and Echave and Fernández (2010) referred to as the Linearly Forced Elastic Network Model (LFENM) treats protein structures as a collection of Cα atoms connected by spring forces. The major
benefit of LFENMs is that they do not assume independence of atomic coordinates and take into account non-local dependencies due to physical interactions. In their current formulation LFENMs do not distinguish between the differing chemical nature of different amino acids and therefore do not account for the variable effect of sequence mutation on protein structure evolution.

Rather than using a Cartesian coordinate representation, our model, ETDBN, uses a dihedral angle representation motivated by the non-evolutionary TorusDBN model (Boomsma et al. 2008, 2014). TorusDBN represents a single protein structure as a sequence of \((\phi, \psi)\) dihedral angle pairs, which are modeled using continuous bivariate angular distributions (Frellsen et al. 2012). Likewise, ETDBN treats protein structure as a random walk in space, again making use of the \(\phi\) and \(\psi\) dihedral angles (top of fig. 1).

The dihedral angle representation is informed by the chemical nature of peptide bonds. Each amino acid in a protein peptide chain is covalently bonded to the next via a peptide bond. Peptide bonds have a partial double bond nature that results in a planar configuration of atoms in space. This configuration allows the protein backbone structure to be largely described in terms of a series of \(\phi\) and \(\psi\) dihedral angles that defines the relationship between the planes in three-dimensional space. A benefit of this representation is that it bypasses the need for structural alignment, unlike in models on Cartesian coordinates which typically need to additionally superimpose the structures for comparison purposes (Herman et al. 2014). Accordingly, having to account for superimposition introduces an additional source of uncertainty. A further advantage of the dihedral angle representation is that there are fewer degrees of freedom per amino acid and therefore typically fewer parameters required in order to model their evolution.

The evolution of dihedral angles in ETDBN is modeled using a novel stochastic diffusion process developed in García-Portugués et al. (2017). In addition to this, a coupling is introduced such that an amino acid change can lead to a \textit{jump} in dihedral angles and a change in diffusion process, allowing the model to capture changes in amino acid that are directionally coupled with changes in dihedral angle or secondary structure. As in Challis and Schmidler (2012) and Herman et al. (2014), the insertion and deletion (indel) evolutionary process is also modeled in order to account for alignment uncertainty (Thorne et al. 1992).

The OU processes used in Challis and Schmidler (2012) and Herman et al. (2014) ignore bond lengths and treat \(C_\alpha\) atoms as evolving independently for the sake of computationally tractability. Furthermore, the OU process makes Gaussian assumptions. From a generative perspective these properties will lead to evolved proteins with \(C_\alpha\) atoms that are unnaturally dispersed in space. Bond lengths are also ignored in ETDBN, but can be plausibly fixed or modeled. As a result, it is expected that the use of angular diffusions will much more naturally capture the underlying protein structure manifold.

Two or more homologous proteins will share a common ancestor, which leads to underlying tree-like dependencies. These dependencies manifest themselves most noticeably in the degree of amino acid sequence similarity between two homologous proteins. The strength of these dependencies is assumed to be a result of two major factors: the time since the common ancestor and the rate of evolution.

Failing to account for evolutionary dependencies can lead to false conclusions (Felsenstein 1985), whereas accounting for evolutionary dependencies allows information from homologous proteins to be incorporated in a principled manner. This can lead to more accurate inferences, such as the prediction of a protein structure from a homologous protein sequence and structure, known as homology modeling (Arnold et al. 2006). Stochastic models such as ETDBN are not expected to compete with homology modeling software such as SWISS-MODEL (Arnold et al. 2006). However, they allow for estimation of evolutionary parameters and statements about uncertainty to be made in a statistically rigorous manner.

Most models of structural evolution ignore dependencies amongst sites because of the increased computational demand and complexity associated with such models. These dependencies are expected to influence patterns of evolution, specifically patterns of amino acid substitution. The current model deals with local dependencies only—dependencies that are expected to arise due to interactions between neighboring amino acids, for example, between amino acids in an \(a\)-helix. ETDBN does not account for global dependencies—dependencies that result in the globular nature of proteins (Boomsma et al. 2008). In ETDBN, we attempt to model local dependencies only by using a Hidden Markov Model (HMM) to capture dependencies amongst neighboring aligned positions. HMMs such as PASSML (Li et al. 1998) have been successfully used to predict protein secondary structure from aligned sequences, however, these models typically have the disadvantage that they assume a canonical secondary structure shared amongst all the sequences being analyzed. This restricts analysis to closely related sequences where conservation of secondary structure is a reasonable assumption. ETDBN does not assume a canonical secondary structure, but instead uses a phylogenetic HMM approach, similar to Siepel and Haussler (2004), that assumes dependencies between evolutionary processes at neighboring aligned positions.

Parameters of ETDBN were estimated using 1,200 homologous protein pairs from the HOMSTRAD database (Mizuguchi et al. 1998). The resulting model provides a realistic prior distribution over proteins and protein structure evolution in comparison to previous stochastic models. Doing so enables biological insights into the relationship between sequence and structure evolution, such as patterns of amino acid change that are informative of patterns of structural change (Grishin 2001). It was with these features in mind that ETDBN was developed.

**Evolutionary Model**

**Overview**

ETDBN is a dynamic Bayesian network model of local protein sequence and structure evolution along a pair of aligned homologous proteins \(p_a\) and \(p_b\). ETDBN can be can be viewed as an HMM (see fig. 1). Each hidden node of the HMM,
corresponding to an aligned position, adopts an evolutionary hidden state specifying a distribution over three different observation pairs: a pair of amino acid characters, a pair of dihedral angles and a pair of secondary structures classifications. A transition probability matrix specifies neighboring dependencies between adjacent evolutionary states. For example, transitions along the alignment between hidden states encoding predominantly $\alpha$-helix evolution would be expected to occur more frequently than transitions between an evolutionary hidden state encoding predominantly $\alpha$-helix evolution and another encoding predominantly $\beta$-sheet evolution.

Ideally, the underlying hidden states would not just vary across the length of the alignment as captured by the HMM in the current model, but also evolve along the branches of the phylogenetic tree. This remains computationally intractable at present. Allowing the hidden states to evolve along the tree would allow capturing large structural changes, even induced by a single mutation. For now we model such events using a jump model (see below).

Partially in order to mitigate this, each hidden state specifies a distribution over a pair of site-classes at each aligned position. This gives rise to the possibility of a ‘jump event’. A jump event allows a large change in dihedral angle or secondary structure (e.g. helix to sheet) to occur at a given aligned position and also introduces a directional coupling between changes in amino acid that are informative of changes in dihedral angle or secondary structure conformation.

**Observation Types**

The two proteins, $P_a$ and $P_b$, in a homologous pair are associated with a pair of observation sequences $O_a$ and $O_b$ obtained from experimental data, respectively. An $i$th site observation pair, $O_i = (O_a(i), O_b(i))$, is associated with every aligned site $i$ in an alignment $M_{ab}$ of $P_a$ and $P_b$ where $M_{ab}$ is the homology relationship at position $i$ of the alignment (homologous, deletion with respect to $P_a$, and insertion with respect to $P_b$, respectively), $i$ is taken to run from $1$ to $m$, $m$ is the length of the alignment $M_{ab}$ and $x \in \{i, j, k\}$ and $y \in \{a, b, c\}$ specify the indices of the positions in $P_a$ and $P_b$, respectively. $|P_a|$ and $|P_b|$ give the number of sites in $P_a$ and $P_b$ respectively.

Each site observation, $O_a(i)$ and $O_b(i)$, contains amino acid and structural information corresponding to the two $C_a$ atoms at aligned site $i$ belonging to each of the two proteins. A site observation corresponding to a particular protein at aligned site $i$, $O_a(i)$, is comprised of three different data types:
associated with the Cα atom: an amino acid (Aα); discrete, one of twenty canonical amino acids, φ and ψ dihedral angles (Xα = (φα, ψα)); continuous, bivariate, and a secondary structure classification (Sα); discrete, one of three classes: helix (H), sheet (S), or coil (C). Therefore, Oα = (Aα, Xα, Sα) and Oβ = (Aβ, Xβ, Sβ).

Model Structure
The sequence of hidden nodes in the HMM is written as \( H = (H^1, H^2, \ldots, H^m) \). Each hidden node \( H^i \) in the HMM corresponds to a site observation pair, \( O^i_{\alpha} \) and \( O^i_{\beta} \), at an aligned site \( i \) in the alignment \( M_{\alpha\beta} \). Initially we treat the alignment \( M_{\alpha\beta} \) as given a priori, but later modify the HMM to marginalize out an unobserved alignment.

The model is parameterized by \( h \) hidden states. Every hidden node \( H^i \) corresponding to an aligned site \( i \) takes an integer value from 1 to \( q \) for the hidden state at node \( H^i \). In turn, each hidden state specifies a distribution over a site-class pair: \((r^i, r^j)\) as a function of evolutionary time. A site-class pair consists of two site-classes: \( r^i \) and \( r^j \). Each of the two site-classes takes an integer value 1 or 2, that is, \((r^i, r^j) \in \{(1,1),(1,2),(2,1),(2,2)\}\). We return to the specific role of the site-classes in the next sections.

The state of \( H \) together with the site-class pair, \((r^i, r^j)\), and the evolutionary time separating proteins \( p_{\alpha\beta} \) and \( p_{\beta\alpha} \), specify a distribution over three conditionally independent stochastic processes: \( A^i = (A^i_{\alpha\beta}, A^i_{\beta\alpha}) \), \( X^i = (X^i_{\alpha\beta}, X^i_{\beta\alpha}) \) and \( S^i = (S^i_{\alpha\beta}, S^i_{\beta\alpha}) \). This conditional independence structure allows the likelihood of a site observation pair at an aligned site \( i \) to be written as follows:

\[
p(O^i_{\alpha}, r^i_{\alpha}, r^j_{\alpha}, t_{\alpha\beta}) = p(A^i_{\alpha\beta}, r^i_{\alpha}, r^j_{\alpha}, t_{\alpha\beta}) \times p(X^i_{\alpha\beta}, r^i_{\alpha}, t_{\alpha\beta}) \times p(S^i_{\alpha\beta}, r^i_{\alpha}, r^j_{\alpha}, t_{\alpha\beta}).
\]

The assumption of conditional independence provides computational tractability, allowing us to avoid costly marginalization when certain combinations of data are missing (e.g. amino acid sequences present, but secondary structures and dihedral angles missing).

Stochastic Processes: Modeling Evolutionary Dependencies
Each site-class couples together three time-reversible stochastic processes that separately describe the evolution of the three pairs of observation types, as in equation (1). Each site-class is intended to capture both physical and evolutionary features pertaining to sequence and structure. Parameters that correspond to a particular site-class are termed site-class specific, whereas parameters that are shared across all site-classes are termed global. The use of site-class specific parameters, such as site-class specific amino acid frequencies and dihedral angle diffusion parameters, as described in the next section, is intended to model site-specific physical–chemical properties (Halpern and Bruno 1998; Koshi and Goldstein 1998; Lartillot and Philippe 2004).

Amino Acid Evolution
As is typical with models of sequence evolution, amino acid evolution, \( p(A^i_{\alpha\beta}, A^j_{\beta\alpha} | H^i, r^i_{\alpha}, r^j_{\alpha}, t_{\alpha\beta}) \), is described by a Continuous-Time Markov Chain (CTMC). Each amino acid CTMC is parameterized in the following way: the exchangeability of amino acids is described by a \( 20 \times 20 \) symmetric global exchangeability matrix \( S \) (190 free parameters; Whelan and Goldman 2001), a site-class specific set of 20 amino acid equilibrium frequencies \( \Pi^i_H = \text{diag} \{ \pi_1, \pi_2, \ldots, \pi_{20} \} \) (19 free parameters per site-class) and a site-class specific scaling factor \( \Lambda^i_H \) (one free parameter per site-class). Together these parameters define a site-class specific time-reversible amino acid rate matrix \( Q^i_H = \Lambda^i_H \Pi^i_H \). The stationary distribution of \( Q^i_H \) is given by the amino acid equilibrium frequencies: \( \Pi^i_H \).

Secondary Structure Evolution
Secondary structure evolution, \( p(S^i_{\alpha\beta}, p_{\alpha\beta}, r_{\alpha\beta}, t_{\alpha\beta}) \), is also described by a CTMC. For the sake of simplicity we use only three discrete classes to describe secondary structure at each position: helix (H), sheet (S), and random coil (C).

The exchangeability of secondary structure classes at a position is described by a \( 3 \times 3 \) symmetric global exchangeability matrix \( V \) and a site-class specific set of three secondary structure equilibrium frequencies \( \Omega^i_H = \text{diag} \{ \pi_1, \pi_2, \pi_3 \} \). Together they define a site-class specific time-reversible secondary structure rate matrix \( R^i_H = VR^i_H, \) with stationary distribution: \( \Omega^i_H \).

Dihedral Angle Evolution
Central to our model is evolutionary dependence between dihedral angles, \( p(X^i_{\alpha\beta}, X^j_{\beta\alpha} | H^i, r^i_{\alpha}, r^j_{\alpha}, t_{\alpha\beta}) \). Typically, the continuous-time evolution of the continuous-state random variables is modeled by a diffusive process such as the OU process, as in Challis and Schmidler (2012). However, an OU process is not appropriate for dihedral angles as they have a natural periodicity. For this reason, a bivariate diffusion that captures the periodic nature of dihedral angles, the Wrapped Normal (WN) diffusion, was specifically developed for this paper in García-Portugués et al. (2017).

Topologically, the WN diffusion (see fig. 2 for a pictorial example) can be thought of as the analogue of the OU process on the torus \( T^2 = [-\pi, \pi] \times [-\pi, \pi] \). The WN diffusion arises as the wrapping on \( T^2 \) of the following Euclidean diffusion:

\[
dX_t = \sum_{k \in Z^2} \mu - X_t - 2k\pi)w_k(X_t) \, dt + \Sigma^i dW_t,
\]

where \( W_t \) is the two-dimensional Wiener process, \( \mu \) is the drift matrix, \( \mu \in T^2 \) is the stationary mean, \( \Sigma \) is the infinitesimal covariance matrix and
is a probability density function (pdf) for $k \in \mathbb{Z}^2$. $\phi_k$ stands for the pdf of a bivariate Gaussian $\mathcal{N}(0, \Sigma)$. The pdf (eq. 3) weights the linear drifts of equation (2) such that they become smooth and periodic.

It is shown in García-Portugués et al. (2017) that the stationary distribution of the OU diffusion is a WN($\mu, \Sigma$), which has pdf:

$$p_{WN}(\theta | \mu, \Sigma) = \sum_{k \in \mathbb{Z}^2} \phi_k(\theta - \mu + 2k\pi).$$ (4)

Despite involving an infinite sum over $\mathbb{Z}^2$, taking just the first few terms of this sum provides a tractable and accurate approximation to the stationary density for most of the realistic parameter values.

Maximum Likelihood Estimation (MLE) for diffusions is based on the transition probability density (tpd), which only has a tractable analytical form for very few specific processes. A highly tractable and accurate approximation to the tpd is given for the WN diffusion. This approximation results from weighting the tpd of the OU process in the same fashion as the linear drifts are weighted in equation (2), yielding the following multimodal pseudo-tpd:

$$\tilde{p}(\theta_2 | \theta_1, A, \mu, \Sigma, t) = \sum_{m \in \mathbb{Z}^2} p_{WN}(\theta_2 | \mu_m, \Gamma_m) \omega_m(\theta_1),$$ (5)

with $\theta_1, \theta_2 \in \mathbb{T}^2$, $\mu_m = \mu + e^{-tA}(\theta_1 - \mu + 2\pi m)$ and $\Gamma_t = \int_0^t e^{-sA} \Sigma e^{-sA} ds$. The pseudo-tpd provides a good approximation to the true tpd in key circumstances: 1) $t \to 0$, since it collapses in the Dirac delta; 2) $t \to \infty$, since it converges to the stationary distribution; 3) high concentration, since the WN diffusion becomes an OU process. Furthermore, it is shown in García-Portugués et al. (2017) that the pseudo-tpd has a lower Kullback–Leibler divergence with respect to the true tpd than the Euler and Shoji-Ozaki pseudo-tpds, for most typical scenarios and discretization times in the diffusion trajectory.

A further desirable property of the pseudo-tpd is that it obeys the time-reversibility equation, which in terms of $(X^{\varphi(i)}_b, X^{\psi(i)}_b)$ is

$$\tilde{p}(X^{\varphi(i)}_b | X^{\psi(i)}_a; A, \mu, \Sigma, t_{ab}) p_{WN}(X^{\psi(i)}_a | \mu, \frac{1}{2} A^{-1}\Sigma)$$

$$= \tilde{p}(X^{\psi(i)}_a | X^{\varphi(i)}_b; A, \mu, \Sigma, t_{ab}) p_{WN}(X^{\varphi(i)}_b | \mu, \frac{1}{2} A^{-1}\Sigma).$$

Indeed, the WN diffusion is the unique time-reversible diffusion with constant diffusion coefficient and stationary pdf (eq. 4), in the same way the OU is with respect to a Gaussian. Time-reversibility is an assumption of the overall model and many other models of sequence evolution. A benefit of time-reversibility in a pairwise model such as ETDBN is that one of the proteins in a pair may be arbitrarily chosen as the ancestor, thus avoiding a computationally expensive marginalization of an unobserved ancestor.

The likelihood of a dihedral angle observation pair $(X^{\varphi(i)}_a, X^{\psi(i)}_b)$, assuming that $X^{\psi(i)}_a$ is drawn from the stationary distribution, is given by:

$$p(X^{\varphi(i)}_a, X^{\psi(i)}_b | [H, r_a, r_b, t_{ab}])$$

$$= p(X^{\varphi(i)}_a, X^{\psi(i)}_b | A, \mu, \Sigma, t_{ab})$$

$$\approx \tilde{p}(X^{\varphi(i)}_b | X^{\psi(i)}_a; A, \mu, \Sigma, t_{ab}) p_{WN}(X^{\psi(i)}_a | \mu, \frac{1}{2} A^{-1}\Sigma),$$ (6)

$A$ and $\Sigma$ are constrained to yield a covariance matrix $A^{-1}\Sigma$. A parameterization that achieves this is $\Sigma = \text{diag}(\sigma_1^2, \sigma_2^2)$ and $A = (\alpha_1, \alpha_2, \alpha_3, \alpha_2, \alpha_2)$. $\alpha_1, \alpha_2, \alpha_2$ and $\alpha_2$ are the drift components for the $\varphi$ and $\psi$ dihedral angles, respectively. Dependence (correlation) between the dihedral angles is captured by $\alpha_3$. A depiction of a WN diffusion with given drift and diffusion parameters is shown in figure 2.

**Site-Classes: Constant Evolution and Jump Events**

We now turn to the meaning of the site-class pairs. Two modes of evolution are modeled: constant evolution and jump events. Constant evolution occurs when the site-class starting in protein $p_a$ at aligned site $i$, $r^a_i$, is the same as the site-class ending in protein $p_b$ at aligned site $i$, $r^b_i$, that is, $r^a_i = r^b_i$. Thus the distribution over observation pairs at a site is specified by a single site-class.

As already stated, a site-class specifies the parameters of the three conditionally independent stochastic processes describing amino acid, dihedral angle, and secondary structure evolution. A limitation of “constant evolution” is that the coupling between the three stochastic processes is somewhat weak. This in part stems from the time-reversibility of the stochastic processes—swapping the order of one of the three observation pairs at a homologous site, e.g. (Glycine, Proline) instead of (Proline, Glycine), does not alter the likelihood in equation (1). Alternatively restated from a generative perspective: a ‘directional coupling’ of an amino acid interchange does not inform the direction of change in dihedral angle or secondary structure. For example, replacing a glycine in an $\alpha$-helix in one protein with a proline at the homologous position in a second protein would be expected to break the $\alpha$-helix in the second protein and to strongly inform the plausible dihedral angle conformation of the second protein.

Ideally, we would consider a model in which the underlying site-classes were not fixed over the evolutionary trajectory separating the two proteins, as in the case of constant evolution as described above, but instead were able to ‘evolve’ in time. This would allow occasional switches in the underlying site-class at a particular homologous site, which would create a stronger dependency between amino acid, dihedral angle and secondary structure evolution, that furthermore captures the directional coupling we desire. Such an approach is considered computationally intractable due to the introduction of context-dependence when having to consider neighboring dependencies amongst evolutionary trajectories at adjacent sites.

In order to approximate this ‘ideal’ model in a computationally efficient manner we introduce the notion of a jump
A jump event occurs when \( r'_a \neq r'_b \). Whereas constant evolution is intended to capture angular drift (changes in dihedral angles localized to a region of the Ramachandran plot), a jump event is intended to create a directional coupling between amino acid and structure evolution, and is also expected to capture angular shift (large changes in dihedral angles). The hidden state at node \( H' \), together with the evolutionary time \( t_{ab} \), separating proteins \( p_a \) and \( p_b \), specifies a joint distribution over a site-class pair:

\[
p( r'_a | r'_b, H', t_{ab} ) = p( r'_a | H', r'_b, t_{ab} ) p( r'_b | H' ), \tag{7}
\]

where

\[
p( r'_a | H', r'_b, t_{ab} ) = \begin{cases} 
  e^{-\gamma_{ab} t_{ab}} + \pi_{ab} r'_a (1 - e^{-\gamma_{ab} t_{ab}}), & \text{if } r'_a = r'_b, \\
  \pi_{ab} r'_b (1 - e^{-\gamma_{ab} t_{ab}}), & \text{if } r'_a \neq r'_b,
\end{cases}
\]

and \( p( r'_a | H' ) = \pi_{ab} r'_a \) and \( p( r'_b | H' ) = \pi_{ab} r'_b \), \( \pi_{ab} < 1 \) and \( \gamma_{ab} > 0 \) are model parameters specifying the probability of starting in site-class \( r'_a \) or \( r'_b \) respectively, corresponding to the hidden state specified by node \( H' \). A jump event is intended to capture angular drift (changes in dihedral angles for both proteins \( (A_a, A_b, X_a, X_b) \). Homologous sites corresponding to a single hidden state and with evidence of a jump event (\( r'_a \neq r'_b \)) at posterior probability > 0.90 were identified, that is, the \( i \)'s such that \( p(H', r'_a \neq r'_b | A_a, A_b, X_a, X_b) > 0.90 \).

In a second filtering step, amino acid sequences and a single set of dihedral angles corresponding to one of the

Identification of Evolutionary Motifs Encoding Jump Events

In order to identify aligned sites having potential evolutionary motifs encoding jump events, a specific criterion was developed.

For a particular protein pair, inference was performed under the model conditioned on the amino acid sequence and dihedral angles for both proteins \( (A_a, A_b, X_a, X_b) \). Homologous sites corresponding to a single hidden state and with evidence of a jump event (\( r'_a \neq r'_b \)) at posterior probability > 0.90 were identified, that is, the \( i \)'s such that \( p(H', r'_a \neq r'_b | A_a, A_b, X_a, X_b) > 0.90 \).
proteins were used \((A_a, A_b, X_a)\) or \((A_a, A_b, X_b)\) to infer the posterior probability, this time at a lower threshold: \(p(H, r'_a \neq r'_b | A_a, A_b, X_a) > 0.50\) or \(p(H, r'_a \neq r'_b | A_a, A_b, X_b) > 0.50\). This second criterion ensured that the evolutionary motif was identifiable under typical conditions where one has limited access to structural information (in this case a single protein structure in a pair). Only those aligned sites meeting both criteria were selected for downstream analysis.

**Statistical Alignment: Modeling Insertions and Deletions**

Protein sequences can not only undergo amino transitions due to underlying nucleotide mutations in the coding sequence, but also indel events. To account this, a modified pairwise TKF92 alignment HMM based on Miklós et al. (2004) was implemented. The TKF92 alignment HMM was augmented with the ETDBN evolutionary hidden states in order to capture local sequence and structure evolutionary dependencies. Furthermore, it was modified such that neighboring dependencies amongst hidden states at adjacent alignment sites were modeled. For more details see ‘Statistical alignment’ in Supplementary Material online.

Whilst it is possible to fix the alignment in advance by pre-aligning the sequences using one of the many available alignment methods (Katoh et al. 2002; Edgar 2004) or using a curated alignment (such as from the HOMSTRAD database), doing so ignores alignment uncertainty.

**Training and Test Datasets**

A training dataset of 1,200 protein pairs (2,400 proteins; 417,870 site observation pairs) and a test dataset of 38 protein pairs (76 proteins; 14,125 site observation pairs) were assembled from 1,032 protein families in the HOMSTRAD database. For further details see ‘Construction of test and training datasets’ in the supplementary Material, Supplementary Material online.

**Model Training and Selection**

Maximum Likelihood Estimation of the model parameters, \(\Psi\), was done using Stochastic Expectation Maximization (StEM, Gilks et al. 1995).

For further details of the E- and M-steps of the StEM algorithm and for details about model selection please refer to ‘Model training and selection’ in the supplementary Material, Supplementary Material online.

**Results and Discussion**

**Selecting the Number of Hidden States**

Fourteen models were trained \((8, 16, 32, 48, 52, 56, 60, 64, 68, 72, 76, 80, 96,\) and 112 hidden state models). The 64 hidden state model was chosen as the best model, as it had the lowest Bayesian Information Criterion (BIC, fig. 3).

**Stationary Distributions over Dihedral Angles Capture the Empirical Distribution**

Figure 4 illustrates the sampled and empirical dihedral angle distributions. There is a good correspondence between dihedral angles sampled under the model (fig. 4, left) and the empirical distribution of dihedral angles in our training dataset (fig. 4, right) for all three cases illustrated (all amino acids, glycine only, and proline only). The correspondence is not surprising given that ETDBN is effectively a mixture model with a large number of mixture components.

**Estimates of Evolutionary Time from Dihedral Angles Are Consistent with Estimates from Sequence**

Whilst ETDBN has a general scope with respect to applications (including acting as proposal distribution or as a building block in a homology modeling application), we envision the primary application being inference of evolutionary parameters.

Figure 5 compares evolutionary times estimated using only pairs of homologous amino acid sequences versus only pairs of homologous dihedral angles. As desired, the two estimates of evolutionary time for each protein pair are similar, as can be seen by the proximity of the points to the identity line.

A paired t-test gave a \(p\)-value of 0.578, thus failing to reject the null hypothesis that there is no difference between branch lengths estimated using sequence only versus angles only. This indicates that there is sufficient evolutionary information in the dihedral angles to estimate the evolutionary times and that the model is consistent in its estimates, lacking a significant tendency to underestimate or overestimate the evolutionary times when either sequence or dihedral angles are used.

Interestingly, the variance in the sampled evolutionary times is higher when dihedral angles only are used, as compared with sequence only (see fig. S1, Supplementary Material online).

**The Relationship between Evolutionary Time and Angular Distance Is Adequately Modeled**

We investigated the relationship between evolutionary time and angular distance between real protein pairs and protein pairs where the dihedral angles of \(p_b(X_b)\) were treated as missing and hence sampled (fig. 6).

As expected, for both real and sampled pairs, angular distance tends to increase as a function evolutionary time. For larger evolutionary times a plateau begins to emerge, which is expected as the maximum possible theoretical angular distance is \(\sqrt{8} \approx 2.828\).

When the evolutionary time is exactly zero \((t_{ab} = 0)\) under our model, the angular distance between sampled dihedral angles is exactly zero (not shown in fig. 6). However, this is not expected to be the case for real protein pairs when the two sequences are identical (due to the inherently flexible nature of proteins, different experimental conditions, experimental noise, etc.). It is therefore not surprising that the regression curve for the real protein pairs does not pass through zero.
Predictive accuracy was measured using 38 homologous protein pairs in the test dataset. For every protein pair \((p_a, p_b)\), the dihedral angles of \(p_a\) in each pair were treated as missing, and these missing dihedral angles were sampled under the model given a particular combination of observation types. The average angular distance between the sampled and known dihedral angles was used as the measure of predictive accuracy.

Figure 7 gives an example of predictive accuracy under different combinations of observations types overlaid on a cartoon structure of the protein structure being predicted, whereas figure 8 provides a representative view of predictive accuracy across 10 different protein pairs in the test dataset for different combinations of observations types. We highlight some of the key patterns identified in figures 7 and 8 as follows.

Combination 1 refers to random sampling from the model, implying no data observations were conditioned on besides the respective lengths of proteins \(p_a\) and \(p_b\). The average angular distance between the true and predicted dihedral angles was 1.6. Random sampling acts as a baseline for predictive accuracy. It is apparent from figure 7 that the model has a propensity to predict right-handed \(\alpha\)-helices, which is the most populated region in the Ramachandran plot.

Under combination 2, only the amino acid sequence corresponding to \(p_b\) is observed. As expected in figures 7 and 8 there is an increase in predictive accuracy with the addition of the amino acid sequence relative to combination 1.

Under combination 3, we add in the amino acid sequence of a homologous protein \((p_a)\). In all ten cases there is an improvement in predictive accuracy. The improvement in predictive accuracy is reasonable, as knowledge of the sequence evolutionary trajectory is expected to encode information about structure evolution and hence will inform the dihedral angle conformational possibilities.

Under combination 4, in addition to the two amino acid sequences we treat the homologous secondary structure as observed. This results in a substantial improvement in predictive accuracy as one would expect. Knowledge of the amino acid sequence and a homologous secondary structure strongly informs regions of the Ramachandran plot that are likely to be occupied.

Under combination 5 (which we consider the canonical combination—the standard homology modeling scenario), we treat both amino acid sequences as observed, as well as the dihedral angles of the homologous protein \((p_a)\)—in all cases the predictive accuracy improves over combination 4. This is anticipated as the homologous dihedral angles are expected to be the best proxy for missing dihedral angles and are therefore expected to be more informative than secondary structure alone. Note that the availability of a homologous amino acid sequence pair here and in combination 4 is consequential as it informs the evolutionary time \(t_{ew}\) parameter, which will typically constrain the distribution over dihedral angles and reduce the associated uncertainty.

Finally, in combination 6, the same data observations as in combination 5 are used, except the alignment is treated as given a priori (by the HOMSTRAD alignment).
rather than as unobserved. The HOMSTRAD alignment is based on a structural and sequence alignment of $p_a$ and $p_b$ and therefore is expected to encode a higher degree of homology and structural information than combination 5 (where the alignment is treated as unobserved and therefore a marginalization over alignments is performed). On average, there is a slight improvement in predictive accuracy when fixing the alignment, albeit the magnitude of improvement is not substantial. This demonstrates the accuracy of the alignment HMM.
Evolutionary Hidden States Reveal a Common Evolutionary Motif

One benefit of ETDBN is that the 64 evolutionary hidden states learned during the training phase are interpretable. We give an example of a hidden state encoding a jump event that was subsequently found to represent an evolutionary motif present in a large number of protein pairs in our test and training datasets.

Evolutionary hidden state 3 (fig. 9) was selected from the 64 hidden states as an example of a hidden state encoding a jump event and capturing angular shift (a large change in dihedral angle). A notable feature of this hidden state is that the change in dihedral angles between site-classes \( r_1 \) and \( r_2 \) is associated with specific amino acid changes. In site-class \( r_1 \) the amino acid frequencies are relatively spread out amongst a number of amino acids, whereas in site-class \( r_2 \) the frequencies are particularly concentrated in favor of glycine (Gly) and asparagine (Asp), with glycine being significantly more probable in site-class \( r_2 \) than \( r_1 \). This suggests that, conditioned on hidden state 3, an exchange between a glycine to another amino acid is likely indicative of a jump and hence a corresponding change in dihedral angle. This is consistent with what we find in a subsequent analysis of evolutionary motifs. This particular jump occurs in coil regions.

Having selected hidden state 3, positions in 238 protein pairs were analyzed for evidence of the corresponding evolutionary motif. 38 protein pairs in the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section. Using the first criterion, 84 protein sites were selected for the test dataset and a further 200 from the training dataset were analyzed using the criteria described in the Methods section.
Most positions in the homologous pair have low posterior jump probabilities (≈ 0.0), with the exception of positions N38/N38 and E39/G39, which both have high posterior jump probabilities (≈ 1.0). The exchange between a glutamate (at position 39 in 1poh) and a glycine (at position 39 in 1pch) appears to be responsible for the shift in dihedral angle. This exchange corresponds to a significant jump in dihedral angle: \( \langle \phi_{1poh,E39} \rangle, \langle \psi_{1poh,E39} \rangle = \langle -1.63, -0.06 \rangle \rightarrow \langle \phi_{1poh,G39}, \psi_{1poh,G39} \rangle = \langle 1.40, 0.22 \rangle \). The angular distance between the two dihedral angles is 2.01. This is consistent with the amino acid frequency parameters specified by the two site-classes for hidden state 3 (fig. 9).

Site-class \( r \) indicates that a number of amino acids (alanine, aspartic acid, glycine, histidine, lysine, asparagine, proline, glutamine, arginine, serine, and theorine) other than glutamate plausibly coincide with the particular dihedral angle conformation specified by site-class \( r \). The involvement of glycine in a jump is not surprising as it is a small and flexible amino acid, whereas the role of asparagine is less clear. In our analysis of 238 protein pairs we found that of the seven positions meeting the criteria for hidden state 3 and involving an exchange with asparagine (Asn), four were an exchange between an asparagine and a glycine, whereas the remaining three were between asparagine and one of lysine, histidine, or serine.

**Using Dihedral Angles for Alignment**

A valuable feature of our model is its ability to account for alignment uncertainty by summing over possible pairwise alignments using the TKF92 model as a prior distribution over indel histories, whilst simultaneously taking into account neighboring dependencies amongst aligned sites. Doing so results in a sample of alignments rather than a single alignment. Nevertheless, a single Maximum A Posteriori (MAP) pairwise alignment may be obtained from the alignment samples and used for downstream analysis.

ETDBN and several other alignment methods (namely StatAlign, BALI-Phy, MUSCLE, and MAFFT) were used to infer pairwise alignments from simulated and real data under various combinations of data observations, for example: an amino acid sequence pair \( (A, B) \), a secondary structure sequence pair \( (S, S) \), a dihedral angle sequence pair \( (X, X) \) and combinations thereof.

In the first set of benchmarks (fig. 11A), pairs of proteins were simulated from the ETDBN model conditioned on 38...
Fig. 8. Benchmarks of predictive accuracy (measured using angular distance, lower is better) on a random subset of ten protein pairs in the test dataset, giving a representative view of predictive accuracy under six different combinations of observations. The dihedral angles $X_{b}$ of $p_{b}$ were treated as missing and were sampled under the model, whereas $p_{a}$ was a homologous protein used for the purposes of prediction. See the legend of figure 7 for a description of each combination (A–F). The final set of bars, denoted 'Mean (N = 38)', are the mean values for the entire test dataset of $N = 38$ protein pairs. The error bars are the standard errors.

Fig. 9. Depiction of evolutionary hidden state 3. This hidden state was sampled at 0.69% of sites (the average was 1.56%). The equilibrium frequencies of $r_{1}$ and $r_{2}$ were $\pi_{1} = 0.691$ and $\pi_{2} = 0.309$, respectively. The jump rate was $\gamma = 31.76$. The corresponding site-class pair probabilities are depicted to the right as a function of evolutionary time. Note that the dashed red lines depicting the probabilities for (1, 2) and (2, 1) superimpose exactly, because the probabilities are equal—this holds for the jump probabilities of all hidden states as it is required for time-reversibility. In the main figure, the two rows depict the parameters encoded by the two site-classes, respectively. Columns 1 and 3 depict the parameters governing the amino acid and secondary structure stochastic processes, respectively. The secondary structure classes correspond to $H =$ helix, $S =$ sheet, and $C =$ coil. Column 2 depicts the WN diffusions. The stationary distributions of the WN diffusions are shown using black contour lines, the direction of the drifts are indicated by the arrows and the magnitude of the drifts at each position indicated using the color gradient.
different pairwise alignments and corresponding evolutionary
times. This resulted in a set of 38 simulated pairwise align-
ments together with corresponding observations, implying
that the true underlying alignments were known for each
of the simulated protein pairs. ETDBN and a number other
alignment methods were used to infer pairwise alignments
for each. The alignment similarity metric (Schwartz et al.
2005) was used to measure the similarity between the inferred
alignments and the true alignments, where higher similarity
indicates better predictions. It was found that, when using the
simulated amino acid sequences alone, ETDBN (11A.5) out-
performed all four other methods tested (11A.1 MUSCLE,
However, the greater performance of ETDBN compared with
other methods cannot be considered a fair comparison, as the
data were simulated under the ETDBN model.

More revealing in figure 11A was the alignment similar-
ity under ETDBN when using different combinations of
simulated data observations. It was found that secondary
structure alone (11A.6) performed the worst, which is un-
surprising given that only three states were available to
align the proteins. The second worst in terms of alignment
similarity was amino acid sequences alone (11A.5), fol-
lowed by amino acid sequences and secondary structures
(11A.7). Interestingly, using dihedral angles only (11A.8)
outperformed both 11A.5 (sequences only) and 11A.6 (sec-
ondary structures only). Finally, using amino acid sequence
 together with dihedral angles (11A.9) or all three data
types combined (11A.10) outperformed all other combina-
tions. This illustrates that, at least under simulation
conditions, increasing the number of data observations
results in better alignment accuracy.

Following that, the various alignment methods were
 benchmarked against 38 pairwise alignments consisting of
real sequence and structure observations in the test dataset.
These pairwise alignments were obtained from the
HOMSTRAD alignments. The sequence identity of these pair-
wise alignments ranged from 10% to 93%, with an average
sequence identity of 39%. In addition to methods 1–10 in
figure 11A, five structural alignment methods were also used:
11. StatAlign (Herman et al. 2014), 12. TMAlign (Zhang and
(Holm and Rosenström 2010), and 15. CE (Shindyalov and
Bourne 1998). These methods were not used in 11A due to
the lack of an appropriate model for simulating the evolution
of three-dimensional protein structures.

When benchmarking the MAP estimated alignments
against the HOMSTRAD alignments (fig. 11B), using real se-
quen ces alone for inference (A, A), ETDBN (11B.5) had a
similar degree of accuracy when compared with several other
sequence-based methods (11B.1 StatAlign, 11B.2 BaliPhy,
11B.3 MUSCLE, and 11B.4 MAFFT). This demonstrates that
ETDBN has performance comparable to that of other
commonly-used sequence alignment methods.

Using (, ) alone, ETDBN (11B.6) had substantially lower
alignment similarity compared with sequence only, which
was expected given that a similar result was obtained for
the simulated data (11A.6). However, when including the
real sequences (11B.7) the predictive accuracy was once again
comparable to sequence only inferences (11B.1–11B.5).

**Fig. 10.** Depiction of two histidine-containing phosphocarriers, PDB 1pch and 1poh, superimposed. On the left is a cartoon representation of the
two proteins corresponding to regions F29-K49 and F29-A42, respectively, with posterior jump probabilities at each position overlaid. On the right
is a ball-and-stick representation giving atomic detail for a smaller region (I36-G42 and T36-A42, respectively). The exchange between a glutamate
(E39 in 1poh) and a glycine (G39 in 1pch) is associated with a large change in dihedral angle as indicated by the curved arrows.
When using $(X_a, X_b)$ alone (11B.8), the alignment similarity was found to be somewhat worse than the sequence only cases. Furthermore, when introducing the sequences (11.9) and secondary structures (11B.10) in addition to the dihedral angles, the similarity remained worse than the sequence only methods (11B.1–11B.5), despite the additional information. These results are in contrast to the results we obtained for simulated data (11A.8–11A.10).

The non-statistical structural alignment methods (11B.12–11B.15) fared the best, likely because they use a criteria similar to that used to align the HOMSTRAD alignments. When interpreting these results it is important to note the HOMSTRAD alignments should not be considered the true underlying alignments and may even be strongly biased. For example, they may favor the closest structural superimposition of structures or the most parsimonious alignments, with the fewest number of indels. In the evolutionary modeling context our goal is to distinguish between homologous sites (sites that have evolved via mutation alone) and indels. In practice, it is extremely difficult to obtain the true underlying alignment (sets of homologies and indels), because it would require an experiment where every indel event since the common ancestor is observed, a seeming impossible task outside of simulation or laboratory conditions.

After further investigation, the trend of lower alignment similarity seen in figure 11B.6–11B.10 when using ETDBN with structural observations compared with sequence only or non-statistical structural alignment methods was found to reverse (11C.6–11C.10) upon calculating the precision of predicting homologous sites (the fraction of sites which were predicted as homologous and were correctly predicted as such). Therefore when only dihedral angle observations are used, ETDBN underpredicts the number of homologous sites, however, when a homologous site is predicted, it is correctly predicted more often than when using only amino acid sequences. In particular, ETDBN predicted fewer homologous sites with coiled secondary structure compared with homologous sites with helical or sheet secondary structure. This pattern of results may be in part due to the WN diffusion used to model evolution of dihedral angles. The WN diffusion is suitable for modeling angular drift (small changes in angles localized around a region of the Ramachandran plot) but does not sufficiently capture angular shift (large changes in angles between regions of the Ramachandran plot, which are more likely in coiled regions) due to stationarity. As noted before, the jump model is an abstraction intended to capture the end-points of evolution by allowing a jump between two regions of the Ramachandran plot, abstracting a potential intermediate evolutionary trajectory for the sake of computational tractability. Note that the jump model accurately captures the common cases where a single mutation induces a large conformational shift.

**Concluding Remarks**

The main achievement of this work is a computationally tractable, generative and interpretable probabilistic model of protein sequence and structure evolution on a local scale.
ETDBN, like TorusDBN (Boomsma et al. 2008), could be used for the inference of evolutionary parameters in its present form. However, we have shown that the current model can already be used for the inference of evolutionary parameters in its present form.

In addition to multi-level modeling, probabilistic models such as ETDBN allow one to account for and to make statements about uncertainty (e.g. with respect to evolutionary time, alignment, etc.) in a rigorous manner. In principle, ETDBN, like TorusDBN (Boomsma et al. 2008), could be used as a proposal distribution. In other words, ETDBN could be used to sample protein structures (possibly conditioned on various data observations) in a computationally efficient manner, such that the resulting samples are expected to be located in regions of high probability density with respect to the true underlying distribution.

A final key feature of our evolutionary model is its interpretability. This interpretability enables the identification of potential evolutionary motifs—common patterns of sequence–structure evolution. We identify one such evolutionary motif in 34 different homologous protein pairs. A major direction for future research is the further identification of such evolutionary motifs. Understanding these evolutionary motifs, may 1) improve homology modeling predictions; 2) provide more accurate estimates of evolutionary parameters; and 3) produce better models of protein evolution that more realistically capture evolutionary trajectories through sequence and structure space, which may help identify functionally relevant positions that are potential drug targets.

**Future Challenges**

**Pairwise to Phylogeny**

For reasons of computational tractability the implemented model is pairwise, but it is theoretically possible to generalize it to a phylogeny, such as in Herman et al. (2014). In practice, for three or more sequences on a phylogeny it is necessary to marginalize out the unobserved ancestral protein states in order to compute likelihoods. Felsenstein’s algorithm can be used to marginalize over discrete ancestral states, such as amino acids in a computationally efficient manner. However, we do not know whether a similar efficient algorithm exists for marginalizing the continuous ancestral dihedral angle states under the WN diffusion, thereby necessitating a more expensive MCMC algorithm. A possibly greater computational hindrance to considering a phylogeny is the alignment problem, which scales $O(l_1 \times l_2 \times \ldots \times l_N)$, where $l_i$ is the length of sequence $i$ and $N$ is the number of sequences, although MCMC approaches are possible (Herman et al. 2014).

**Context-Dependence**

Although we believe our model provides a substantial improvement over current stochastic models of sequence and structural evolution, there is still scope for improvement. The WN diffusions used to model dihedral angle evolution adequately capture angular drift (small local changes in dihedral angle), but are less capable of capturing angular shift (large changes in dihedral angle). This is to a considerable extent mitigated by the introduction of jump events, as discussed before. A more realistic model would model the entire evolutionary trajectory, allowing an arbitrary number of switches between site-classes together with neighboring dependencies amongst adjacent sites along the evolutionary trajectory. Similar context-dependent models are typically computationally expensive and require sophisticated inference procedures (Robinson et al. 2003; Yu and Thorne 2006).

**Software Availability**

Julia code (tested on both Windows and Linux platforms) is available at: http://www.computingforbiology.org/software/etdbn.
**Supplementary Material**

Supplementary material are available at *Molecular Biology and Evolution* online.

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