RAD51B in Familial Breast Cancer

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Abstract

Common variation on 14q24.1, close to \textit{RAD51B}, has been associated with breast cancer: rs999737 and rs2588809 with the risk of female breast cancer and rs1314913 with the risk of male breast cancer. The aim of this study was to investigate the role of \textit{RAD51B} variants in breast cancer predisposition, particularly in the context of familial breast cancer in Finland. We sequenced the coding region of \textit{RAD51B} in 168 Finnish breast cancer patients from the Helsinki region for identification of possible recurrent founder mutations. In addition, we studied the known rs999737, rs2588809, and rs1314913 SNPs and \textit{RAD51B} haplotypes in 44,791 breast cancer cases and 43,583 controls from 40 studies participating in the Breast Cancer Association Consortium (BCAC) that were genotyped on a custom chip (iCOGS). We identified one putatively pathogenic missense mutation c.541C>T among the Finnish cancer patients and subsequently genotyped the mutation in additional breast cancer cases (n = 5259) and population controls (n = 3586) from Finland and Belarus. No significant association with breast cancer risk was seen in the meta-analysis of the Finnish datasets or in the large BCAC dataset. The association with previously identified risk variants rs999737, rs2588809, and rs1314913 was replicated among all breast cancer cases and also among familial cases in the BCAC dataset. The most significant association was observed for the haplotype carrying the risk-alleles of all the three SNPs both among all breast cancer cases (odds ratio (OR): 1.15, 95% confidence interval (CI): 1.11–1.19, P = 8.88 x 10^-11) and among familial cases (OR: 1.24, 95% CI: 1.16–1.32, P = 6.19 x 10^-11), compared to the haplotype with the respective protective alleles. Our results suggest that loss-of-function mutations in \textit{RAD51B} are rare, but common variation at the \textit{RAD51B} region is significantly associated with familial breast cancer risk.

Introduction

Breast cancer is the most frequent cancer among women worldwide and also the leading cause of cancer-related death [1]. Several susceptibility loci for breast cancer have been identified and most of the currently known high- and moderate-penetration predisposition genes have a role in DNA repair. The major high-penetration breast and ovarian cancer susceptibility genes \textit{BRCA1} and \textit{BRCA2} are important for DNA double-strand break (DSB) repair through homologous recombination (HR) [2]. Proteins encoded by the moderate-penetration genes \textit{ATM}, \textit{BRIP1}, and \textit{CHEK2} interact with \textit{BRCA1} in DNA damage repair whereas \textit{PALB2} associates with both \textit{BRCA1} and \textit{BRCA2} [2,3]. In the HR repair of DSBs, the RAD51 recombinase has a key role. Binding of RAD51 to single-stranded DNA at the break site initiates the repair of a DSB [4]. In humans, there are five RAD51 paralogs RAD51B, RAD51C, RAD51D, XRCC2, and XRCC3, and they promote the binding of RAD51 to the DNA. Rare pathogenic mutations in \textit{RAD51} paralogs \textit{RAD51C} and \textit{RAD51D} have been identified in breast and ovarian cancer families and confer a high risk specifically for ovarian cancer [5–7] whereas a homozygous missense mutation in \textit{RAD51C} (\textit{FANCO}) was found in a Fanconi anemia patient [8]. A homozygous \textit{XRCC2} mutation has also been detected in a Fanconi anemia patient [9] and rare mutations in the gene in breast cancer families were identified in an exome sequencing study [10]; however, the association of \textit{XRCC2} with breast cancer risk could not be confirmed in a large follow-up study [11].
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Variants in the RAD51B region, also known as RAD51LI, have been associated with breast cancer risk in genome-wide association studies (GWAS) [12–14]. The major allele of the common polymorphism rs999737 in intron 10 of RAD51B and the minor-allele of rs2588809 in intron 7 have been associated with an increased risk of female breast cancer [12,14]. The rs999737 has also been associated with breast cancer risk among BRCA1 mutation carriers whereas no association was found for rs999737 or rs2588809 with breast cancer subtypes among BRCA1 or BRCA2 mutation carriers [15,16]. Another common polymorphism located in the intron 7 of the gene, rs1314913, has been associated with the risk of male breast cancer [13]. RAD51B is located at 14q24.1 and is expressed widely, with the highest levels in tissues that are active in recombination [17]. The RAD51B and RAD51C proteins form a stable complex which interacts weakly with RAD51 and promotes the assembly of RAD51 nuclear foci [18,19]. Furthermore, RAD51B is part of a larger BCDX2 complex that is formed with RAD51C, RAD51D, and XRCC2 [4]. The BCDX2 complex acts upstream of RAD51 recruitment to DNA damage foci [20]. Haploinsufficiency of RAD51B leads to aberrant HR repair of DNA and causes centrosome fragmentation and aneuploidy in human cells which suggests that loss of the proper biallelic expression of RAD51B may lead to chromosome instability in tumor cells [21].

In the Finnish population, recurrent founder mutations have been observed in many of the known breast and ovarian cancer susceptibility genes, including the RAD51 paralogs RAD51C and RAD51D [22,23]. Thus, the Finnish population is a valuable resource for the identification of new susceptibility alleles.

To identify putative recurrent founder mutations and to study the role of RAD51B in female and male breast cancer predisposition especially in familial breast cancer, we comprehensively screened the RAD51B gene in 172 Finnish patients. One identified missense mutation was in silico predicted to be pathogenic and was subsequently screened in a larger set of breast cancer cases and population controls from four datasets. In addition, the known RAD51B risk SNPs rs999737, rs2588809, and rs1314913 and RAD51B haplotypes were studied in the previously published 40 studies participating in the Breast Cancer Association Consortium (BCAC) including 44,791 breast cancer cases and 43,583 controls.

Materials and Methods
Screening of the RAD51B gene
The coding region and the exon-intron boundaries of the RAD51B gene (RefSeq NG_023267.1, NM_133509.3) were screened in 172 cancer patients from Southern Finland: in 87 female breast cancer patients with a family history of female breast cancer or breast and ovarian cancer, in 4 familial ovarian cancer patients, and in 4 female breast cancer patients with a family history of male and female breast cancer from the Helsinki region of Finland as well as in 77 male breast cancer patients (33 from the Helsinki region and 44 from the Tampere region) (S1 Text). All the female breast or ovarian cancer families and 60 of the male breast cancer families and patients were previously screened negative for BRCA1/2 mutations whereas 21 patients had not been tested for BRCA1/2 mutations. Genomic DNA isolated from blood was amplified by PCR and subsequently sequenced using ABI BigDyeTerminator 3.1 Cycle Sequencing kit (Life Technologies) (S1 Table). The sequencing results were analyzed with Variant Reporter Software v1.0 (Life Technologies).

Genotyping of c.541C>T
The identified c.541C>T, p.(Arg181Trp) missense change was genotyped in additional familial and unselected breast cancer patients and population controls from the Helsinki (cases = 2203,
Bioinformatics and statistical methods

The pathogenicity of the variants identified in the sequencing of \textit{RAD51B} was predicted using the MutationTaster software as it considers both exonic and intronic variants [25]. In addition, PON-P that utilizes results from SIFT, PhD-SNP, PolyPhen-2, SNAP, and I-Mutant 3.0 was used to predict the pathogenicity of the missense variants [26]. Secondary structure prediction was done with RaptorX [27] and protein-protein interaction with PredictProtein [28]. Statistical analyses were performed in R version 3.0.2 (http://www.r-project.org/). To study the association of the c.541C>T mutation with breast cancer risk, two-sided \textit{P}-values were calculated using Pearson’s chi-squared test or, if the expected number of cell count was less than five, Fisher’s exact test. For meta-analysis, the estimates were combined using a fixed-effects meta-analysis, using the inverse variance-weighted method. Student’s \textit{t}-test was used to compare the age at diagnosis between the mutation carriers and non-carriers. Per-allele odds ratios (OR) and confidence intervals (CI) for the common \textit{RAD51B} polymorphisms were estimated with logistic regression. Multivariate regression models including any two of the three SNPs at a time were used to study the independence of the association signals. Haplotype-specific ORs and CIs were estimated with SHAPEIT and IMPUTE2 by using the 1000Genomes project as the reference panel [24]. The analyses were restricted to cases with European ancestry. All participants gave written informed consent and all the studies were approved by the respective Institutional Review Boards or Ethics Committees (S2 Table).
regional association plot for visualization of the SNP associations and the extent of linkage disequilibrium (LD) [29].

Results

We sequenced the coding region and the exon-intron boundaries of the RAD51B gene in 168 breast (female and male) and 4 familial ovarian cancer patients from Southern Finland. Nine intronic and six missense variants were identified (Table 1). The c.541C>T, p.(Arg181Trp), missense change was the only variant that was predicted to be pathogenic by both MutationTaster and PON-P software and was selected for further genotyping. Based on Finnish subjects in the ExAC dataset, the minor-allele frequency (MAF) for the c.541C>T variant is estimated to be 1.2% whereas in the non-Finnish Europeans the MAF is 0.05% (Exome Aggregation Consortium (ExAC), Cambridge, MA (URL: http://exac.broadinstitute.org) [May, 2015]). According to RaptorX secondary structure prediction software the arginine in position 181 is located in beta-sheet with the likelihood of 83.4% but, as determined by PredictProtein, the amino acid is not predicted to directly participate in protein-protein interactions.

The c.541C>T missense variant was genotyped in additional 3359 female and male breast cancer patients and families, and in 2351 population controls from Southern (Helsinki and Tampere datasets) and Northern Finland (Oulu dataset). Altogether, when combining the two stages of mutation testing, c.541C>T was screened in 2331 patients from Helsinki, 748 patients from Tampere, and 452 patients from Oulu. There was no evidence of association in any of the population-based case-control studies (Table 2). There was a suggestive association among breast cancer families with at least three first or second-degree relatives affected with breast or ovarian cancer, compared to population controls, in the Helsinki dataset (OR: 2.31, 95% CI: 1.20–4.48, \(P = 0.010\)) (Table 2). The mean age at breast cancer diagnosis was similar between carriers and non-carriers.

Table 1. Variants identified in the screening of the RAD51B gene (RefSeq NM_133509.3).

<table>
<thead>
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<th>DNA change</th>
<th>protein change</th>
<th>rs-number</th>
<th>position</th>
<th>AA (^a)</th>
<th>Aa (^b)</th>
<th>Aa (^c)</th>
<th>MAF (^d)</th>
<th>MAF(_{ExAC}) (^e)</th>
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<td>c.84+28T&gt;C</td>
<td></td>
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<td>71</td>
<td>84</td>
<td>17</td>
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<td>0.287</td>
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<td>NA</td>
</tr>
</tbody>
</table>

Counts of \(^a\) common homozygotes
\(^b\) heterozygotes and
\(^c\) rare homozygotes in this study
\(^d\) Minor-allele frequency (MAF) in this study
\(^e\) MAF in the Finnish population in the ExAC database

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Table 2. Frequencies and ORs for the c.541C>T mutation among the different patient subgroups in the Helsinki, Tampere, Oulu, and Belarus series.

<table>
<thead>
<tr>
<th>HELSINKI dataset</th>
<th>Total</th>
<th>CC</th>
<th>%</th>
<th>CT</th>
<th>%</th>
<th>OR (95% CI)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls</td>
<td>1278</td>
<td>1257</td>
<td>98.36%</td>
<td>21</td>
<td>1.64%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>All female BC cases</td>
<td>2293</td>
<td>2240</td>
<td>97.69%</td>
<td>53</td>
<td>2.31%</td>
<td>1.42 (0.85–2.36)</td>
<td>0.179</td>
</tr>
<tr>
<td>Familial BC (≥3 affected)</td>
<td>430</td>
<td>414</td>
<td>96.28%</td>
<td>16</td>
<td>3.72%</td>
<td>2.31 (1.20–4.48)</td>
<td>0.010</td>
</tr>
<tr>
<td>Familial BC (2 affected)</td>
<td>523</td>
<td>513</td>
<td>98.09%</td>
<td>10</td>
<td>1.91%</td>
<td>1.17 (0.55–2.50)</td>
<td>0.690</td>
</tr>
<tr>
<td>Unselected BC</td>
<td>1728</td>
<td>1690</td>
<td>97.80%</td>
<td>38</td>
<td>2.20%</td>
<td>1.35 (0.79–2.30)</td>
<td>0.277</td>
</tr>
<tr>
<td>Male BC c</td>
<td>37</td>
<td>36</td>
<td>97.30%</td>
<td>1</td>
<td>2.70%</td>
<td>1.66 (0.22–12.70)</td>
<td>0.469</td>
</tr>
<tr>
<td>Familial OC</td>
<td>4</td>
<td>4</td>
<td>100%</td>
<td>0</td>
<td>0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>TAMPERE dataset</strong></td>
<td><strong>Total</strong></td>
<td><strong>CC</strong></td>
<td>%</td>
<td><strong>CT/TT</strong></td>
<td>%</td>
<td><strong>OR (95% CI)</strong></td>
<td><strong>P</strong></td>
</tr>
<tr>
<td>Controls</td>
<td>800</td>
<td>778</td>
<td>97.25%</td>
<td>22</td>
<td>2.75%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Unselected BC</td>
<td>645</td>
<td>620</td>
<td>96.12%</td>
<td>25</td>
<td>3.88%</td>
<td>1.43 (0.80–2.55)</td>
<td>0.230</td>
</tr>
<tr>
<td>Male BC e</td>
<td>103</td>
<td>102</td>
<td>99.03%</td>
<td>1</td>
<td>0.97%</td>
<td>0.35 (0.05–2.60)</td>
<td>0.503</td>
</tr>
<tr>
<td><strong>OULU dataset</strong></td>
<td><strong>Total</strong></td>
<td><strong>CC</strong></td>
<td>%</td>
<td><strong>CT</strong></td>
<td>%</td>
<td><strong>OR (95% CI)</strong></td>
<td><strong>P</strong></td>
</tr>
<tr>
<td>Controls</td>
<td>273</td>
<td>267</td>
<td>97.80%</td>
<td>6</td>
<td>2.20%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>All female BC cases</td>
<td>452</td>
<td>443</td>
<td>98.01%</td>
<td>9</td>
<td>1.99%</td>
<td>0.90 (0.32–2.57)</td>
<td>0.850</td>
</tr>
<tr>
<td>Familial BC (≥3 affected)</td>
<td>83</td>
<td>82</td>
<td>98.80%</td>
<td>1</td>
<td>1.20%</td>
<td>0.54 (0.06–4.57)</td>
<td>1</td>
</tr>
<tr>
<td>Familial BC (2 affected)</td>
<td>49</td>
<td>48</td>
<td>97.96%</td>
<td>1</td>
<td>2.04%</td>
<td>0.93 (0.11–7.87)</td>
<td>1</td>
</tr>
<tr>
<td>Unselected BC</td>
<td>320</td>
<td>313</td>
<td>97.81%</td>
<td>7</td>
<td>2.19%</td>
<td>1.00 (0.33–3.00)</td>
<td>0.993</td>
</tr>
<tr>
<td><strong>BELARUS dataset</strong></td>
<td><strong>Total</strong></td>
<td><strong>CC</strong></td>
<td>%</td>
<td><strong>CT</strong></td>
<td>%</td>
<td><strong>OR (95% CI)</strong></td>
<td><strong>P</strong></td>
</tr>
<tr>
<td>Controls</td>
<td>1235</td>
<td>1235</td>
<td>100%</td>
<td>0</td>
<td>0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BC cases</td>
<td>1900</td>
<td>1897</td>
<td>99.85%</td>
<td>3</td>
<td>0.15%</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

BC = breast cancer; OC = ovarian cancer

a Families with ≥3 BC or OC among first- or second-degree relatives
b Two first-degree relatives affected with BC or OC
c Includes 33 male BC cases and 4 female BC cases with family history of male BC
d The Tampere dataset included one case with homozygous mutation
e Male BC cases only

non-carriers among the familial patients (54.2 for carriers and 53.1 for non-carriers, $P = 0.725$) and among all cases (55.9 for carriers and 56.4 for non-carriers, $P = 0.744$). However, no association was observed for familial breast cancer in the Oulu dataset, and there was no evidence of association when all datasets were combined (OR: 1.35, 95% CI: 0.66–2.73, $P = 0.410$). The variant was also genotyped in 1900 breast cancer cases and 1235 population controls from Belarus but only three carriers were identified among cases and none among controls (Table 2).

We further studied the missense variant c.541C>T and the previously reported common risk SNPs rs2588809, rs1314913, and rs999737 in a large breast cancer dataset from BCAC comprising 40 studies with 44,791 invasive breast cancer cases and 43,583 controls of predominantly European ancestry. The subjects were genotyped on an Illumina Infinium custom chip (iCOGS) for over 200,000 SNPs [14], including the rs2588809, rs1314913, and rs999737 SNPs. Genotypes for over 11 million SNPs, including the c.541C>T missense variant, were imputed by using the 1000Genomes project as a reference panel [24]. For the missense variant, we first performed an association test stratified by study including all the 40 BCAC studies. The overall information score for the c.541C>T was 0.673. To increase the imputation accuracy of this rare variant, studies with information score less than 0.5 in the stratified analysis were excluded from the final analysis (S3 Table). Altogether 26,969 cases and 27,092 controls from 23 studies were included and the information score for the variant in the final analysis was 0.755. The
MAF of the c.541C>T missense variant was 0.2% among all cases, familial cases, and controls and it did not associate with breast cancer among all cases (OR: 0.97, 95% CI: 0.76–1.25, \( P = 0.538 \)) or familial cases (OR: 1.04, 95% CI: 0.65–1.66, \( P = 0.794 \)). The highest frequency was observed in the Finnish studies HEBCS, KBCP, and OBCS with the overall MAF ranging from 0.6% to 0.7%. The HEBCS and OBCS studies are overlapping with the Finnish Helsinki and Oulu datasets that were genotyped for the variant.

Breast cancer associations for two of the genotyped SNPs, rs2588809 and rs999737, in the BCAC dataset have been published before [14,24,30] and the results were replicated here. The minor-alleles of rs2588809 and rs1314913 and the major-allele of rs999737 were associated with an increased risk of breast cancer with ORs between 1.07–1.08 among all breast cancer cases and also among familial cases with ORs between 1.10–1.15 (Table 3). The associations were also significant among the subset of cases with ER-positive tumors but only rs999737 showed association among the ER-negative subset (Table 3). Rs2588809 and rs1314913 are strongly correlated (\( r^2 = 0.816 \)), whereas rs999737 is not correlated with them (\( r^2 = 0.003 \) with rs2588809 and \( r^2 = 0.070 \) with rs1314913) (S1 Fig). In multivariate logistic regression models, both rs2588809 and rs1314913 remained significant after adjustment with rs999737 (OR: 1.07, 95% CI: 1.04–1.09, \( P = 1.74 \times 10^{-6} \)) and OR: 1.06, 95% CI: 1.03–1.09, \( P = 9.72 \times 10^{-5} \), respectively). Likewise, rs999737 remained an independent predictor for risk after adjustment with rs2588809 or rs1314913 (OR: 1.08, 95% CI: 1.06–1.11, \( P = 2.91 \times 10^{-11} \) and \( P = 2.54 \times 10^{-11} \), respectively). Neither rs2588809 nor rs1314913 remained significant when adjusted with each other (OR: 1.07, 95% CI: 1.00–1.16, \( P = 0.066 \) and OR: 1.00, 95% CI: 0.93–1.08, \( P = 0.977 \), respectively).

To evaluate the allele combination associated with the highest risk, we performed haplotype analysis for rs2588809, rs1314913, and rs999737. The reference haplotype CCT, carrying the major-alleles of rs2588809 and rs1314913 and the minor-allele of rs999737 had an estimated frequency of 21.0% among controls and 19.4% among cases. The strongest association was observed for the TTC haplotype that carries the risk-alleles of all the three SNPs with an odds ratio of 1.15 for all cases (95% CI: 1.11–1.19, \( P = 8.88 \times 10^{-16} \)) and 1.24 for familial cases (95% CI: 1.16–1.32, \( P = 6.19 \times 10^{-11} \)) (Table 4). This haplotype had an estimated frequency of 12.6% among controls, 13.5% among all cases, and 14.1% among familial cases. The most common haplotype, with over 60% frequency among cases and controls, was the haplotype CCC carrying the risk-allele of rs999737 and also this haplotype associated with an increased risk of breast cancer among all the cases (OR: 1.09, 95% CI: 1.06–1.12, \( P = 1.86 \times 10^{-10} \)) and the familial cases (OR: 1.14, 95% CI: 1.09–1.20, \( P = 1.30 \times 10^{-7} \)) when compared to the CCT haplotype.

**Discussion**

To study the role of RAD51B in breast cancer predisposition, we screened the coding sequence in 172 Finnish breast or ovarian cancer patients. We also studied the common susceptibility variants in the genomic region of the gene in a large dataset from BCAC including 44,791

<table>
<thead>
<tr>
<th>Rs-number</th>
<th>MAF controls</th>
<th>MAF cases</th>
<th>ORALL (95%CI)</th>
<th>( P ) ALL</th>
<th>ORFAM (95%CI)</th>
<th>( P ) FAM</th>
<th>ORER+ (95%CI)</th>
<th>( P ) ER+</th>
<th>ORER- (95%CI)</th>
<th>( P ) ER-</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs2588809</td>
<td>0.158</td>
<td>0.169</td>
<td>1.08 (1.05–1.10)</td>
<td>( 4.62 \times 10^{-5} )</td>
<td>1.10 (1.05–1.16)</td>
<td>( 9.72 \times 10^{-5} )</td>
<td>1.09 (1.06–1.12)</td>
<td>( 1.85 \times 10^{-4} )</td>
<td>1.00 (0.95–1.05)</td>
<td>( 8.90 \times 10^{-1} )</td>
</tr>
<tr>
<td>rs1314913</td>
<td>0.144</td>
<td>0.154</td>
<td>1.07 (1.04–1.10)</td>
<td>( 2.74 \times 10^{-4} )</td>
<td>1.10 (1.04–1.16)</td>
<td>( 2.45 \times 10^{-4} )</td>
<td>1.08 (1.05–1.12)</td>
<td>( 3.60 \times 10^{-4} )</td>
<td>0.98 (0.93–1.03)</td>
<td>( 3.85 \times 10^{-1} )</td>
</tr>
<tr>
<td>rs999737</td>
<td>0.231</td>
<td>0.215</td>
<td>1.09 (1.06–1.11)</td>
<td>( 7.78 \times 10^{-13} )</td>
<td>1.15 (1.10–1.20)</td>
<td>( 1.08 \times 10^{-9} )</td>
<td>1.10 (1.07–1.13)</td>
<td>( 7.70 \times 10^{-12} )</td>
<td>1.05 (1.00–1.10)</td>
<td>( 3.03 \times 10^{-5} )</td>
</tr>
</tbody>
</table>

ALL = all breast cancer cases; FAM = familial breast cancer cases; ER+ = ER-positive; ER- = ER- negative.

doi:10.1371/journal.pone.0153788.t003
Among the Finnish patients, we identified one putatively pathogenic missense variant, c.541C>T, p.(Arg181Trp), that was further screened in a larger set of unselected and familial breast cancer patients and population controls.

In the Helsinki dataset, the c.541C>T mutation had a suggestive association with familial breast cancer but this association was not confirmed in the other Finnish sample sets. In Belarus, the mutation was rare and was identified only in three cases but not among controls. For the large BCAC dataset that had been genotyped on the iCOGS chip, c.541C>T genotypes were imputed. The missense was very rare with a 0.2% MAF and no association with breast cancer was seen among all breast cancer cases or in the subset of familial patients.

Out of the common variants at the RAD51B region, rs999737 and rs2588809 have been associated with female breast cancer and rs1314913 with male breast cancer [12–14,30]. For rs999737 and rs2588809 the results from the previous BCAC studies for all cases and by ER status were replicated here [14,30]. The rs1314913 SNP has been previously associated with male breast cancer with an OR of 1.57 (P = 3.02 × 10^{-13}) [13]. It has not been previously reported in female breast cancer, and showed here similar associations as rs2588809 with OR of 1.07 among all cases and 1.09 among ER-positive cases. The rs1314913 SNP is correlated with rs2588809 (r^2 = 0.816) but not with rs999737 (r^2 = 0.070). Our results suggest two independent associations at the region as rs999737 adjusted with rs1314913 or rs2588809 showed significant association whereas neither rs1314913 nor rs2588809 remained significant after adjustment with each other, suggesting both may be tagging another, causative allele.

All three variants were also associated with familial breast cancer. The strongest association was observed for a haplotype carrying the risk-alleles of all the three SNPs both among all cases and familial cases with ORs of 1.15 and 1.24, respectively. This haplotype is observed at a frequency of 12.6% among controls and 13.5% among cases.

As of today, only three studies have reported truncating germline mutations in RAD51B: two deleterious mutations were detected among ovarian cancer cases, one splicing mutation in a breast and ovarian cancer family, and one nonsense mutation in a melanoma family [31–33]. In an Australian study, no pathogenic mutations were identified among 188 breast cancer families [34]. These reports together with our results suggest that loss-of-function mutations in RAD51B are very rare yet we cannot exclude such mutations and possibly a higher breast cancer risk associated with these. However, the common variants in the RAD51B region are associated with an increased breast cancer risk. Further fine-mapping studies are needed to identify the causative variants underlying the associations, and functional studies to determine whether RAD51B or another gene in the region is the target of these associations. RAD51B is a plausible candidate gene for the association as it functions in HR repair of DNA damage like most of the known breast cancer genes.

Table 4. Estimated haplotype frequencies and haplotype-specific ORs among the BCAC dataset. The haplotypes are based on genotypes for SNPs rs2588809, rs1314913, and rs999737.

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>All breast cancer cases</th>
<th></th>
<th></th>
<th></th>
<th>Familial breast cancer cases</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Freq controls</td>
<td>Freq cases</td>
<td>OR (95% CI)</td>
<td>P</td>
<td>Freq controls</td>
<td>Freq cases</td>
<td>OR (95% CI)</td>
</tr>
<tr>
<td>CCT (reference)</td>
<td>0.210</td>
<td>0.194</td>
<td>1.15 (1.11–1.19)</td>
<td>8.88 × 10^{-16}</td>
<td>0.211</td>
<td>0.188</td>
<td>1.24 (1.16–1.32)</td>
</tr>
<tr>
<td>TTC</td>
<td>0.126</td>
<td>0.135</td>
<td>1.09 (1.06–1.12)</td>
<td>1.86 × 10^{-10}</td>
<td>0.126</td>
<td>0.141</td>
<td>1.14 (1.09–1.20)</td>
</tr>
<tr>
<td>CCC</td>
<td>0.632</td>
<td>0.636</td>
<td>1.15 (1.04–1.27)</td>
<td>5.48 × 10^{-3}</td>
<td>0.628</td>
<td>0.635</td>
<td>1.24 (1.16–1.32)</td>
</tr>
<tr>
<td>TCC</td>
<td>0.011</td>
<td>0.013</td>
<td>1.11 (0.99–1.23)</td>
<td>7.12 × 10^{-2}</td>
<td>0.012</td>
<td>0.013</td>
<td>1.21 (1.09–1.34)</td>
</tr>
<tr>
<td>TTT</td>
<td>0.018</td>
<td>0.018</td>
<td></td>
<td></td>
<td>0.018</td>
<td>0.017</td>
<td>1.06 (0.86–1.31)</td>
</tr>
</tbody>
</table>

Freq = frequency

doi:10.1371/journal.pone.0153788.t004
In conclusion, no pathogenic \textit{RAD51B} mutations were identified among 172 Finnish breast or ovarian cancer patients. However, we cannot rule out rare risk-variants in the Finnish or other populations. The minor-alleles of the common polymorphisms rs2588809 and rs1314913 and the major-allele of rs999737 were associated with breast cancer risk in the large BCAC dataset. The strongest association was observed for a risk haplotype carrying the risk-alleles of all the three SNPs with an OR of 1.15 among all cases and 1.24 among familial cases compared to the haplotype with the respective protective alleles.

**Supporting Information**

S1 Fig. Regional association plot for the \textit{RAD51B} SNPs rs2588809, rs1314913, and rs999737 and the c.541C>T missense variant. Each variant is represented with a dot and the color of the dot represents the extent of LD ($r^2$) with rs2588809. Association among all breast cancer cases in the BCAC dataset is represented at the -log10 scale. The x axis shows the genomic positions of the variants based on hg19 build. The right y axis shows the estimated recombination rate at the region (centiMorgans/megabase, cM/Mb). LD and recombination rate were estimated using the 1000Genomes Nov 2014 EUR as the reference population. (PDF)

S1 Table. Primers and PCR conditions for the screening of the \textit{RAD51B} gene and the c.541C>T missense mutation. (DOCX)

S2 Table. Description of the BCAC studies. BCAC studies genotyped on the iCOGS array, ethics approval committees/institutional review boards and the numbers of controls, all invasive cases, cases with first-degree family history of breast cancer, and cases with ER-positive and negative tumors included in this study. All subjects are genetically European. (DOCX)

S3 Table. The imputed c.541C>T variant in the BCAC dataset. Minor-allele frequencies (MAF) of the imputed c.541C>T missense variant in the BCAC studies and the information scores from the association analysis stratified by study. Studies with information score <0.5 were excluded from the final analysis. (DOCX)

S1 Text. Supplementary Materials and Methods. (DOCX)

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