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**Lab Resource: Stem Cell Line**

**Generation of a gene-corrected isogenic control cell line from an Alzheimer's disease patient iPSC line carrying a A79V mutation in PSEN1**

Carlota Pires, Benjamin Schmid, Carina Petraæus, Anna Poon, Natakarn Nimsanorc, Troels T. Nielsen, Gunhild Waldemar, Lena E. Hjermind, Jørgen E. Nielsen, Poul Hyttel, Kristine K. Freude

A79V-hiPSC (c.236 T→C) was gene-corrected with the CRISPR/Cas9 system (Ran et al., 2013) by replacing the point mutation “T” with the wild-type nucleotide “C” (Fig. 1A). The nucleotide substitution/correction was validated by sequencing (Fig. 1B). Gene-integrity was also checked by confirming that the DNA sequence surrounding the mutation and the CRISPR cutting site were not altered, with no frameshifts or other mutation(s) being generated upon gene editing.

The study was approved by the “De Videnskabsetiske Komiteer for Region Hovedstaden” (protocol number H-4-2011-157), Copenhagen, Denmark and written informed consent was obtained in all cases. To protect the patient family privacy, no personal patient information is presented here. Mutations in presenilin 1 (PSEN1) gene are the most common known causes of inherited Alzheimer’s disease (AD).

A human induced pluripotent stem cell (hiPSC) line was previously generated from a skin biopsy obtained from a 48-year-old woman carrying a heterozygous mutation in exon 4 of the PSEN1 gene, which causes a change in amino acid A79V. Episomal plasmids carrying gene sequences for hOCT4, hSOX2, hKLF4, hL-MYC, hLIN28, and shRNA against TP53 (Okita et al., 2011) were used to reprogram the fibroblasts into iPSCs, successfully establishing integration and feeder-free iPSCs. This cell line has been described previously as a bonafide iPSC line with normal karyotype (Li et al., 2016).

The A79V-hiPSC (c.236 C>T) was gene-corrected with the CRISPR/Cas9 system (Ran et al., 2013) by replacing the point mutation “T” with the wild-type nucleotide “C” (Fig. 1A). The nucleotide substitution/correction was validated by sequencing (Fig. 1B). Gene-integrity was also checked by confirming that the DNA sequence surrounding the mutation and the CRISPR cutting site were not altered, with no frameshifts or other mutation(s) being generated upon gene editing, confirming successful gene correction of A79V-hiPSC, named A79V-GC-hiPSC (“GC” = gene corrected). The sequencing of the isogenic line confirmed the c.236 T>C replacement in exon 4 of the PSEN1 gene.
gene corresponding to the wild-type (healthy) sequence (Fig. 1B). Immunocytochemical (ICC) analysis of the pluripotency markers OCT4, NANOG, SSEA3, SSEA4, TRA-1-60 and TRA-1-81 (Fig. 1C) confirmed the gene-edited line maintained its pluripotency like the mother line previously published. Finally, karyotyping was performed to confirm that no chromosomal aberrations were introduced in the isogenic line upon gene correction (Fig. 1D).

Materials and methods

CRISPR design

Isogenic gene-corrected controls were obtained using the CRISPR/Cas9 technology in combination with single-stranded oligonucleotides (ssODNs), which provided the wild-type template for gene-correction. CRISPRs were designed with the help of the software available at http://crispr.mit.edu/. The single-guide RNAs (sgRNA) were ordered from Sigma and the cloning of CRISPRs followed the protocol from Ran et al. (2013), with a single plasmid containing both the sgRNA and the Cas9 (pSpCas9(BB)-2A-Puro (PX459) V2.0; addgene ID 48139).

ssODNs were designed to contain 120 bp in total, with 60 bp upstream and 60 bp downstream of the cutting site. Screening of clones was performed with restriction digestions where the wild-type sequence was recognized (and cut) by specific restriction enzymes (Ehel, 5′-GAGTTTG-3′, ThermoScientific), while the mutated sequence was not, which means Ehel only digested clones where the ssODN was successfully integrated.

Nucleofection

A79V-hiPSCs growing on matrigel-coated (Corning Bioscience) dishes with E8 medium (Gibco) were detached with Accutase (Gibco). 2 × 10^6 cells were nucleofected with 10 μg of CRISPR/Cas9 plasmid and 100 pmol of ssODN using the Amaxa 4D Nucleofector (program CA137, Lonza) and P3 Primary Cell Kit (Lonza) according to the manufacturer’s instructions. Nucleofected hiPSCs were plated back in a matrigel-coated dish with E8 medium supplemented with 1 mM ROCK inhibitor (Sigma). hiPSCs were subjected to puromycin selection 4 h after nucleofection and allowed to recover for 4–5 days. Surviving (resistant) colonies were manually picked into 96 well plates coated with matrigel with E8 and then expanded for genotyping and sequencing.

Genotyping and restriction assays

DNA for genotyping was extracted using the prepGem™ Kit (ZyGEM, PTI0500) (Qiagen). PCR genotyping was performed using TEMPase Hot Start DNA Polymerase (Ampliqon) according to the manufacturer’s instructions, using an annealing temperature of 60 °C. The following primers were designed to cover the mutation and

![Fig. 1. A. CRISPR/Cas9 and ssODN used to repair the point mutation in A79V-hiPSC. A) Genomic sequence surrounding the mutation site: mutated nucleotide (T, red); sgRNA recognition site containing 20 bp (yellow); CRISPR cutting site between the 17th and 18th bp (bold); forward and reverse primers (pink). B) ssODN with 120 bp, 60 bp upstream and 60 bp downstream the mutation site containing the WT nucleotide (C, green). B) Sequencing of exon 4 of the PSEN1 gene in hiPSCs. A) Heterozygous c.236C > T substitution in the mother line previously published. B) Successful correction of the point mutation (WT nucleotide – C; Mutated nucleotide – T). C. Immunocytochemical detection in hiPSCs of the pluripotency markers OCT4, NANOG, SSEA3, SSEA4, TRA1-60 and TRA1-81. Scale bars correspond to 200 μm. D. Representative karyotype of A79V-GC-iPSCs.](image-url)
sgRNA cutting site, generating a PCR product of 409 base pairs: A79V_Forward2 5′-TCCCAGGTCTAACCGTTACCT-3′ and A79V_Reverse2 5′-ATGCAGAGGCCTTCAAGGTG-3′. PCR products were digested using the restriction enzyme EheI (ThermoScientific) according to the manufacturer’s instructions and clones showing correct digestion patterns were then sequenced to confirm the correction and that no frame-shifts or other mutations were introduced.

**Sequencing**

Sanger sequencing of exon 4 of PSEN1 gene was carried out in an ABI PRISM 310 Genetic Analyzer using a designed set of primers that flank the mutation site (A79V_Forward2 5′-TCCCAGGTCTAACCGTTACCT-3′ and A79V_Reverse2 5′-ATGCAGAGGCCTTCAAGGTG-3′) according to standard procedures.

**Immunofluorescence staining**

hiPSCs were fixed with 4% paraformaldehyde (PFA) in PBS for 15 min and stained by standard immunofluorescence staining procedures. The primary antibodies (Table 1) were visualized with the secondary antibodies Alexa488 or Alexa594 diluted 1:200 (Life technologies) and counterstained with Hoechst bisbenzimide 33,258. Images were acquired with a Leica DMRB-fluorescence microscope (Leica Microsystems, Wetzlar, Germany).

**Karyotyping**

A79V-GC-hiPSCs were treated for 45 min with KaryoMAX colcemid (Life Technologies), harvested in fresh fixative consisting of 25% acetic acid and 75% methanol and sent for G-band karyotyping (Institute of Medical Genetics and Applied Genomics, University of Tübingen, Tübingen, Germany).

**Verification and authentication**

Karyotyping was performed by Institute of Medical Genetics and Applied Genomics, University of Tübingen (Tübingen, Germany) and a minimum of 20 metaphases were analyzed. The results showed a normal 46, XX karyotype, without any detectable abnormalities (Fig. 1D). A79V-GC-hiPSC line identity and purity was confirmed by sequencing of PSEN1 (Fig. 1B) and ICC for pluripotency genes expression (Fig. 1C).

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**Table 1**

Primary antibodies used for immunocytochemistry.

<table>
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<th>Antibody and host species</th>
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<tr>
<td>Pluripotency Rabbit anti-NANOG</td>
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**References**

