Multiple DNA binding proteins contribute to timing of chromosome replication in *E. coli*

Riber, Leise; Frimodt-Møller, Jakob; Charbon, Godefroid; Løbner-Olesen, Anders

*Published in:*
Frontiers in Molecular Biosciences

*DOI:*
10.3389/fmolb.2016.00029

*Publication date:*
2016

*Document version*
Publisher's PDF, also known as Version of record

*Document license:*
CC BY

*Citation for published version (APA):*
https://doi.org/10.3389/fmolb.2016.00029
Multiple DNA Binding Proteins Contribute to Timing of Chromosome Replication in *E. coli*

Leise Riber †, Jakob Frimodt-Møller †, Godefroid Charbon and Anders Løbner-Olesen *

Section for Functional Genomics and Center for Bacterial Stress Response and Persistence, Department of Biology, University of Copenhagen, Copenhagen, Denmark

Chromosome replication in *Escherichia coli* is initiated from a single origin, oriC. Initiation involves a number of DNA binding proteins, but only DnaA is essential and specific for the initiation process. DnaA is an AAA+ protein that binds both ATP and ADP with similar high affinities. DnaA associated with either ATP or ADP binds to a set of strong DnaA binding sites in oriC, whereas only DnaA\textsubscript{ATP} is capable of binding additional and weaker sites to promote initiation. Additional DNA binding proteins act to ensure that initiation occurs timely by affecting either the cellular mass at which DNA replication is initiated, or the time window in which all origins present in a single cell are initiated, i.e. initiation synchrony, or both. Overall, these DNA binding proteins modulate the initiation frequency from oriC by: (i) binding directly to oriC to affect DnaA binding, (ii) altering the DNA topology in or around oriC, (iii) altering the nucleotide bound status of DnaA by interacting with non-coding chromosomal sequences, distant from oriC, that are important for DnaA activity. Thus, although DnaA is the key protein for initiation of replication, other DNA-binding proteins act not only on oriC for modulation of its activity but also at additional regulatory sites to control the nucleotide bound status of DnaA. Here we review the contribution of key DNA binding proteins to the tight regulation of chromosome replication in *E. coli* cells.

Keywords: *E. coli*, chromosome replication, DNA binding proteins, cell mass, initiation synchrony

**TIMING OF INITIATION OF CHROMOSOME REPLICATION IN *E. COLI***

Chromosome replication in *Escherichia coli* is initiated from a single replication origin, oriC. The oriC-encoded structural and functional instructions for initiation are well-described (Leonard and Mechali, 2013; Skarstad and Katayama, 2013). In brief, the minimal oriC contains two functional regions: the Duplex Unwinding Element (DUE), which comprises three AT-rich repeat sequences of each 13 bp, and the flanking DnaA Assembly Region (DAR) (Figure 1; Mott and Berger, 2007; Ozaki and Katayama, 2012). DnaA is the initiator protein responsible for DUE opening and for the recruitment of replisome components and is the only protein that is both essential and specific for the initiation process (Kaguni, 2011; Leonard and Grimwade, 2011). DnaA belongs to the AAA+ proteins (ATPases Associated with diverse Activities) and can bind both ATP and ADP with similar high affinities (Sekimizu et al., 1987). The DAR region contains high affinity DnaA Boxes (R1, R4, and R2) that bind both DnaA\textsubscript{ATP} and DnaA\textsubscript{ADP}, along with multiple low affinity sites (R3, R5/M, I1, I2, I3, C1, C2, C3, τ1, and τ2) that bind DnaA\textsubscript{ATP}.
Throughout most of the cell cycle oriC is bound by DnaA located at R1, R2, and R4. This origin recognition complex (ORC) serves dual purposes in setting the stage for proper orisome assembly and preventing premature DNA unwinding. The ratio of DnaA<sub>ATP</sub> to DnaA<sub>ADP</sub> varies through the cell cycle and the peak at about 70–80% DnaA<sub>ATP</sub> coincides with replication initiation (Kurokawa et al., 1999). In the current model for orisome formation, two converging DnaA<sub>ATP</sub> filaments are formed (Rozgaja et al., 2011). One filament originates from R4 and grows leftward. This R4-filament displaces Fis from its binding site next to R2, which allows IHF to bind its recognition sequence next to R1. IHF bends the DNA 180° thereby bringing R1 in proximity of R5 and allows for the formation of the rightward filament responsible for duplex opening at the DUE, DnaC assisted helicase loading and assembly of the replisome (Leonard and Grimwade, 2011; Ozaki et al., 2012). Following initiation, DnaA<sub>ATP</sub> is converted to DnaA<sub>ADP</sub> primarily by a process called regulatory inactivation of DnaA (RIDA), which is dependent on the Hda protein bound to ADP and the DNA-loaded β-clamp of the polymerase III holoenzyme (Kato and Katayama, 2001), and by the less efficient datA-dependent DnaA<sub>ATP</sub> hydrolysis (DDAH). DDAH takes place at datA and is dependent on IHF (Figure 1; Kasho and Katayama, 2013).

Coordination of initiations with Cell Mass Increase

A long standing observation is that initiation of chromosome replication occurs when a certain cellular mass per origin, the initiation mass, is reached (Donachie, 1968; Hill et al., 2012). This coupling of replication initiation to cell growth depends on the DnaA protein. Earlier studies indicate that accumulation of DnaA protein sets the time of initiation in the cell cycle especially around or below wild-type level (Løbner-Olesen et al., 1989).

On the other hand, a coordinated increase in DnaA<sub>ATP</sub> and DnaA<sub>ADP</sub> does not significantly increase initiation (Kurokawa et al., 1999; Flatten et al., 2015), suggesting that accumulation of DnaA<sub>ATP</sub> is insufficient to trigger initiation. However, in the absence of RIDA, where DnaA is mainly ATP bound, a modest increase in DnaA<sub>ATP</sub> level leads to excessive initiations from oriC (Riber et al., 2006; Fujimitsu et al., 2008), as does expression of a DnaA mutant protein insensitive to RIDA (Simmons et al., 2004). Together, this indeed suggests that accumulation of DnaA<sub>ATP</sub> triggers initiation, whereas this effect can be offset by a similar increase in DnaA<sub>ADP</sub> (Donachie and Blakely, 2003). The participation of DnaA<sub>ADP</sub> in orisome formation remains unclear (Leonard and Grimwade, 2015), but the above observations suggest that it affects initiation negatively. Overall, accumulation of DnaA protein during steady-state growth, along with the cell cycle specific peak in DnaA<sub>ATP</sub>/DnaA<sub>ADP</sub> ratio, determines the onset of initiation with little variation between individual cells.

Coordination of initiations within a Single Cell

In individual cells, initiation at all origins occurs within approximately 1/10 of the doubling time (Initiation period, I<sup>P</sup>; Figure 2A). Rapidly growing cells with overlapping replication cycles therefore predominantly contain 2<sup>n</sup> (n = 1, 2, 3) copies of oriC, referred to as initiation synchrony (Skarstad et al., 1986). Initiation synchrony depends on the immediate inactivation of newly replicated origins by sequestration. oriC contain 11 copies of the sequence GATC that are methylated by Dam methyltransferase and bound, i.e., sequestered, by SeqA when hemimethylated. Sequestration prevents DnaA binding to its weak sites in oriC (Nievera et al., 2006) for approximately 1/3 generation (Sequestration period, S<sup>P</sup>; Figure 2A) and serves to keep track of which origins have been initiated (Boye and Løbner-Olesen, 1990; Campbell and Kleckner, 1990; Lu et al., 1994). The ability to initiate all origins in synchrony could result from maintaining a high DnaA<sub>ATP</sub> level throughout I<sup>P</sup>. Alternatively the first origin initiated may release its DnaA<sub>ATP</sub> to assist in triggering successive initiations at remaining origins in a cascade-like manner to ensure that free DnaA<sub>ATP</sub> increases through I<sup>P</sup> and enforces synchrony (Løbner-Olesen et al., 1994). These models predict different outcomes for sequestration deficient cells. A high DnaA<sub>ATP</sub> level throughout I<sup>P</sup> would result in re-initiation(s) within I<sup>P</sup>, asynchrony and overinitiation. The cascade model predicts a delay between successive initiations due to newly initiated origins competing with old origins for a limited amount of DnaA<sub>ATP</sub>. The initiation frequency would be directly proportional with accumulation of DnaA<sub>ATP</sub> resulting in asynchrony but an unchanged overall initiation frequency, which is in accordance with experimental observations for Dam deficient cells (Boye and Løbner-Olesen, 1990; Løbner-Olesen et al., 1994).

Synchrony is only observed when I<sup>P</sup> < S<sup>P</sup> (Figure 2A). In cells with aberrant timing of initiation, the I<sup>P</sup> and S<sup>P</sup> periods change, i.e., either start earlier in the cell cycle at a decreased initiation mass, i.e., overinitiation, or are delayed with an increased initiation mass, i.e., underinitiation. Alternatively, the duration of I<sup>P</sup> and S<sup>P</sup> may change relative to each other, and when I<sup>P</sup> > S<sup>P</sup>, newly initiated origins, released from sequestration, compete with origins not yet initiated. Consequently, some origins are re-initiated while others are not initiated at all, leading to loss of synchrony (Olsson et al., 2003; Skarstad and Løbner-Olesen, 2003). This is exemplified by dam mutants without a sequestration period that initiate throughout the cell cycle (Figure 2B; Boye and Løbner-Olesen, 1990; Lu et al., 1994). seqA mutants are also asynchronous but have a higher origin concentration, possibly because DnaA is increased, relative to dam mutants (Figure 2C; Campbell and Kleckner, 1990; von Freiesleben et al., 1994). Increased levels of Dam will, due to faster re-methylation rates, reduce S<sup>P</sup> and when this becomes shorter than I<sup>P</sup>, asynchrony follows (Figure 2C; Boye and Løbner-Olesen, 1990; von Freiesleben et al., 2000a). Excess SeqA protein delays initiation, prolongs the sequestration period but does not affect synchrony (Figure 2D; Bach et al., 2003; Charbon...
et al., 2011). During sequestration the activity of DnaA is lowered by RIDA and DDAH. RIDA is presumably accelerated by generation of new replication forks at initiation and hence more DNA loaded β-clamps (Moolman et al., 2014). Similarly, DDAH is increased shortly after initiation when the datA locus is duplicated and together they ensure a post-initiation decrease in the DnaA_{ATP}/DnaA_{ADP} ratio (Figure 1). RIDA (Δhda) and to a lesser degree DDAH (ΔdatA) deficient cells fail to lower the ratio of DnaA_{ATP}/DnaA_{ADP} to prevent re-initiation following sequestration. This results in asynchrony and early initiation at a reduced cell mass (Figure 2E; Kitagawa et al., 1998; Fujimitsu et al., 2008; Kasho and Katayama, 2013). On the other hand, the dnaN_{G157C} mutant, which is more active in RIDA (dnaN encodes the β-clamp), or extra copies of datA, results in delayed initiation and, for dnaN_{G157C} cells, also produces asynchrony (Figures 2F,G; Morigen et al., 2001; Gon et al., 2006; Charbon et al., 2011; Johnsen et al., 2011). During sequestration, the overall level of free DnaA is reduced by titration (Hansen et al., 1991;
FIGURE 2 | Timing of replication initiation. Examples of mutants/plasmids with altered initiation (I^P; green) and sequestration (S^P; blue) periods. The horizontal line represents one doubling time, whereas the vertical (hyphenated) line illustrates the time of initiation of the first origin in wild-type cells. Note that the start of S^P always coincides with the first origin initiated, i.e., start of I^P. In the graphical representation of initiation synchrony, the number of origins per cell are on the X-axis, whereas the cell number is on the Y-axis of each histogram. When more than one mutation/plasmid is listed for a specific example (e.g., in C,E–G,I), the histograms are representative of the initiation phenotype of each individual mutation/plasmid.

MODULATION OF TIMING OF REPLICATION INITIATION BY DNA BINDING PROTEINS

Several DNA binding proteins affect either the cell mass at initiation, the initiation synchrony, or both. These proteins either bind specifically to oriC to affect DnaA binding, non-specifically to DNA to alter oriC topology, or they bind sequences important for the nucleotide bound status of DnaA.

Proteins That Specifically Interact with oriC Prior to Initiation

The most important protein to interact with oriC prior to initiation is DnaA. Mutations in DnaA that affect nucleotide binding, such as dnaA46, are presumably somewhat deficient in formation of DnaA multimers on oriC, which results in delayed initiation and a prolonged initiation period (Skarstad et al., 1988; Boye et al., 1996). As sequestration remains unchanged (I^g > S^g), dnaA46 cells are asynchronous (Figure 2F; Skarstad and Lobner-Olesen, 2003). Mutations in DnaA that affect DNA binding, but not nucleotide binding (e.g., dnaA204), lead to late but synchronous initiation (Figure 2G; Skarstad et al., 1988; Torheim et al., 2000). The ability to form DnaA^{ATP} filaments on oriC therefore seems of greater importance for initiation synchrony than a tight anchoring to DnaA binding sites.

Conflicting data exist on the role of Fis for timing of initiation. Binding Fis to oriC in vitro is reported to either inhibit initiation of replication by inducing conformational changes at oriC that prevent orisome formation (Wold et al., 1996; Ryan et al., 2002, 2004), or have no effect on initiation (Margulies and Kaguni, 1998). Cells with a mutated primary Fis binding in oriC (oriC131) have an origin concentration similar to wild-type (Figure 2H; Weigel et al., 2001; Riber et al., 2009; Flatten and Skarstad, 2013). Fis-deficient cells, on the other hand, have a lowered origin concentration (Flatten and Skarstad, 2013; Kasha et al., 2014), suggesting that initiation is delayed (Figure 2F). However, because Fis affects multiple cellular processes due to its involvement in DNA organization one should be careful in assessing its role in initiation solely based on the behavior of Fis-deficient cells. Both Fis deficiency or loss of its primary oriC binding site result in initiation asynchrony (Figures 2F,H; Riber et al., 2009; Flatten and Skarstad, 2013), indicating that these cells are deficient for proper orisome assembly and/or for preventing premature DNA unwinding. The role of IHF in replication timing is less controversial. An oriC mutant with a disrupted IHF binding site (oriC132) is somewhat deficient in orisome formation and has delayed but synchronous initiation (Figure 2G; Weigel et al., 2001; Skarstad and Lobner-Olesen, 2003; Riber et al., 2009). ihf mutant cells also initiate replication at an increased mass per origin consistent with a stimulatory role of IHF on initiation. Cells deficient in IHF are on the other hand asynchronous (Figure 2F; von Freiesleben et al., 2000b).

This is in agreement with an additional role of IHF for DnaA^{ATP} generation at DARS2 (see below).

A number of proteins negatively regulate initiation of replication in vitro. These include ArcA that binds to 13 mer AT rich repeats, to DnaA box R1 and to the IHF binding site in oriC, and IciA that binds to 13-mer AT-rich repeats in oriC (Hwang and Kornberg, 1990; Lee et al., 2001). The impact of ArcA and IciA on replication initiation in vivo is modest (Nystrom et al., 1996) or not known, respectively. The stationary-phase induced CspD protein binds ssDNA to inhibit replication initiation and elongation in vitro, whereas no in vivo data are available (Yamanaka et al., 2001). Upon association with Cnu and/or Hha, H-NS (see below) binds to a specific sequence in oriC that overlaps DnaA box R5 (Kim et al., 2005; Yun et al., 2012). Cells deficient in Cnu and/or Hha are, however, similar to wild-type (Kim et al., 2005). Finally, the protein Rob binds to a single site in oriC in vitro, but does not affect initiation in vivo (Skarstad et al., 1993).

DNA Binding Proteins That Affect Topology of oriC

In E. coli the genomic DNA is mostly negatively supercoiled (Wang et al., 2013). Unconstrained supercoiling of oriC contributes to the ease of duplex opening and is determined by transcription (not covered here; for review see Magnan and Bates, 2015) along with the actions of topoisomerase I and DNA gyrase enzymes (Wu et al., 1988). Mutations in topoisomerase I, which removes negative supercoils, result in initiation at a slightly reduced mass while synchrony is maintained (Figure 2I; von Freiesleben and Rasmussen, 1992; Olsson et al., 2003). Conversely, temperature sensitive gyrB mutant cells, with moderately reduced negative superhelicity of the chromosome, enhance the temperature sensitivity of a dnaA46 mutant (Filutowicz, 1980) and show delayed synchronous initiations (Figure 2G; von Freiesleben and Rasmussen, 1991; Usongo et al., 2013). This suggests that initiation is facilitated by an increase in negative superhelicity of the chromosome. However, topA-gyr mutations influence chromosome segregation, R-loop formation and possibly induce stable DNA replication independent of oriC (Usongo et al., 2013, 2016) making it difficult to assess the effect of large changes in overall supercoiling on replication initiation. In vivo, nucleoid-associated proteins (NAPs; Dillon and Dorman, 2010), such as IHF, Fis, H-NS, HU, and MukF constrain negative supercoils to condense the chromosome and could therefore affect initiation of chromosome replication (Badrinarayanan et al., 2015; Lal et al., 2016). H-NS deficient cells have an increased negative superhelicity of the genome (Mojica and Higgins, 1997; Hardy and Cozzarelli, 2005). Yet, genetic evidence suggests that loss of H-NS hampers initiation (Katayama et al., 1996), and H-NS deficient cells initiate replication in synchrony at an increased cell mass (Figure 2G; Kaidow et al., 1995; Atlung and Hansen, 2002). The HU protein can substitute for IHF in DnaA-mediated unwinding of oriC in vitro (Hwang and Kornberg, 1992) although their mechanisms of action differ (Ryan et al., 2002). In vivo, genetic evidence suggests that loss of HU stimulates initiation despite decreased negative
supercoiling (Louarn et al., 1984). Loss of MukB, involved in condensation of the bacterial chromosome (Hiraga et al., 1989; Cui et al., 2008), results in reduced negative supercoiling (Weitao et al., 2000), but initiations remain synchronous (Weitao et al., 1999). It is not known whether MukB affects the initiation mass. Finally, the starvation-induced NAP, Dps, binds non-specifically to oriC, and interacts with the N-terminus of DnaA, inhibiting DNA unwinding in vitro. Loss of Dps does not result in loss of synchrony, but increases the cellular origin content somewhat (Chodavarapu et al., 2008). In summary, it seems that NAPs modulate replication initiation but that the effect is not solely mediated through an effect on DNA supercoiling.

GETTING READY FOR THE NEXT ROUND OF REPLICATION

At later cell cycle stages DnaA<sub>ATP</sub> is regenerated for the next initiation to take place (Figure 1). E. coli can rejuvenate DnaA<sub>ADP</sub> to DnaA<sub>ATP</sub> in a process assisted by acidic phospholipids (Saxena et al., 2013) or at two non-coding chromosomal sites called DARS1 and DARS2 (Fujimitsu et al., 2009). DARS1 and DARS2 are located in each replichore halfway between oriC and terC, and are duplicated after the end of sequestration. Multiple DnaA<sub>ADP</sub> molecules form complexes with DARS to facilitate release of ADP resulting in apo-DnaA, which will primarily rebind ATP as this is more abundant than ADP within the cell (Petersen and Møller, 2000).

DARS1 is not known to be regulated by any proteins, whereas rejuvenation at the more efficient DARS2 locus is dependent on binding of both IFH and Fis (Kasho et al., 2014). While Fis binds DARS2 throughout the cell cycle, IFH provides cell cycle specificity to DARS2 activity by only binding and activating DARS2 immediately prior to initiation to ensure an increase in DnaA<sub>ATP</sub> level (Fujimitsu et al., 2009; Kasho et al., 2014). Extra copies of DARS1 or DARS2 will increase the overall DnaA<sub>ATP</sub> level, which results in early initiation (Figures 2E, I) and for DARS2 also extends Π<sup>E</sup>, thereby resembling RIDA deficiency (Figure 2E; Fujimitsu et al., 2009; Charbon et al., 2011). Deletion of DARS1, DARS2, or both reduces the ability to reactivate DnaA for new initiations in the following cell cycle and results in delayed initiation (Figures 2F, G; Fujimitsu et al., 2009; Kasho et al., 2014; Frimodt-Møller et al., 2015). Loss of DARS2 also increases the relative duration of the initiation period, leading to initiation asynchrony (Figure 2F; Fujimitsu et al., 2009; Frimodt-Møller et al., 2015). This suggests that both DARS1 and DARS2 are important for coupling initiation to cell mass increase, whereas only the cell-cycle regulated DARS2 is crucial for maintaining initiation synchrony.

CONCLUDING REMARKS

Overall, timing of chromosome replication in E. coli takes place at least at two levels. First, initiation of replication is tightly coupled to cell mass increase through accumulation of DnaA<sub>ATP</sub>. Second, synchrony of initiations within the single cell is not necessarily connected to initiation mass but results from each origin being simultaneously initiated only once per generation, with asynchrony originating from failure to obey this once-and-only-once rule. DnaA remains the only replication protein solely required for initiation at oriC, but additional proteins act on oriC and elsewhere to assist in coupling of replication to cell growth and synchrony. In particular IFH and Fis display complex functions, targeting several regulatory sites. IFH has a dual role on replication initiation, acting both positively (i.e., binding to DARS2 and oriC) and negatively (i.e., binding to datA). Also, IFH binds oriC at the pre-initiation stage and interacts with datA and DARS2 following initiation. Binding of IFH to these regions is suggested to be temporally regulated so that IFH binds to oriC, to datA and to DARS2 in a successive manner during cell cycle progression (Kasho and Katayama, 2013; Kasho et al., 2014). In vivo, ihf mutants display an initiation-compromised phenotype, indicating that the overall role of IFH on initiation of replication appears positive.

For a long time, the contribution of Fis in initiation regulation has been questioned. Recent studies do, however, suggest an overall positive role of Fis in replication initiation (Flatten and Skarstad, 2013; Kasho et al., 2014), which likely results from ensuring ordered orisome formation by preventing premature IFH binding and DNA unwinding (Leonard and Grimwade, 2015) and from stimulating DnaA<sub>ATP</sub> rejuvenation at DARS2. As the cellular Fis level depends on both growth-rate and phase, it could adjust chromosome replication to the bacterial growth rate through its activity on DARS2 (Kasho et al., 2014).

AUTHOR CONTRIBUTIONS

All authors listed, have made substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

This research was part of the Center for Bacterial Stress Response and Persistence (BASP) funded by a grant from the Danish National Research Foundation (DNRF120) and by a grant from the Novo Nordisk Foundation.

REFERENCES


Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer MM declared a past co-authorship with the author ALO to the handling Editor, who ensured that the process met the standards of a fair and objective review.

Copyright © 2016 Riber, Frimodt-Møller, Charbon and Løbner-Olesen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.