Real-time and label free determination of ligand binding-kinetics to primary cancer tissue specimens; a novel tool for the assessment of biomarker targeting

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A B S T R A C T

In clinical oncology, diagnosis and evaluation of optimal treatment strategies are mostly based on histopathological examination combined with immunohistochemical (IHC) expression analysis of cancer-associated antigens in formalin fixed paraffin-embedded (FFPE) tissue biopsies. However, informative IHC analysis depends on both the specificity and affinity of the binding reagent, which are inherently difficult to quantify in situ. Here we describe a label-free method that allows for the direct and real-time assessment of molecular binding kinetics in situ on FFPE tissue specimens using quartz crystal microbalance (QCM) enabled biosensor technology. We analysed the interaction between the rVAR2 protein and its placental-like chondroitin sulfate (pl-CS) receptor in primary human placenta tissue and in breast and prostate tumour specimens in situ. rVAR2 interacted with FFPE human placenta and cancer tissue with an affinity in the nanomolar range, and showed no detectable interaction with pl-CS negative normal tissue. We further validated the method by including analysis with the androgen receptor N-20 antibody (anti-AR). As the Kd value produced by this method is independent of the number of epitopes available, this readout offers a quantitative and unbiased readout for in situ binding avidity and amount of binding epitopes. In summary, this method adds a new and important dimension to classical IHC-based molecular pathology by adding information about the binding characteristics in biologically relevant conditions. This can potentially be used to select optimal biologics for diagnostic and therapeutic applications as well as guide the development of novel high affinity binding drugs.

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1. Introduction

Cancer is a capital health problem in the modern world and therefore a major focus of modern medical research. Currently, diagnosis and disease management of most types of cancer is based on histopathological examination, combined with a molecular pathology analysis of primary tumour and potential metastatic lesions [3]. Classical molecular pathology analysis involves IHC evaluation of biomarkers that can inform on disease status and cancer subtype, as well as on molecular signatures able to guide therapy. The standard analysis is two-dimensional in the sense that the output only informs on expression intensity and pattern of the biomarker(s) in question. Nevertheless, molecular pathology assessment plays an important role in the diagnosis and disease management of several types of cancer. For example, IHC staining intensity and pattern of ErbB2/HER2 is widely used to guide treatment in human breast cancer [4,10,30]. In prostate cancer, the expression intensity and pattern of the androgen receptor (AR) is used...
2. Materials and methods

2.1. Materials and reagents

Antibodies used for immunohistochemistry (IHC) and immunofluorescence (IF) were anti-AR rabbit polyclonal antibody (1:200, N-20, Santa Cruz, sc-816), anti-V5-FITC antibody (1:500, Life Technologies, 10308) and fluorescein anti-rabbit IgG antibody (1:500, Vector Laboratories, Fl-1000).

2.2. Immunohistochemistry

Tissue stainings were performed using the Ventana Discovery platform. Sectioned paraffin embedded tissue samples were deparaffinized in EZ prep solution (Ventana) and stained with V5-tagged rVAR2 or anti-AR (N-20, Santa Cruz) antibody. In brief, tissue sections were incubated in Cell Conditioning 1 (CC1) or Cell Conditioning 2 (CC2) solution (Ventana) to retrieve antigen, followed by incubation with primary staining molecules. Bound rVAR2 was detected with an anti-V5 antibody, and an anti-mouse-HRP detection step. Bound antibodies were incubated with universal secondary antibody and visualized using Streptavidin-biotin peroxidase detection system and 3,3′-diaminobenzidine as chromogen.

2.3. Tissue immobilization

Attana COP-1 cartridges were disassembled to retrieve the COP-1 gold chip. The chip was coated in poly-L-lysine solution 5min at RT (Sigma) prior to tissue immobilization. The chip was rinsed in PBS and allowed to dry at RT for 2h. The FFPE tissue, tested positive for the given target in IHC, was micro-dissected at 5µm thickness on a microtome. The tissue was mounted on the chip and baked at 60°C for 1h.

2.4. Chip deparaffinization and antigen retrieval

The tissue chips were deparaffinized in EZ prep solution at 65°C in a water bath for 30min, and rinsed three times in PBS. Antigen retrieval was performed in 10mM sodium citrate, 0.05% Tween, pH 6 for 30min at 95°C. The chip was again rinsed three times in PBS. For AR experiments, the tissue was permeabilized in 0.5% saponin in PBS 10min at RT. For antibody experiments the chips were blocked in 1% BSA, 10% FBS in PBS at 1h at RT. For rVAR2 experiments, the chips were blocked in Synbiol (Immunochemistry Technologies) solution for 1h at RT. The treated tissue chips were mounted in the COP-1 cartridges, being very careful not to allow leakage of fluid to under the chip. In the optimization step the tissue was stained with ethidium bromide, and the integrity of the tissue was viewed in a top-down fluorescence microscope.

2.5. Cell chip preparation

Cells were seeded on COP-1 surfaces (Attana AB) at 80.000 cells in 700 µL of appropriate supplemented growth media. Following a 24h incubation at 37°C, 5% CO₂, the cells were fixed 10min in 4% PFA in PBS. The cells were washed in PBS and the cartridge was assembled, being careful of leakage. In the optimization step the cells were stained with ethidium bromide, and viewed in a top-down fluorescence microscope.

2.6. Purification of pl-CS from human placenta

5g of frozen placental tissue was minced and extensively washed with cold PBS. The tissue was then treated with 10mg collagenase type IV (Life Technologies) in 10mL PBS containing Ca²⁺ and Mg²⁺, for 16h at 37°C. The resulting supernatant was cleared at 3000g for 10min. The pellet was further digested with Trypsin EDTA (Lonza) for 2h at 37°C. The resulting supernatant was cleared at 3000g for 10min. All the supernatants were combined and lyophilized. The dried extract was delipidated using a series of chloroform and methanol washes using 2:1, 1:1, 1:2 v/v ratios and dried at 55°C. The sample was then digested with 20mg Pronase (Roche) in 20mL 50mM Tris/HCl, pH7 and 5mM CaCl₂ for 16h at 50°C. After heat inactivation of the Pronase, 2mM MgCl₂ and 1000U Benzonase (Sigma-Aldrich) were added and the sample was incubated for 2h at 37°C. The sample was run on a DEAE column and eluted in 20mM NaOAc, pH 6, 1 M NaCl. The GAGs were precipitated in 3 volumes of ethanol 24h at 4°C. The precipitate was collected at 9000g for 30min and dried at 50°C. The dried sample was then resuspended in water and desalted using a PD-10 column (GE Healthcare). The sample was then treated with hyaluronidase from Streptomyces hyalurolyticus (Seikagaku Biobusiness Corporation, Japan), and A mixture of Flavobacterium heparinum heparin lyase I, II, and III in succession. The chondroitin sulfate was then precipitated in three volumes of ethanol, dried, and resuspended in water. The structure, purity and concentration was determined by MS.
2.7. Biotinylation of the pl-CS

Purified CS from the placenta was modified with an activated biotin reagent (sulfo-NHS-LC-biotin, Thermo-Pierce, Rockford, IL) at the amine group of the residual peptide left at the reducing end of CS chain after the trypsin digestion purification step. In Short the placental CS-peptide (100μg) in 50μL of 50mM phosphate buffer, pH 7.4, was mixed with the biotin reagent (1.0μmol) and incubated at room temperature for 2h. 5μL of 1M ethanolamine was then added to the mix and incubated at room temperature for 20min to terminate the reaction. The biotin-modified placental CS was purified on a Superose 12 HR 10/30 column (GE Healthcare, Piscataway, NJ) using 0.2M ammonium acetate as the eluent. The eluted product was finally freeze-dried three times. The placental CS-Biotin powder was resuspended in H2O prior to use in experiments.

For immobilization of the purified biotinylated pl-CS on Biotin Chips a biotin chip (Attana AB) was allowed to stabilize in 10mM HEPES, 150mM NaCl, 0.005% Tween-20 at a flow rate of 100μL/min on the Attana Cell 200 instrument. The flow rate was lowered to 20μL/min and 100μg/ml streptavidin (Attana AB) was injected and allowed to interact with the surface. The pl-CS-biotin was then diluted to 50μg/ml and injected on the surface for immobilization.

2.8. Kinetic analysis

The chips were inserted in the Attana Cell 200 instrument (Attana AB) and allowed to stabilize in running buffer at 25μL/min (100μL/min for the AR experiments). The purified pl-CS experiment and tests of monoclonal antibodies were run in PBS. rVAR2 cell and tissue experiments were run in PBS containing Symbloc diluted to 0.1× stock solution (Immunochrometry Technologies). Once stable, the baseline was verified with repeated injections of running buffer. The analyte to be tested was dissolved in the appropriate running buffer and diluted two-fold to yield the given concentration range. All injections were performed by the C-Fast autosampler (Attana AB). Baseline was checked between sample injections, by injections with running buffer. Blank injections were subtracted from the sample injections in the final analysis. The analyses were tested for reactivity against blank surfaces and against the given negative controls. The data was prepared using the Attana Attache software (Attana AB) and curve fitting was performed in the TraceDrawer software (Ridgeview Instruments) using the fit models given in Table 1. For some experiments a low level of background binding was accounted for by fitting a 1:2 model. Only the true high affinity values (Kd) values are listed in Table 1. For CA8 inhibition, rVAR2 at a concentration of 100nM was pre-incubated with 400μg/ml bovine CS (Sigma) and injected over the surface. For the verification of tissue adherence, rVAR2 was injected over the tissue surface until saturation. The lid on the COP-1 Cartridge was removed and the tissue incubated 30min with anti-V5-FITC (Invitrogen) at RT. The chip was washed and viewed under a top-down fluorescence microscope.

2.9. Immunofluorescence

The given cancer cell lines were seeded at a subconfluent concentration on glass slides and allowed to adhere at 37°C, 5% CO2. The cells were then fixed in 4% PFA and washed in PBS. The cells were blocked for 1h at RT in 1% BSA, 5% PBS in PBS. The cells were then stained with V5 tagged rVAR2 or anti-AR (N-20, Santa Cruz) for 1h at RT in PBS containing 0.25% BSA. The cells were washed and incubated with anti-V5-FITC or antio-mouse-FITC in PBS containing 0.25% BSA. The slides were stained with DAPI, mounted, and viewed using confocal microscopy.

For the measurement of fluorescence intensity in the Attana chip rVAR2 detection on the prostate and tonsil tissues, 7 distinct areas of the tissue were selected and quantificated using the ImageJ software. Data is presented as Corrected Total Tissue Fluorescence (CTTF), for which the following formula was used, as described before: CTTF = Integrated density − (Area of selected region × Mean fluorescence of the background) [2,20]. Quantification shown represents one measured experiment.

2.10. Microscopy

A Nikon C1 confocal microscope with a 60× oil objective was used for imaging the IF cell stainings. A total of 5 representative pictures were taken per sample. Stainings were repeated twice. For analysis of the integrity of the immobilized tissue, a Leica DMLB with a Rolera-rx Fast 1394 camera and a 20× objective was used.

3. Results

3.1. Preparation of FFPE tissue specimens on QCM biosensor chips

Chips for the Attana QCM biosensor come with a wide variety of surfaces for the immobilization of different ligands to be analysed. Traditionally the kinetics of biomolecule interactions was studied with purified ligand immobilized on polystyrene or covalently linked through primary amines to reactive surfaces [22]. Recently, cells were grown on polystyrene chips optimized for cellular adherence (COP-1) and either fixed to the surface using PFA [8,23] or tested as live cells [16]. Our aim was to immobilize a piece of FFPE primary human tissue to the chip for subsequent binding analysis. The cell COP-1 chips are plasma-treated to become hydrophilic, which is optimal for cellular adherence. We macro-dissected a small circular piece of placental tissue, at a thickness of 5μm and placed it in the middle of a COP-1 surface that was covered with Poly-L-Lysine and incubated at room temperature for 20min to yield the given concentration range. All injections were performed by the C-Fast autosampler (Attana AB).

The detection of protein targets in FFPE tissue requires removal of the paraffin matrix and often requires further treatment for the regeneration and availability of the target antigens. We performed deparaffinization in EZprep solution and antigen retrieval in 10mM sodium citrate, 0.05% tween, pH 6 [27]. The chips were visually inspected at each step to ensure the presence of the tissue on the chip.

Table 1

<table>
<thead>
<tr>
<th>Sample</th>
<th>Fit model</th>
<th>Kd_1/(M⁻¹s)</th>
<th>Kd_2/(s)</th>
<th>Kd_3/(nM)</th>
<th>Est. Error (nM)</th>
</tr>
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<tbody>
<tr>
<td>rVAR2 Placental</td>
<td>1:2</td>
<td>9.2 × 10⁴</td>
<td>3.9 × 10⁻⁴</td>
<td>4.3</td>
<td>±0.01</td>
</tr>
<tr>
<td>Prostate cancer tissue</td>
<td>1:1</td>
<td>1.27 × 10⁴</td>
<td>11.3 × 10⁻⁴</td>
<td>8.9</td>
<td>±0.06</td>
</tr>
<tr>
<td>Tonsil tissue</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Breast cancer tissue</td>
<td>1:2</td>
<td>7.9 × 10⁴</td>
<td>5.3 × 10⁻⁴</td>
<td>6.7</td>
<td>±0.3</td>
</tr>
<tr>
<td>Purified placental CS</td>
<td>1:1</td>
<td>12.3 × 10⁴</td>
<td>4.42 × 10⁻⁴</td>
<td>3.6</td>
<td>±0.05</td>
</tr>
<tr>
<td>LNCap cells</td>
<td>1:2</td>
<td>9.4 × 10⁴</td>
<td>5.2 × 10⁻⁴</td>
<td>5.5</td>
<td>±0.0004</td>
</tr>
<tr>
<td>SKBR3 cells</td>
<td>1:2</td>
<td>6.8 × 10⁴</td>
<td>10.5 × 10⁻⁴</td>
<td>15.6</td>
<td>±0.2</td>
</tr>
<tr>
<td>CHO+745A cells</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Prostate cancer tissue</td>
<td>1:2</td>
<td>13.0 × 10⁴</td>
<td>13.1 × 10⁻⁴</td>
<td>10</td>
<td>±0.04</td>
</tr>
<tr>
<td>Placental tissue</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>LNCap cells</td>
<td>1:2</td>
<td>7.4 × 10⁴</td>
<td>14.0 × 10⁻⁴</td>
<td>18.9</td>
<td>±0.07</td>
</tr>
<tr>
<td>PCI cells</td>
<td>1:2</td>
<td>23.7 × 10⁴</td>
<td>55.5 × 10⁻⁴</td>
<td>23.5</td>
<td>±0.6</td>
</tr>
</tbody>
</table>
To test the availability of the pl-CS in the immobilized placental tissue we performed a binding experiment using a titration of rVAR2 protein (Fig. 1b). The integrity of the tissue was confirmed by visualizing the tissue with ethidium bromide DNA staining, before and after the binding experiment. rVAR2 highly interacted with the tissue with a $K_D$ value of 4.3 nM. Furthermore, the interaction was CSA specific, as shown by the efficacy of inhibiting rVAR2 adhesion to the placental tissue with soluble CSA (Fig. 1c).

3.2. Affinity of the rVAR2:pl-CS interaction in primary tissue

We have shown that rVAR2 specifically interacts with pl-CS in the placenta and in most cancers, without binding CS on normal tissue [27]. This observation was mainly based on staining FFPE tissues with rVAR2 in IHC. We wanted to study the kinetics of the interactions and the relative abundance of pl-CS in patient-derived FFPE primary cancer tissue. For this purpose, we immobilized FFPE tissue from a prostate cancer and a breast cancer patient on COP-1 chips, and performed binding analysis with a titration of rVAR2 (Fig. 2a). A piece of tonsil tissue was used as the control for normal tissue. rVAR2 interacted to a high degree with both the prostate and breast cancer tissue with comparable $K_D$ values (8.9 nM and 6.7 nM, respectively). As previously observed [27], rVAR2 staining of the tonsil tissue in IHC was very weak, with only a limited amount of focal staining (Fig. 2a). In accordance with this rVAR2 did not interact with the tonsil tissue immobilized on the chip, in real time. To confirm these results and verify that rVAR2 interacted with the tissue, we disassembled the prostate cancer and tonsil chips following an experiment and confirmed the presence and localization of rVAR2 binding using an anti-V5-FITC antibody (Fig. 2b and c). This showed an intense staining of the prostate cancer tissue with very limited signal seen on the tonsil chip.

To date, biosensor analysis has relied on the interaction between an analyte and an immobilized cell or purified receptor [5,8,16,22,23]. In line with this, we have previously investigated the interaction between several recombinant VAR2CSA proteins and immobilized bovine decorin [6,7], as well as the interaction of rVAR2 with a melanoma cell [27]. To verify the interaction between rVAR2 and the immobilized tissue, we performed the same analysis on purified placental-like CS and cancer cells of prostate and breast cancer origin (Fig. 3). We purified pl-CS from placenta, biotinylated it through the peptide remaining from protease treatment, and immobilized it on an Attana biotin chip. As with the tissue, rVAR2 interacted highly with the pl-CS on the surface in the same concentration range and with a comparable $K_D$ value (3.6 nM) (Fig. 3a). To further verify the interaction we cultured an AR positive prostate cancer cell line (LNCap), and a breast cancer cell line (SKBR3) on the surface of a COP-1 chip, fixed them in PFA, and subjected them to the same analysis. Both cell lines were shown positive for the presence of pl-CS by rVAR2 staining in IF (Fig. 3b). The interaction analysis was comparable between the two cells lines and the analysis performed on the tissues and the purified pl-CS (Fig. 3b). A CHO cell...
selected for low xylotransferase activity (CHO-s745A) [9], and therefore low overall glycosaminoglycan (GAG) content, was used as the negative control for binding. This cell line was shown negative for the presence of pl-CS in IF, and was negative for the interaction with rVAR2 in the biosensor setup (Fig. 3b).

### 3.3. Affinity of an anti-AR antibody to prostate cancer tissue

To further extend the validation of the tissue biosensor setup, we selected a well-characterized cancer marker for which a validated monoclonal antibody is available. We analysed the interaction of the anti-Androgen Receptor (AR) N-20 antibody, used in the characterization of prostate cancer [13], with endogenous AR in cancer cells and in FFPE prostate cancer tissue. LNCap cells are known to express high levels of AR [21]. In accordance with this the LNCap cells was stained by anti-AR in IF and the anti-AR antibody interacted with the LNCap cells immobilized on a COP-1 surface, with a Kd value of 18.9 nM (Fig. 4a). Contrary to this PC3 cells, which are known to be androgen independent [32], showed very little AR staining in IF and a corresponding low level of interaction with the AR-antibody when immobilized on the biosensor chip (Fig. 4a). Having shown that the antibody worked in the biosensor setup, we selected an AR positive FFPE primary prostate cancer tissue biopsy, immobilized it on a COP-1 surface, and performed binding analysis on the Attana Cell 200. As with the rVAR2 analysis, interaction to the tissue was comparable to that of the immobilized AR positive cells (Kd value of 10 nM) (Fig. 4b). A piece of AR negative placental tissue showed no interaction with the AR antibody (Fig. 4c), confirming the target specificity. When comparing the peak responses measured in Hz between rVAR2 and anti-AR binding to the primary tissues it is evident that rVAR2 binding is up to 10 fold higher than the with the antibody.

### 4. Discussion

Discovery and characterization of cancer-associated markers effective in cancer diagnosis and treatment is a main focus in modern
medicine. Several targets have been described\textsuperscript{[13,14]}. Furthermore, we recently showed that the malarial VAR2CSA protein targets a specific type of placental-like chondroitin sulfate found in the placenta and on most cancer cells\textsuperscript{[27]}. The field of medical science increasingly recognizes that a thorough molecular analysis of the biomarker recognition event is needed for full evaluation of a target for diagnostic and treatment purposes\textsuperscript{[11]}. Such thorough analysis requires an in depth understanding of the molecular mechanism underlying the specific interaction as well as its kinetic properties. For this purpose scientists are utilizing a wide range of techniques including nuclear magnetic resonance (NMR)\textsuperscript{[12,26]}, mass-spectrometry (MS)\textsuperscript{[12]}, Enzyme-linked-immunosorbent-assay (ELISA), isothermal titration calorimetry (ITC)\textsuperscript{[15]}, and biosensor technology\textsuperscript{[1,5,19,22,24,31]}. Common for these techniques is a requirement for the isolation and purification or recombinant expression of the target molecule. While these methods are valuable in describing a given biomarker and how to target it effectively, biomarker extraction is laborious and may interrupt key features only present in the biomarker’s, or target’s, native environment.

Current molecular pathology relies on the intensity and pattern of expression of the given biomarker using two-dimensional histopathological and IHC analysis. This work aims at adding an extra dimension of interaction time to classic pathology by performing kinetics of binding analysis directly on FFPE primary tissue specimens. We believe that this provides pathologists with a tool to evaluate disease specific targets not only by their presence and specific localization, but also by their molecular characteristics and how this relates to its direct interaction with a targeting molecule.

We macro-dissected out a small circular piece of paraffin-embedded tissue and put it on COP-1 Attana QCM chip surfaces. We then removed the paraffin and performed antigen retrieval. One problem was that the tissue did not stick sufficiently to non-treated COP-1 surfaces. A coating with poly-L-lysine was needed for proper immobilization. The antigen retrieval needed was optimized by IHC staining using the analytes in question. Subsequently, the tissue-coated chips were treated in an identical way, allowing for direct comparison. Since no light can go through the gold COP-1 surface, we visualized the immobilized tissue by a DNA stain and a fluorescent microscope before and after the experiments to ensure the integrity of the tissue. No change in morphology was seen, showing that the proposed method maintains proper tissue integrity for subsequent analysis. The exact epitope of recognition of the anti-AR (N-20) antibody and its molecular nature is not known. It is likely that antibody specific optimization on tissue treatments is needed to accommodate analysis of other antibodies, such as antibodies targeting conformational epitopes.

Having optimized the immobilization of the FFPE tissue, we performed binding experiments using rVAR2 and Anti-AR (N-20) monoclonal antibodies on tissues verified for the presence of the target antigen in question. All binding experiments were verified against appropriate cell lines, and in the case of rVAR2 also purified pl-CS. rVAR2 showed similar binding kinetics to purified placental-like CS, cell lines, and tissue. This confirms our strategy and supports the use of primary tissue for QCM experiments. It is common to evaluate specific binding in biosensor experiments by subtracting binding to a blank reference\textsuperscript{[25]}. However, in working with immobilized cells and tissues we found
that binding of an analyte to a blank chip is not representative of the background seen in binding to a chip coated with cells or tissues. When investigating the interaction between an analyte and a purified receptor in classic SPR, the surface area available for non-specific interactions on the chip can be considered comparable. This is not the case when investigating a chip covered in tissue or cells. Subtracting the signal from the reference chip therefore introduced a negative bias on the ‘true’ interaction, visualized as a negative spike in the real-time observation of the on-rate (Data not shown). Instead we selected cells and tissues negative for the ligand in question as proper negative controls, and presented them as separate figures to demonstrate the background independently. In this, rVAR2 did not interact with normal tissue (tonsil), confirming our previous observations that rVAR2 staining is placenta and cancer specific [27]. Furthermore, rVAR2 did not interact with the CS negative CHO-A475a cells and binding could be inhibited with soluble CSα, confirming the CS specificity. The analysis
of the AR interaction showed the same correspondence with AR expression in both cell line and tissue setups. It is notable that such analysis was possible using a nuclear target.

Interestingly the data shows that pl-CS is abundantly present in tumour tissue compared to AR. This is evident from the high difference in max binding responses, with the Bmax response for rVAR2 being up to 10 times higher. This difference is not obvious in the IF staining in max binding responses, with the Bmax response for rVAR2 beingmour tissue compared to AR. This is evident from the high difference was possible using a nuclear target.

is interesting for cancer therapy as targeting an abundant target may offer a more efficient anti-cancer effect.

We demonstrate that this novel method can be broadly used to determine the relative presence of a cancer specific target as well as the affinity of the targeting reagent for its receptor. Implementation of the method has the potential to inform cancer diagnostics as well as provide an important tool for screening of high affinity targeting reagents in drug development, and aid in patient stratification, companion diagnostics and dosage determination during therapeutic treatment.

Contributions

T.M.C, A.S., M.D., and T.G.T., designed the research; T.M.C., M.A.P., H.Z.O., Y.M., N.S., M.R., and J.L., performed the experiments; L.F., provided useful reagents and helpful discussions; T.M.C., A.S. and M.D. wrote the manuscript.

Conflicts of interest

The authors declare no conflicts of interest.

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