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Genetic Basis for *Saccharomyces cerevisiae* Biofilm in Liquid Medium

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**ABSTRACT** Biofilm-forming microorganisms switch between two forms: free-living planktonic and sessile multicellular. Sessile communities of yeast biofilms in liquid medium provide a primitive example of multicellularity and are clinically important because biofilms tend to have other growth characteristics than free-living cells. We investigated the genetic basis for yeast, *Saccharomyces cerevisiae*, biofilm on solid surfaces in liquid medium by screening a comprehensive deletion mutant collection in the Σ1278b background and found 71 genes that were essential for biofilm development. Quantitative northern blots further revealed that AIM1, ASG1, AVT1, DRN1, ELP4, FLO8, FMP10, HMT1, KAR5, MIT1, MRPL32, MSS11, NCP1, NMR1, PEP5, PEX25, Rim8, Rim101, Snf8, SPC2, STB6, STP22, Tec1, Vfd24, Vps20, Vtc3, Ybl029W, Ybl029c-A, Yfl054c, Ygr161W-c, Yl014c-A, Yir024c, Ykl151c, Ynl200c, Yor034c-A, and Yor223w controlled biofilm through FLO11 induction. Almost all deletion mutants that were unable to form biofilms in liquid medium also lost the ability to form surface-spreading biofilm colonies (mats) on agar and 69% also lost the ability to grow invasively. The protein kinase A isoform Tpk3p functioned specifically in biofilm and mat formation. In a tpk3 mutant, transcription of FLO11 was induced three-fold compared with wild-type, but biofilm development and cell–cell adhesion was absent, suggesting that Tpk3p regulates FLO11 positive posttranscriptionally and negative transcriptionally.

The study provides a resource of biofilm-influencing genes for additional research on biofilm development and suggests that the regulation of FLO11 is more complex than previously anticipated.

Many microorganisms have the ability to form the multicellular, sessile, surface-bound communities known as biofilms. Biofilm formation has been described in prokaryotes such as the Gram-negative *Pseudomonas aeruginosa* and the eukaryotic yeasts *S. cerevisiae*, *Candida albicans*, and *Candida glabrata* (Hawser and Douglas 1994; Reynolds and Fink 2001; Hall-Stoodley et al. 2004). Cells in biofilms are reported to have a higher degree of diversity, so they have more possible fates than cells in free-living planktonic form. One consequence of this diversity is the high number of antibiotic-persistent cells in *P. aeruginosa* biofilms, a result of the high frequency of slow-growing or dormant cells in mature biofilms (Nguyen et al. 2011).

Although the molecular basis for biofilm development and biofilm cell diversification has been studied extensively in bacteria, less is known about the genetic basis for biofilm formation and cell diversification in eukaryotic microbes such as yeasts. Laboratory *S. cerevisiae* strains have, in some cases, been selected to not form biofilms (Liu et al. 1996). The trait can reappear in suppressor mutants that derepress expression of the cell wall protein Flo11p or other members of the Flo protein family that induce cell–cell adhesion (Fichtner et al. 2007; Torbensen et al. 2012). The *S. cerevisiae* strain Σ1278b naturally forms biofilm in liquid medium on solid surfaces such as polystyrenes because it expresses Flo11p (Reynolds and Fink 2001). In addition to its importance for biofilm formation, Flo11p is also essential for other morphotypes, including haploid-invasive growth on complex solid medium and diploid-pseudohyphal growth.
A fourth Flo11p-dependent phenotype is a type of giant colony that develops on semisolid complex mediums at room temperature. The giant colonies have been denoted surface-spreading biofilm as well as mats in the literature (Reynolds and Fink, 2001; Ryan et al. 2012). Although biofilm in liquid medium, surface-spreading biofilm (mats), invasive, and pseudohyphal growth are dependent on FLO11, they cannot be expected to be regulated in identical fashions because the growth conditions required for induction of these phenotypes are different.

The FLO11 gene is located in the middle of the right arm of chromosome IX (Lo and Dranginis 1996) and has a 2.8-kb promoter (Rupp et al. 1999). The relatively large promoter contains an extensive set of cis-acting elements that respond to multiple signaling pathways (Brückner and Mösch 2011). FLO11 is regulated by a mitogen-activated protein kinase (MAPK) pathway via the Ste12p/Tec1p transcription factors (Roberts and Fink 1994; Köhler et al. 2002; Rupp et al. 1999). The pH-sensitive Rim101p pathway regulates FLO11 (Barrales et al. 2008; Bayly et al. 2005; Lamb and Mitchell 2003), and the response is believed to include components from the endosomal sorting complex required for transport (ESCRT), because ESCRT I, II, and III proteins are required for activation of Rim101p and transcription of FLO11 (Sarode et al. 2011; Xu et al. 2004). Nutrient levels regulate FLO11 transcription through other pathways. Amino acid levels influence FLO11 transcription via the general control nonderepressible (GCN) pathway (Braus et al. 2002), which induces transcription on amino acid starvation (Lucchini et al. 1984). The presence of amino acids induces the Ssy1p-Ptr3p-Ssy5p-sensor complex transcription on amino acid per- meases (Torbensen et al. 2012). Furthermore, glucose depletion induces FLO11 via the AMP kinase homolog Snf1p by inactivating the transcriptional repressors Nrg1p and Nrg2p (Kuchin et al. 2002; Van De Velde and Thevelein 2008). Low glucose is also known to induce transcription of FLO11 through G protein-coupled glucose receptor Gpr1p, cAMP (Van De Velde and Thevelein 2008), the protein kinase A (PKA) isofrom Tpk2p, and the competing transcription regulators Sfl1p and Flo8p (Robertson and Fink 1998; Rupp et al. 1999). FLO11 is repressed when Sfl1p is bound and a noncoding RNA is transcribed in the FLO11 promoter and in a transcriptionally permissive state when Flo8p is bound to the FLO11 promoter and the ncRNA gene is transcriptionally inactive (Bumgarner et al. 2009). Flo8p activity is thought to facilitate the binding of other positive transcription factors such as Tec1p, Ste12p, and Pol II that reinforce the active state of the FLO11 promoter (Bumgarner et al. 2012). An interesting aspect of FLO11 regulation is the toggle switch that results from competition between Sfl1p and Flo8p and leads to variegated FLO11 expression (Bumgarner et al. 2009, 2012). Because of variable expression, only a subpopulation of cells expresses FLO11 and contributes to cell-cell adhesion. Variegated FLO11 expression is seen in pseudohyphal and invasive growth and could play a role in development of biofilm in liquid medium.

In contrast to FLO11 induction through active Tpk2p, the PKA Tpk3p is reported to repress FLO11. This has been shown with tpk3 mutants that have three-fold higher levels of FLO11 mRNA than wild-type TPK3 cells and show more robust invasive and pseudohyphal growth (Robertson and Fink 1998). Because FLO11 expression is repressed in a tpk3 tpk2 double mutant, Tpk3p is thought to inhibit Tpk2p activity (Robertson and Fink 1998). FLO11 regulation has mainly been investigated under conditions in which cells grow invasively or form mats or pseudohyphae (Brückner and Mösch 2011). Because the growth conditions essential for biofilm in liquid medium are very different from those favoring mats and pseudohyphal and invasive growth, it is unknown if the transcriptional program that regulates biofilm also regulates the other FLO11-dependent phenotypes. On abiotic surfaces, biofilms are formed in synthetic media with glucose as the carbon source (Torbensen et al. 2012; Reynolds and Fink 2001), whereas surface-spreading biofilm formation and invasive growth both occur on complex solid medium, and pseudohyphal growth is formed by diploid cells on solid, synthetic, nitrogen-poor medium (Reynolds and Fink 2001; Gimeno et al. 1992; Roberts and Fink 1994). A recent screen for genes essential for mat formation and invasive and pseudohyphal growth found limited overlap between genes regulating the three phenotypes (Ryan et al. 2012), suggesting a dedicated transcriptional program for FLO11-dependent biofilm formation. In addition to FLO11, other FLO genes and conditions might influence biofilms, including genes regulating the extracellular matrix that strengthens the three-dimensional structure of biofilms (Kuthan et al. 2003; Vachova et al. 2011; Guo et al. 2000), and quorum signaling that might coordinate the developmental program. Quorum signaling is reported for yeast, but its involvement in biofilm development is unknown (Chen and Fink 2006; Smukalla et al. 2008; Palkova et al. 1997).

In the current work, we investigated the molecular basis for Σ1278b biofilm development, including the extent to which biofilm development was dependent on FLO11. To do this, we screened a global collection of deletion mutants in the Σ1278b background for biofilm-forming ability on a solid abiotic surface. The extent to which the molecular program for biofilm formation overlapped with other FLO11-dependent phenotypes was investigated by testing biofilm-deficient mutants for ability to form mats and grow invasively. We found that a substantial fraction of Σ1278b cells were not part of the biofilm but existed as planktonic cells. Genes involved in the planktonic phenotype were identified by screening for Σ1278b mutants with more biofilm-phenotype cells in the total cell mass. Our study gives comprehensive insight into the molecular program controlling biofilm development in the genetic tractable yeast S. cerevisiae.
into synthetic medium to OD_{600nm} 0.1 for 2 hr before biofilm assays. Assays were in 200 μL in flat-well polystyrene microtiter plates (Frisenette). Cell density of the total population or of biofilm or planktonic subpopulations was recorded at OD_{430nm} at indicated time points. Planktonic subpopulations were measured by removing nonadhering cells by pipetting and measuring cell density in new microtiter wells. Biofilm subpopulations were measured by addition of 200 μL fresh medium to adhering cells and measuring OD_{595nm}. To visualize biofilms, crystal violet (HT901-8FOZ; Sigma-Aldrich) was added to wells for 24 hr at a final concentration of 0.05%. Planktonic cells and medium were removed and wells were washed four times with 200 μL H₂O. Biofilms were dried and resuspended in 170 μL 96% ethanol for 1 hr. Biomass was determined at OD_{595nm}.

**Biofilm screens of the deletion strain collection**

Deletion mutants were grown on solid YPD for 2 d. Cells were subsequently transferred to 96-well flat-bottom microtiter plates (Frisenette) containing 200 μL SC medium with 0.2% glucose. Cells were propagated for 46 hr or 96 hr at 30°C and biomass was determined at OD_{600nm} using a Synergy 1H Hybrid Reader (Biotek). Normalized biomass was calculated by pipetting and measuring cell density in new microtiter wells. Planktonic subpopulations were measured by removing nonadhering cells, and medium were used to wash four times with 200 μL H₂O. Biofilms were dried and resuspended in 170 μL 96% ethanol for 1 hr. Biomass was determined at OD_{595nm}.

**Data analysis**

Crystal violet biofilm measurements for Σ1278b YS-11 mutants were normalized to the total biomass and log-transformed as ln(OD_{595nm}/OD_{600nm}). Normalized biofilm scores were used to determine the median biofilm score for each mutant for both 46 hr and 96 hr. Median values were used for all further analysis (Supporting Information, File S5 and File S6). Samples with a total biomass less than OD_{600nm} < 0.01 were excluded from analysis. Replicate biofilm assays of the parental YS-11 strain (n = 288) showed normalized biofilm values that followed a Gaussian distribution at both 46 hr and 96 hr. The average normalized biofilm value ±2σ of the parental strain was used to determine mutants that had a significantly different biofilm score at 46 hr and 96 hr. Mutants with median-normalized biofilm scores less than 0.584 were considered to form significantly less biofilm than the parental strain and mutants with scores more than 1.972 were considered to form significantly more biofilm than the parental strain.

**Synthetic genetic array**

Selection of tpk3 geneX double mutants in the parental (MATa can1Δ::STE2p-pHi5 SLY113 STE3p-LEU2 his3::HisG leu2Δ ura3Δ) background was essentially conducted as described previously (Tong et al. 2001).

**Confocal laser scanning microscopy**

Overnight cultures were diluted to OD_{600nm} 0.1 in synthetic complete medium (0.2% glucose) and incubated on Rinzl plastic coverslips (Electron Mictoscopy Sciences) for 96 hr at 30°C. Biomasses were washed twice in saline before 30 min of FUN-1 (Invitrogen) staining. Imaging used a plan-Neofluar 40×/1.3 oil microscopy objective with a Zeiss LSM510 microscope.

**RNA FISH**

RNA FISH was conducted essential as described previously (McIsaac et al. 2013) using Stellaris probes from Biosearch Technologies. Thirty-five 20mer Quasar 670 probes covering the stretch +9 to +708 of FLO11 were used for detection of FLO11 mRNA, whereas 35 20mer Quasar 570 probes covering the stretch +2 to +701 of ACT1 were used as positive hybridization control. Precultures were grown overnight at 30°C in synthetic medium. Cells were subsequently inoculated into synthetic medium to OD_{600nm} 0.1 for 2 hr before fixation. Cells were fixed with 3% paraformaldehyde for 30 min at 30°C followed by 4-hr incubation at 5°C. Expression of FLO11 was subsequently recorded with a Zeiss LSM780 microscope. Cells were counted as recordable if they were labeled with the ACT1 probe. All ACT1 positive cells were subsequently recorded for their expression of FLO11 mRNA by counting cells with one or more red foci as positive for FLO11 mRNA. A total of 1329 wild-type cells, 1167 sfl1 cells, and 542 flo11 cells were counted as blinded samples. The flo11 cells served a negative control for FLO11 mRNA expression. None of the flo11 cells showed any signal for FLO11 mRNA labeling.

**Glucose concentration**

Glucose concentration was determined enzymatically with a glucose assay kit (GAGO-20; Sigma-Aldrich) adjusted for use with microtiter plates.

**Invasive growth**

Invasive growth was assayed essentially as described previously (Roberts and Fink 1994). Cells were patched on solid 2.0% agar YPD using an inoculation loop and propagated for 3 d at 30°C. Plates were covered with water for 30 min and shaken carefully, and nonadhering cells were removed. Complete colonies or colonies with portions remaining in or on the agar were categorized as invasive.

**Mat formation**

Cells were patched in the center of 25 ml YPD with 0.3% agar and incubated for 5 d at room temperature (22°C–25°C). Colonies with a structured hub and spokes were categorized as mat formers. Colonies that were completely smooth were categorized as having lost the ability to form mats.

**Northern blot**

RNA for northern blot was purified from mutants and the parental strain, YS-11, grown for 96 hr in square 120-mm × 120-mm Petri dishes (Frisenette). Total RNA was purified as previously described (Torbensen et al. 2012). All samples were treated with DNase and tested for removal of DNA by PCR using ACT1 primers 5’-TGGATTCTGGTATGTCTCAG-3’ and 5’-GAACGAGCTGGTGAAACACC-3’. Samples that still contained DNA were treated further with DNase until no trace of DNA was detected by PCR. DNAse-treated RNA, 2 μg in 3 μL, was dropped onto Hydrobond-N+ membranes (GE Healthcare) and dried. Membranes were wrapped in plastic wrap and RNA was cross-linked by UV light for 30 sec at 302 nm. One set of membranes was hybridized to a FLO11 [32P]-labeled probe and another set was hybridized to a [32P]-labeled ACT1 probe. Hybridization and probes were as described (Torbensen et al. 2012). Hybridization to each dot was recorded with a Storm 840 phosphorimager (Bio-Rad), and Molecular Dynamics ImageQuant TL software was used for quantification.
RESULTS

*S. cerevisiae* Σ1278b formation of mixed biofilm and planktonic populations is FLO11-dependent

In low glucose, haploid *S. cerevisiae* Σ1278b forms biofilm on polystyrene (Reynolds and Fink 2001). We tested this phenotype in liquid synthetic medium with different carbon sources and found that haploid Σ1278b formed a biofilm when cultured in 0.2% glucose, 1% maltose, 1% glycerol, or 1% ethanol, whereas biofilm formation was repressed in 2% glucose (Figure 1A). Further examination of biofilms formed in 0.2% glucose revealed that only 25% of cells were part of the biofilm (Figure 1B). Confocal laser scanning microscopy (CLSM) revealed that the proportion of biofilm-forming cells decreased substantially in a flo11 mutant, confirming that FLO11 was responsible for biofilm formation (Figure 1, C and D). Fluorescent in situ hybridization (FISH) revealed that only a minor fraction of reference cells expressed FLO11 mRNA (21%, n = 1329) in the synthetic 0.2% glucose medium (Figure 1F) corresponding to the fraction of cells participating in biofilm (Figure 1B). The proportion of cells expressing FLO11 mRNA increased to 73% (n = 1167) in a sfl1 mutant (Figure 1H), suggesting that SFl1p took part in participation of cells in a planktonic and a biofilm-forming subpopulation (Figure 1B). This hypothesis was supported by CLSM of biofilm formed by sfl1 and the reference strain (Figure 1, C and E). Both strains formed microcolonies, and microcolonies formed by the sfl1 mutant appeared to be larger than those of the reference strain (Figure 1, C and E and Figure S1). Based on these initial experiments, we expected that other molecular factors involved in biofilm development and repression of biofilm could be found.

Screening of a Σ1278b deletion collection for genes essential for biofilm development

To identify genes essential for biofilm in liquid medium in Σ1278b, as well as genes that regulate the proportion of biofilm-forming cells in a population, we screened a complete library of Σ1278b haploid mutants deleted in 4019 nonessential genes for biofilm-forming ability. The mutants were constructed by substituting each open reading frame (ORF) in Σ1278b (MATA can1Δ::STE2p-spHIS5 lyp1Δ::STE3p-LEU2 his3Δ::HisG leu2Δ::ura3Δ) with a kanMX cassette derived from the S288c deletion collection (Ryan et al. 2012). Thus, mutant alleles in the Σ1278b mutant collection were identical to the mutant alleles in the S288c collection (Giaever et al. 2002).

The biofilm screen was conducted by growing each of the 4019 mutants in liquid synthetic complete medium with 0.2% glucose and ammonium. Biofilm development was tested in triplicate after 46 hr and 96 hr by staining with crystal violet and removing planktonic cells. Mutants varied greatly in the amount of biomass formed (Figure 2A). To compensate for differences in biomass, values from crystal violet staining were normalized to the total cell mass of planktonic and biofilm-forming cells. (OD450nm). Median normalized biofilm values ln(OD450nm/OD600nm) were calculated for each mutant at 46 hr and 96 hr (Figure 2, C and D and File S1, File S2). Biofilm formation by the parental strain (n = 288) followed a Gaussian distribution. Thus, the average normalized biofilm value ± 2σ for the parental strain was used to identify mutants that formed significantly more or less biofilm than the parental strain (P < 0.05). We found that 137 mutants formed significantly less biofilm after 46 hr and, of these, 71 mutants still formed significantly less biofilm after 96 hr (Figure 2, C and D, blue bars, and File S3). A larger set of 427 mutants formed significantly more biofilm after 46 hr (Figure 2C, yellow bars). After 96 hr, 371 mutants formed significantly more biofilm than the parental strain (Figure 1, C and E). Both strains formed microcolonies, and microcolonies formed by the sfl1 mutant appeared to be larger than those of the reference strain (Figure 1, C and E and Figure S1). Based on these initial experiments, we expected that other molecular factors involved in biofilm development and repression of biofilm could be found.

**Figure 1** *S. cerevisiae* Σ1278b forms mixed populations of biofilm and planktonic cells. (A) Wild-type cells grown at 30°C in polystyrene wells in synthetic complete (SC) media in indicated carbon and nitrogen sources (Gln = glutamine 100 mM; Pro = proline 100 mM; NH4+ = ammonium 100 mM) for 96 hr and stained with crystal violet. Dark staining indicates biofilm formation. (B) Biofilm formation on polystyrene by a wild-type (wt) population in SC 0.2% glucose and NH4+. Blue triangles = nonadhering planktonic cells; green circles = biofilm-forming cells. Planktonic cells were separated from biofilms by pipetting. Cell density was measured at OD450nm. Black circles = measured glucose concentration left in growth medium over time. (C, D, E) Biofilm was recorded with confocal laser scanning microscopy (CLSM) after 96 hr of growth in SC 0.2% glucose and NH4+. Blue triangles = nonadhering planktonic cells; green circles = biofilm-forming cells. Planktonic cells were separated from biofilms by pipetting. Cell density was measured at OD450nm. Black circles = measured glucose concentration left in growth medium over time. (C, D, E) Biofilm was recorded with confocal laser scanning microscopy (CLSM) after 96 hr of growth in SC 0.2% glucose and NH4+. Nonadhering cells were removed by a single gentle pipetting and adhering cells were dyed with FUN-1. White bar = 50 μm. (F, G, H) Single cell fluorescence in situ hybridization (FISH) of representative samples of cells grown in SC 0.2% glucose and NH4+ for 20 hr. Bars = 5 μm. (C, F) Wild-type. (D, G) flo11Δ::kanMX in the wild-type background. (E, H) sfl1Δ::kanMX in the wild-type background. MATA can1Δ::STE2p-spHIS5 lyp1Δ::STE3p-LEU2 his3Δ::HisG leu2Δ::ura3Δ.
parental strain (Figure 2D, yellow bars), with 100 mutants forming more biofilm at both time points (File S3). To obtain a conservative estimate of the genes involved in biofilm development and to avoid growth rate effects, we considered only mutants with significantly altered biofilm effects after both 46 hr and 96 hr. This resulted in 71 candidate genes essential for biofilm development and 100 genes that repressed biofilm development by maintaining a high proportion of planktonic cells (Figure 2, C and D, blue and yellow bars, and File S3).

**Biofilm genes form a large complex regulatory network**

To obtain insight into the mechanisms that regulate biofilm development, all 171 biofilm-related genes were grouped into functional categories according to their gene ontology (GO) process (Table 1). Among the genes with a positive effect on biofilm when deleted were several functional groups of genes encoding proteins with mitochondrial function (Table 1 and File S4), and genes encoding respiratory chain components \((P = 4.56 \times 10^{-3})\) and mitochondrial ribosomal proteins \((P = 2.19 \times 10^{-6})\) were especially overrepresented. To analyze if the increased proportion of biofilm-forming cells in these mutants was dependent on *FLO11*, we made *FLO11* northern blots of representative mutants with reduced mitochondrial function (Figure 3A and File S5). No change in *FLO11* expression was observed in seven of the tested mutants representing reduced mitochondrial function, suggesting that posttranscriptional mechanisms acted on the *FLO11* gene product, other *FLO* genes were responsible for the increase in biofilm, or mutants with impaired mitochondrial function affected a non-Flo-dependent biofilm mechanism.

Among the 71 genes that were essential for biofilm development were several that were essential for invasive and pseudohyphal growth (Table 1). These included components of the PKA pathway (*FLO8, RAS2, TPK3*), a MAP kinase pathway (*TEC1*), the Rim pathway (*RIM101, RIM8*), and the GCN pathway (*GCN4*), as well as *FLO11* transcription factors that have not been assigned to specific signaling pathways (*MIT1, MFG1, MSS11*) (Gagiano et al. 2003; Ryan et al. 2012; Cain et al. 2012; Braus et al. 2003; Barrales et al. 2008; Roberts and Fink 1994; Rupp et al. 1999; Robertson and Fink 1998). We also found genes encoding components of the ESCRT complexes that are essential for regulation of *FLO11* (*SNF8, STP22, VPS36, VPS20, VPS25*) (Sarode et al. 2011). Additionally, the GO revealed a number of genes not previously associated with *FLO11* expression such as *AGT9* and *VTC3*, which are involved in membrane invagination, the vacuolar amino acid transporter *AVT1*, and *NPR1*, a regulator of amino acid transporter endocytosis. Identification of these genes suggested that vacuolar function or transport of one or more proteins to the vacuole was essential for biofilm development. In addition, there is also a group of genes involved in NADH repair such as NADHX epimerase (*YNL200C*) and NADHX dehydratase (*YKL151C*). In all, we identified 58 genes not previously associated with biofilm formation or *FLO11* expression (File S3).

Because formation of biofilm in the parental strain was dependent on *FLO11* expression (Figure 1D), we tested the level of *FLO11* mRNA in the mutants that did not form biofilm. Quantitative northern blots for *FLO11* mRNA revealed that 38 mutants had significantly less...
**Table 1** Biofilm gene GO processes for 71 genes that were essential for biofilm development and 100 genes that induced biofilm when deleted (all gene names are given in Table S3)

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<tr>
<th>GO Process Annotation</th>
<th>Mutants Making Significantly Less Biofilm</th>
<th>No. of Mutants</th>
<th>Mutants Making Significantly More Biofilm</th>
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<td>Nucleosome-containing small molecule metabolic process</td>
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If two GOs capture identical genes, then only one GO is mentioned (lowest p-value). http://www.yeastgenome.org/cgi-bin/GO/goTermFinder.pl.

_FLO11_ mRNA than the parental strain (Figure 3B and File S5). As expected, _flo11, flo8, mit1, mss11, rim8, rim101, snf8, tec1, and yps20_ had lower levels of _FLO11_ mRNA. A large number of genes not previously implicated in _FLO11_ regulation were also found to reduce _FLO11_ transcription or mRNA stability when deleted (AID1, ASG1, AVT1, DRN1, ELP4, FMP10, HMT1, KARS, MRPL32, NPC1, NPR1, PEP5, PEX25, RGT1, SPG2, STB6, STP22, VID24, VTC3, YBL029W, YBL029C-A, YFL054C, YGR161W-C, YIL014C-A, YIR024C, YKL151C, YNL200C, YOR034C-A, YOR223W).

Several factors known to regulate _FLO11_ expression did not appear as regulators in the biofilm screen. Most notably, deletion of PKA isoform 2 did not significantly affect biofilm development (Figure 4 and File S1, File S2). The involvement of Tpk2p in _FLO11_ transcription and adhesive invasive phenotypes was previously determined by testing on rich complex medium (Roberson and Fink 1998). We found that on liquid synthetic medium, a _tpk2_ mutant formed 33%–50% of the amount of biofilm of the wild-type strain (Figure 4A). The _Tpk2p_ homolog _Tpk3p_, however, was essential for biofilm development (Figure 4A). The positive effect of _Tpk3p_ on biofilm formation was not at the transcriptional level, because the _tpk3_ mutant had a 2.7-fold increase in _FLO11_ transcript compared with wild-type (Figure 4B and File S5). Hence, _Tpk3p_ appeared to have two roles in _FLO11_ regulation: partial repression of _FLO11_ transcription but induction of _Flo11p_ expression at a posttranscriptional level. To investigate if _Flo11p_ was expressed in the cell wall of the _tpk3_ mutant, we tested the mutant for cell–cell adhesion. The _tpk3_ cells grown in liquid synthetic biofilm medium did not adhere to each other as seen for the wild-type strain (data not shown). Hence, the _tpk3_ mutant did not appear to express functional _Flo11p_.

To identify genes involved in regulation of _FLO11_ by _Tpk3p_ at the posttranscriptional level, we screened a library of _tpk3_ geneX double mutants for ability to suppress the _tpk3_ phenotype and fully restore biofilm formation. Double mutants were made by crossing a biofilm-deficient _tpk3::natMX_ mutant with the mutant collection using a synthetic genetic array (Tong et al. 2001). Diploids were sporulated and haploid double mutants were tested in three independent experiments for ability to form biofilm (Figure S2). The screen identified 35 mutant alleles that suppressed the _tpk3_ biofilm phenotype (File S6). However, none of these genes encoded known components of the PKA pathway or components of translation or RNA processing that suggested a pathway or complex through which _Tpk3p_ acted on _FLO11_.

**Genes essential for biofilm formation are also essential for mat formation on semisolid medium**

Next, we tested biofilm-deficient mutants for other _FLO11_ dependent phenotypes (Figure 5, A–C). Mat formation was tested on complex medium with 0.3% agar. Colonies that formed a structured hub and spokes were considered mats, whereas unstructured, smooth colonies were deemed to have lost the ability to form mats (Figure 5B). Of the 71 mutants that lost the ability to form a biofilm in liquid medium on polystyrene, 69 were also unable to form mats, revealing a nearly complete overlap between genes involved in biofilm in synthetic
medium and mats formation on semisolid complex medium (Figure 5, D and E and Figure S3). Next, invasive growth was tested on complex medium with 2% agar by growing cells in patches for 2 d and washing the resulting colonies with water (Figure 5C). Colonies that left no macroscopic traces of cells after washing were considered noninvasive, whereas colonies that remained on plates after washing were considered invasive (Figure 5C and Figure S4). Of the 69 mutants that did not form mats or biofilm in synthetic medium, 49 also did not show invasive growth (Figure 5, D and E), revealing a core group of 49 genes essential for the haploid phenotypes of mat and biofilm formation in synthetic medium and invasive growth.

**DISCUSSION**

In this work, we found that the *S. cerevisiae* strain S1278b showed a stable, dimorphic growth pattern on polystyrene in liquid synthetic complete medium, with both planktonic and biofilm-forming cells. *FLO11* was essential for formation of the biofilm subpopulation. A possible explanation for the dimorphism is that biofilm subpopulations express *FLO11* under control of Flo8p while planktonic populations repress *FLO11* via Sfl1p. It was previously shown that cells in the S1278b background show variegated *FLO11* expression that is dependent on Flo8p and Sfl1p (Halme et al. 2004). Flo8p and Sfl1p compete for regulation of the *FLO11* promoter and concomitant transcription of either of two ncRNAs in the promoter determines whether the *FLO11* promoter is in a repressed state or a permissive form that allows other transcription factors to induce transcription of *FLO11* (Bumgarner et al. 2009). The Sfl1p switch also appeared to be essential for *FLO11*-dependent dimorphism in biofilms, because only a small subpopulation of cells expressed *FLO11* mRNA in the parental strain, whereas this subpopulation increased to 73% in the sfl1 mutant (Figure 1, F and H).

To investigate the molecular basis for *S. cerevisiae* biofilm formation, we screened a deletion mutant collection in the S1278b strain background for loss of biofilm-forming ability. We identified 71 genes with significantly lower biofilm formation than the reference strain (Figure 2, C and D and File S3). Of these, 38 genes resulted in...
a significant reduction in FLO11 mRNA levels when deleted, suggesting that biofilm was primarily regulated through transcriptional induction of FLO11 (Figure 3). A majority of genes have not previously been associated with FLO11 regulation and the regulatory network underlying FLO11 thus appears to be much more diverse than previously suggested.

Some of the novel regulators are probably directly involved in FLO11 regulation while others might have an indirect effect on FLO11. For example, Npr1p might be an indirect factor. Npr1p is essential for correct targeting of plasma membrane proteins such as the ammonium permease Mep2p and the general amino acid permease Gap1p (De Craene et al. 2001; Vandenbol et al. 1990; Lorenz and Heitman 1998). Both Mep2p and Gap1p are involved in FLO11 expression in some strain backgrounds (Lorenz and Heitman 1998; Torbensen et al. 2012). Therefore, the reduction of biofilm formation and expression of FLO11 in an npr1 mutant is likely to be a consequence of intracellular retention of Mep2p and/or Gap1p. Although the biofilm assay uncovered several FLO11 regulators, a number of genes previously described as essential for FLO11 expression were not found. This was partly the result of a stringent cutoff in the biofilm assay that led to a large number of false negatives. The absence of some known FLO11 regulators in the biofilm assay could also mean that certain gene products are only conditionally essential for FLO11 expression. Hence, some genes might be essential for FLO11-dependent pseudohyphal growth but not FLO11-dependent biofilm development (Ryan et al. 2012). One example is the two PKAs, Tpk2p and Tpk3p. Tpk2p is essential for FLO11 expression in pseudohyphal growth, whereas Tpk3p partially represses FLO11 transcription under these conditions (Robertson and Fink 1998). We found that Tpk2p was not essential for biofilm formation, although detailed analysis of the tpk2 mutant revealed that Tpk2p contributed substantially to the biofilm phenotype (Figure 4A). Tpk3p appears to have two effects on biofilm formation. First, a 2.7-fold increase in FLO11 mRNA in the tpk3 mutant suggested that Tpk3p repressed FLO11 transcription in biofilms (Figure 4B). Second, deletion of TPK3 led to complete biofilm loss (Figure 4A), despite the presence of FLO11 mRNA

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**Figure 5** Most genes essential for biofilm are also essential for invasive growth and surface-spreading biofilm formation. (A) Biofilms in microtiter plates stained with crystal violet as described in Figure 2A legend. Color indicates biofilm. Wt, wild-type. Σ1278b parental strain (MATα can1Δ::STE2p-spHIS5 lyp1Δ::STE3p-LEU2 his3::HisG leu2Δ ura3Δ); flo11Δ, flo11::kanMX in the wild-type background. (B) Mat formation of wild-type and lack of mat by flo11 on semisolid YPD (0.3% agar) after 5 d at room temperature (22°C–25°C). (C) Invasive growth of wild-type and flo11 mutant. Invasive growth was tested after 3 d on solid YPD (2% agar) at 30°C by washing colonies gently with water to remove nonadhering cells. (D) Diagram of deletion mutants that lost the ability to form biofilm and mats and grow invasively. (E) List of phenotypes of mutants with gene deletion that eliminated biofilm formation. Images of mat formation and invasive growth of all mutants are in Figure S3 and Figure S4.
in the *tpk3* mutants, Tpk3p thus must also affect posttranscriptional levels of *FLO11*, as indicated in the model in Figure 4C. The positive role of Tpk3p in biofilm development is thus opposite to its role in pseudohyphal growth. Tpk3p is reported to be essential for redistribution of polysomes on glucose starvation (Ashe et al. 2000) and might also be involved in polysome maintenance of, for example, polysomes with *FLO11* mRNA. This would explain why the *tpk3* mutant did not form a biofilm despite the high levels of *FLO11* mRNA.

Although a number of genes previously described as regulating *FLO11* were not found in our screen, we did find a strong correlation between genes essential for biofilm formation and genes essential for two other *FLO11*-related phenotypes (Figure 5, D and E). Of the 71 mutants that did not form a biofilm, 69 also lost the ability to form mats, even though the growth conditions for the two phenotypes are very different. Mats are formed at room temperature on semisolid complex medium and are characterized by large, flat colonies with cable-like structures and a central hub. In our experiments, biofilms were formed in liquid synthetic medium at 30°C. A smaller subset of 49 genes involved in biofilm and mat formation was also essential for invasive growth on solid complex medium. Although both invasive growth and mat formation are assayed on complex agar medium, the overlap between genes essential for invasive growth and mat formation was smaller than the overlap of genes essential for mat and biofilm formation. This result suggested that a single genetic program mediated the three developmental phenotypes of biofilm and mat formation and invasive growth, whereas at least one other genetic program was specific for mat and biofilm formation (Figure 5, D and E). The common genetic program between the three phenotypes appeared to be almost entirely at the level of *FLO11* transcription. Only a few genes, *ICT1*, *TOM70*, *VOA1*, *YGR266W*, *YLR126C*, and *YOL098C*, did not affect *FLO11* mRNA levels significantly when deleted (Figure 3B).

Several of the genes we identified as involved in biofilm formation were recently reported to be involved in biofilm formation or biofilm-related phenotypes, although their function has not been linked to *FLO11* expression. Ryan et al. (2012) found 655 genes essential for mat formation, whereas 211 genes were found to be essential for structured colony morphology (Voordeckers et al. 2012). Furthermore, Granek et al. (2013) reported that *RGT1* was within a quantitative trait locus found using a biofilm-forming clinical isolate of *S. cerevisiae*. We compared genes essential for biofilm and mat (this study) and genes essential for mat/surface-spreading biofilm identified by Ryan et al. (2012) and found an overlap of 38 genes (File S7). Both studies are based on the use of the same deletion strain collection and the *a priori* assumption was an overlap of 69 genes. Some of the discrepancies in our findings might be ascribed to the method by which mutants were recorded as biofilm and mat formers. Although Ryan et al. (2012) applied colony size as a measure of mat formation, we used colony morphology to determine if mutants formed mats and the adhering proportion of a population to determine biofilm.

**CONCLUSION**

A genome-wide screen of yeast mutants identified that 71 genes were essential for biofilm development. Half of the genes were required for *FLO11* transcription, but only a small subset is previously described as regulators of *FLO11* transcription. These results revealed that the regulation of biofilm formation and *FLO11* expression is far more complex than previously anticipated. The results of this study will be beneficial for our research in yeast biofilm in general for identification of targets for antifungal drugs and new targets for studying biofilms, mats, and pseudohyphal and invasive growth. Biofilm formed by genetically identical eukaryotic cells might be considered as a primitive form of multicellularity. In this context, the current study therefore provides genetic data for understanding of the development programs for primitive multicellularity in a eukaryotic organism.

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**LITERATURE CITED**


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