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Effect of Diethylcarbamazine on HIV Load, CD4%, and CD4/CD8 Ratio in HIV-Infected Adult Tanzanians with or without Lymphatic Filariasis: Randomized Double-Blind and Placebo-Controlled Cross-Over Trial

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Abstract. We assessed the effect of anti-filarial treatment (diethylcarbamazine, DEC) on HIV load, CD4%, and CD4/CD8 ratio in HIV-positive individuals with and without infection with the filarial parasite Wuchereria bancrofti in a randomized, double-blind, placebo-controlled cross-over trial. The study was conducted in Tanga Region, Tanzania, in 2002 and involved 27 adults. A significant decrease in HIV load (54%) and an insignificant increase in CD4% were observed in the HIV-positive individuals with filarial co-infection at 12 weeks after treatment. HIV load and CD4% both increased, although not statistically significantly, in the HIV-positive individuals without filarial infection. The findings suggest that DEC affected HIV load through its effect on the filarial infection rather than through a direct (pharmacodynamic) effect on HIV. Global efforts to control lymphatic filariasis by annual mass treatment with DEC may have a beneficial effect on the HIV/AIDS epidemic in areas where HIV and lymphatic filariasis co-exist.

INTRODUCTION
The high prevalence of HIV and parasite infections in sub-Saharan Africa results in large geographical overlaps and thereby potential for interaction between these two types of infections in co-infected individuals. Parasites, helminths in particular, may increase susceptibility to HIV and contribute to a more rapid progression toward AIDS.1–3 Helminth-induced immune activation may be an underlying mechanism, as helminth infections are associated with increased viral replication.4 Th1–Th2 cross-regulation is likely to play a role in this interaction. Helminth infections are commonly associated with an upregulated Th2 response. This seems to prevent production of Th1-related cytokines and cytotoxic T-cell activity,1,4 which play a central role in the immunologic combat against HIV. HIV virus may furthermore replicate more rapidly in Th2 cells than in Th1 cells,5 and a significant level of replication seems to occur in eosinophils in individuals with helminth-induced eosinophilia.6 Treatment of helminth infections is thus likely to be particularly beneficial to HIV patients and has been found to be associated with a significant decrease in HIV plasma load.7 However, it has also been observed that helminth infections were not associated with faster progression of HIV disease, and it was suggested that anthelminthic therapy may not slow down HIV progression in co-infected individuals.8 Apart from helminth infections, some protozoan infections interact with HIV. Thus, malaria has been shown to be an important factor in promoting the spread of HIV in sub-Saharan Africa.9

We previously examined the cross-sectional relationship between subclinical HIV, lymphatic filariasis (Wuchereria bancrofti infection), and other parasitoses (Plasmodium falciparum, Ascasis lumbricoïdes, Trichurus trichiura, and hookworm infections) among 858 adults in Tanga Region, northeastern Tanzania.10 A significant positive association was observed between HIV and W. bancrofti infection and between HIV and P. falciparum infection. By contrast, HIV and hookworm infection were negatively associated. A more detailed immunologic study carried out in a subpopulation of 59 individuals, however, could not provide clear evidence for an interaction between HIV and W. bancrofti infection.11

In the present study, we looked further into the association between HIV and W. bancrofti infection, by analyzing the effect of diethylcarbamazine (DEC) treatment of W. bancrofti on HIV infection in HIV-positive individuals with W. bancrofti co-infection and compared this with the effect in HIV-positive individuals without filarial co-infection.

MATERIALS AND METHODS

Study population and study design. The study was conducted to assess and compare the effect of DEC on viral load, CD4%, and CD4/CD8 ratio in HIV-positive individuals with or without W. bancrofti co-infection in a randomized, double-blind, placebo-controlled cross-over trial. The study was carried out in May–November 2002 in 5 villages (Mwahako, Mwakidila, Pongwe South, Kakindu, and Jitengeni) in Tanga Region, northeastern Tanzania. As part of a cross-sectional study on the association between HIV, lymphatic filariasis, and other parasitic infections, 858 adults (18–70 years) were tested for HIV (antibodies), lymphatic filariasis (W. bancrofti circulating filarial antigens, CFA), malaria parasites, and intestinal helminth eggs.10 Pre- and post-test counseling was provided by the Tanga AIDS Working Group, a local NGO. The 858 individuals constituted the source of study individuals for further detailed investigations on HIV and filariasis interactions.

Thirty-four individuals were willing to participate in the current study and met the inclusion criteria of being 1) HIV-positive and 2) without obvious clinical manifestations of HIV and/or W. bancrofti infection. The study individuals were randomly assigned to receive DEC or placebo at the beginning of the study, and the opposite treatment was given 12 weeks later (Figure 1). Venous blood was collected at baseline and at 1, 12, 13, and 24 weeks after the first round of treatment. Seven of the 34 participants did not complete all follow-up
examinations and were therefore excluded from the analyses. There was no difference in age, sex, or infections between those lost to follow-up and those who remained in the study. Data from 27 individuals are analyzed. We tested the hypothesis that DEC treatment would 1) reduce the viral load, 2) increase the CD4%, 3) increase the CD4/CD8 ratio, and 4) affect HIV-positive individuals with or without *W. bancrofti* co-infection differently. Research clearance to conduct the study was obtained from the Medical Research Coordinating Committee of the National Institute for Medical Research in Tanzania, and the study was approved by the Danish National Committee on Biomedical Research Ethics.

**Treatment with diethylcarbamazine (DEC).** The 34 study participants were treated with DEC either at the beginning of the study or after 12 weeks. The drug was administered as a single dose of 6 mg/kg body weight, which is the yearly dose given for transmission control of lymphatic filariasis. The tablets were prepared specifically for this study by a manufacturer (Almega, Ringsted, Denmark); 50-mg DEC tablets and corresponding placebo tablets were supplied. The identical DEC and placebo tablets were packed in containers with different color codes (“red” or “blue”). Individuals were randomized (1:1) to receive treatment in the order of “red” followed by “blue” or “blue” followed by “red” by using a list of random numbers. The selected study participants were listed and numbered from 1 to 34, and the first 17 numbers (between 1 and 34) encountered in the table (when starting on a randomly chosen figure) were assigned to receive treatment in the order red–blue, and the remaining 17 received treatment in the order blue–red. All study personnel and participants were blinded to treatment assignment throughout the study.

**Venous blood samples.** At each sampling time, 25 mL of venous blood (20 mL heparinized and 5 mL non-heparinized) was collected from each study participant. The blood was transferred in a cool box to the field laboratory. The 5 mL of nonheparinized blood was left overnight at 4°C to clot. The serum was collected the following day and kept at −80°C until used for quantitative determination of HIV load and CFA intensity. The 20 mL of heparinized blood was used for isolation of peripheral blood mononuclear cells (PBMCs) and for cell culturing, separation of plasma, hemoglobin measurements, and preparation of blood slides for further immunologic studies.

**Circulating filarial antigen (CFA).** The *W. bancrofti* CFA status was determined in the field by use of the NOW immunochromatographic test (ICT) for filariasis (Binax, Scarborough, ME), a rapid, qualitative immunodiagnostic test, as de-
The intensity of CFA was later quantified in serum using the TropBio ELISA test kit (TropBio Pty. Ltd., Townsville, Australia). Manufacturers’ procedures were followed, and all tests were carried out in duplicate. Serum samples with a mean optical density (OD) value > standard 2 (32 antigen units) were considered positive. Most positive samples had a mean OD value ≥ that of the highest standard (standard 7 = 32,000 antigen units). These were subsequently diluted 1:10 and retested. Samples with a mean OD value ≥ that of the highest standard after dilution (N = 11) were assigned a value of 320,000 antigen units (corresponding to 32,000 antigen units multiplied by the dilution factor 10).

Assessment of HIV infection. HIV status was determined in the field by 2 different rapid tests (both detecting antibodies to HIV 1 and 2), as described previously. The HIV load in samples from individuals tested positive in both rapid tests (discrepancies did not occur) was quantified using a modification of an in-house developed reverse-transcriptase PCR. HIV RNA was isolated from 125 μL of serum. All samples were analyzed in 2 independent extractions and runs. Samples with discordant results (discrepancy > 0.5 log) were retested in duplicate. The limit of detection was 40 genome equivalents (geq)/mL.

Staining and counting of CD4 and CD8 T cells. An immunoperoxidase (IA) phosphatase method was used for staining and counting of CD4 and CD8 T cells. In brief, 2 thin smears from each patient underwent immunocytochemical labeling with monoclonal antibodies against CD4 and CD8 receptors, respectively. The stained sections (by a secondary antibody conjugated to biotin and an avidin–bixin–tritiated analog conjugated to alkaline phosphatase, both from DakoCytomation, Carpinteria, CA) antigen/antibody binding was visualized as a reddish-stained ring around the lymphocyte nucleus by adding a substrate containing New Fuchsian (fuchsin + substrate–chromogen system, DakoCytomation). Stained CD4 and CD8 cells were counted among 200 lymphocytes, and the CD4% and the CD4/CD8 ratio were estimated. The procedure followed the instructions given by a WHO laboratory manual for characterization of T-cell subsets in peripheral blood.

Examination for malaria and intestinal helminths. Examinations for malaria parasites and intestinal helminth eggs were carried out as described previously.

Statistical analysis. Viral loads and infection intensities were normalized by log_{10} transformation. The two sample Students t test was used to analyze differences in means, and the χ² test for differences in prevalences between groups. The effect of DEC treatment on HIV load, CD4%, and CD4/CD8 ratio was analyzed according to standard procedures for cross-over trials as described by Altman. Welch’s version of the t test was used to test for treatment effect, period effect, and treatment period interaction. No period effect and no treatment period interaction were observed. A linear model was fitted with the period differences as response and treatment, infection status, and their interaction as predictors; a significant interaction term means that the treatment effect differs between the HIV⁺ and HIV⁺/CFA⁻ groups.

RESULTS

Infection status-eligible individuals were recruited from May to June 2002. Collection of baseline data, randomization and the first round of treatment occurred in June 2002. The study participants were followed up at 1, 12, 13, and 24 weeks after the first round of treatment. Cross-over was at 12 weeks after the first treatment. We now describe the baseline characteristics of the HIV-positive study participants (with or without W. bancrofti co-infection) and the effect of DEC on HIV load, CD4%, and CD4/CD8 ratio in these patients during a 12-week period only, as there was no measurable effect of DEC after 1 week. There were no adverse events or side effects related to treatment among the study participants.

Characterization of the study groups. Among the 27 HIV-positive study participants, 10 (37%) were males (Table 1). The mean age was 41 (range 22–70) years, and there was no significant difference in age between males and females. The mean HIV load was 3.71 log units, and the mean CD4% was 27.3. CFA was present in 12 (44%) individuals, and their mean intensity was 33,287 antigen units. P. falciparum was found in 5 individuals (19%) with a geometric mean intensity of 134 parasite/μL blood among those infected. Hookworm eggs were found in 13 (62%) of 21 examined individuals (stool samples were not obtained from 6 individuals) with a geometric mean intensity of 410 eggs per gram (epg) of feces among those infected. Eggs from A. lumbricoides and Schistosoma mansoni were not found in any of the study participants, and T. trichiura eggs were found in 1 only. There was no significant sex difference in the prevalence and intensity of CFA, P. falciparum, or hookworm infection. When the 27 study participants were divided into two groups, with and without filarial co-infection, the prevalence of hookworm was 63.3% and 60.0% (P = 0.86), respectively, and the prevalence of P. falciparum was 27.3% and 13.3% (P = 0.37), respectively.

Effect of DEC treatment on HIV load. The individual HIV load levels at 12 and 24 weeks follow-up in individuals with (CFA⁺) or without (CFA⁻) filarial co-infection, are presented in Figure 2. The effect of DEC on viral load was first assessed in all HIV-positive individuals, disregarding filarial status (CFA⁺/CFA⁻, N = 26), and subsequently in individuals with (CFA⁺, N = 11) or without (CFA⁻, N = 15) filarial co-infection, separately. One sample could not be amplified in

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Background characteristics of the 27 HIV-positive study individuals</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, in years*</td>
<td>41 (± 13)</td>
</tr>
<tr>
<td>Males (%)</td>
<td>37</td>
</tr>
<tr>
<td>HIV load (geq/mL)†</td>
<td>3.71 (± 0.97)</td>
</tr>
<tr>
<td>CD4%*</td>
<td>27.3 (± 11.4)</td>
</tr>
<tr>
<td>Circulating filarial antigen (CFA)</td>
<td></td>
</tr>
<tr>
<td>Positives, in %</td>
<td>44</td>
</tr>
<tr>
<td>Intensity, in CFA units*</td>
<td>33,287 (± 87,519)</td>
</tr>
<tr>
<td>P. falciparum</td>
<td></td>
</tr>
<tr>
<td>Positives, in %</td>
<td>19</td>
</tr>
<tr>
<td>Intensity, in parasites/μL‡</td>
<td>134 (± 5)</td>
</tr>
<tr>
<td>Hookworm</td>
<td></td>
</tr>
<tr>
<td>Positives, in %§</td>
<td>62</td>
</tr>
<tr>
<td>Intensity, in epg¶</td>
<td>410 (± 5)</td>
</tr>
</tbody>
</table>

* Mean (± standard deviation).
† Mean intensity (± standard deviation) expressed as log_{10} viral load (genome equivalents [geq] per mL) in serum from 26 individuals (HIV RNA could not be amplified in one sample).
‡ Intensity expressed as geometric mean (5 is the standard deviation which should be interpreted as a factor: intensities between 26.8 and 670) malaria parasites/μL of infected individuals only (n = 3).
§ n = 21, as stool samples were not obtained from 6 individuals.
¶ Intensity expressed as geometric mean (5 is the standard deviation which should be interpreted as a factor: intensities between 82 and 2,050) eggs per gram of feces in infected individuals only (n = 13).
PCR but tested positive in 2 different HIV field tests and was omitted from the HIV load analyses.

The overall analysis among the HIV-positive individuals (CFA+/CFA−) showed no significant treatment effect on HIV load (0.059 log unit increase = 15% [95% confidence interval [CI]: −0.22; 0.34], P = 0.66). However, when the results were stratified by filarial status, an effect was observed in both groups (Figure 3A). In the CFA+ group, a significant reduction in HIV load (−0.34 log unit [95% CI: −0.637; −0.048], P = 0.03), equaling 54%, was observed. In contrast, a 0.36 log unit increase in HIV load (0.36 log unit [95% CI: −0.10; 0.82], P = 0.11), which equals a doubling (2.3 times the pretreatment level), was observed in the CFA− group, but the increase was not statistically significant. When the effect of DEC on HIV load was compared in the CFA+ and CFA− groups, it was shown to be significantly different (P = 0.007).

Effect of DEC treatment on the CD4% and CD4/CD8 ratio. The individual change in CD4% in the period between the 12- and 24-week follow-up varied considerably (with both increases and decreases observed), and given that day-to-day variation may also occur, these data should be interpreted with caution. The changes in CD4/CD8 ratio in the same period were very small. Because of these facts, individual changes in CD4% and CD4/CD8 ratio are not presented graphically. Instead, the estimated mean effect of DEC on CD4% and CD4/CD8 ratio is presented in Figure 3, B and C, respectively. As with HIV load, the effect was first assessed in all HIV-positive individuals (CFA+/CFA−, N = 27) and then in those with (CFA+, N = 12) or without (CFA−, N = 15) filarial co-infection, separately. The overall analysis among the HIV-positive individuals (CFA+/CFA−) showed no significant change in CD4% (mean increase of 1.20% point [95% CI: −2.24; 4.63], P = 0.48) nor in the CD4/CD8 ratio after treatment (mean increase of 0.08% point [95% CI: −0.09; 0.25], P = 0.36).

In the CFA+ group, the 12-week treatment effect on the CD4% was an increase of 2.5% points (95% CI: −4.90; 9.94, P = 0.45), but the increase was not statistically significant (Figure 3B). In the CFA− group, an insignificant effect (increase of 0.12% point [95% CI: −3.41; 3.64], P = 0.95) was likewise observed. In contrast to HIV load, there was no significant difference in the treatment effect in the two groups (P = 0.48).

The effect of DEC treatment on the mean CD4/CD8 ratio was, as for the CD4%, small and statistically insignificant in both the CFA+ and CFA− group with a mean increase in
CD4/CD8 ratio of 0.15% point (95% CI: −0.24; 0.54, \( P = 0.38 \)) and 0.01% point (95% CI: −0.10; 0.1, \( P = 0.84 \)), respectively (Figure 3C). The treatment effect in the 2 groups was not statistically different (\( P = 0.37 \)).

**DISCUSSION**

The present study analyzed the effect of DEC treatment on HIV load, CD4%, and CD4/CD8 ratio over a 12-week follow-up period in HIV-positive individuals with or without filarial co-infection. Only individuals with asymptomatic infections were included in the study to avoid confounding effects related to immunologic changes in the late stages of HIV infection and in chronic manifestations of filariasis. The pretreatment survey showed a high prevalence of *P. falciparum* malaria and hookworm infection in the study individuals.

The finding of a significant reduction in HIV load in the HIV-positive individuals with filarial co-infection after treatment indicates that DEC treatment reduces viral load in HIV and filariae-positive individuals and may thus slow down the progression of HIV. It seems likely that DEC acts through its effect on the filarial infection. The mechanism behind this is probably immunologically mediated, as treatment of the filarial co-infection may result in a switch from a predominant Th2-type response (known to be associated with filarial infections and infections with other helminths) to an enhanced Th1-type response favorable in the combat against HIV infection. Moreover, the reduction in viral load may be related to a reduced level of immune activation, as antifilarial treatment may cause a reduction in activated T cells and thereby reduce susceptibility to HIV and delay the viral replication. The contrasting finding of an increase, though not statistically significant, in HIV load among HIV-positive individuals without filarial infection may lead to the thought that DEC could be harmful in terms of increasing HIV load. However, the trend of an increase in CD4%, rather than a decrease, in the same group of individuals does not lend support to this suspicion.

The difference in the effect of DEC on HIV load in individuals with and without filarial co-infection supports the idea that DEC acts through its antifilaricidal properties rather than through a direct (pharmacodynamic) effect on HIV load in the dose applied in this study. However, it cannot be excluded that administration of DEC in a higher dose and/or for a longer period of time could also have a direct effect on HIV infection. Previous studies in cats have indicated that DEC may have a direct effect on retroviral infections in cats, and DEC treatment (5 mg/kg body weight administered for 30 days) of HIV-infected humans in Guatemala intriguingly showed that 5 out of 7 individuals, who tested positive for HIV 1 p24 antigen before treatment, were negative (4 persons) or had a lower (1 person) serum p24 value after 4 weeks of treatment.

The largest increase in CD4% after treatment was observed in the CFA+ group, which corresponds well with the significant decrease in HIV load in this group. The trend of an increase in CD4%, rather than a decrease, in the HIV-positive individuals without filarial infection indicates that DEC had no harmful effect on the health of these patients. It could be speculated that the time required for the treatment effect to be reflected in the CD4% is longer than the follow-up period, and the increase in CD4% after treatment might have been bigger had the patients been followed up for a longer time period. In addition, a higher dose of DEC may have resulted in a more distinct increase in CD4%.

Generally, the change in the CD4/CD8 ratio after treatment was small. A more pronounced increase in this ratio might have been expected in the group of HIV-positive individuals with filarial co-infection after antifilarial treatment as a result of the increased CD4%. However, because production and activity of cytotoxic T cells (CD8 cells) appear to be associated with a Th1-type response, an increase in the CD4/
CD8 ratio may have failed to appear because the increase in CD4 cells was accompanied by a simultaneous increase in production of CD8 cells as a result of a switch from a Th2- to a Th1-predominated response after treatment of the filariae.

In summary, the presented data suggest that DEC treatment may have an important positive impact on the host’s ability to respond to the HIV infection in individuals co-infected with filariae and may thereby slow down the progression of HIV. The ongoing global efforts to control lymphatic filariasis, which in many areas rely on annual mass treatment with DEC, may thus have an additional beneficial effect in terms of contributing to alleviation of the HIV/AIDS epidemic in areas where HIV and lymphatic filariasis co-exist. A positive effect of DEC on HIV in W. bancrofti-endemic areas may be even more pronounced if the drug was administered at a higher dose and/or more frequently than once a year, a perspective that deserves to be further investigated.

The prospects of DEC, being an inexpensive and well-tolerated drug, as a tool in the combat against HIV/AIDS may be even wider. If studies similar to the present study but with 1) a higher dose of DEC, 2) continuous or regular administration (contrary to the single dose in the present study), 3) larger study populations, and 4) longer follow-up periods could provide indications of an anti-retroviral effect of DEC, these might form the basis for development of new DEC-based treatment strategies to combat HIV/AIDS. Studies on the anti-retroviral efficacy of DEC (or DEC derivatives) would also be of importance in attempts to be at the forefront of potential development of resistance to the currently used preparations.

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