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Rhizosecretion of stele-synthesized glucosinolates and their catabolites requires GTR-mediated import in Arabidopsis

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Abstract

Casparian strip-generated apoplastic barriers not only control the radial flow of both water and ions but may also constitute a hindrance for the rhizosecretion of stele-synthesized phytochemicals. Here, we establish root-synthesized glucosinolates (GLS) in Arabidopsis as a model to study the transport routes of plant-derived metabolites from the site of synthesis to the rhizosphere. Analysing the expression of GLS synthetic genes in the root indicate that the stele is the major site for the synthesis of aliphatic GLS, whereas indole GLS can be synthesized in both the stele and the cortex. Sampling root exudates from the wild type and the double mutant of the GLS importers GTR1 and GTR2 show that GTR-mediated retention of stele-synthesized GLS is a prerequisite for the exudation of both intact GLS and their catabolites into the rhizosphere. The expression of the GTRs inside the stele, combined with the previous observation that GLS are exported from biosynthetic cells, suggest three possible routes of stele-synthesized aliphatic GLS after their synthesis: (i) GTR-dependent import to cells symplastically connected to the cortical cells and the rhizosphere; (ii) GTR-independent transport via the xylem to the shoot; and (iii) GTR-dependent import to GLS-degrading myrosin cells at the cortex. The study suggests a previously undiscovered role of the import process in the rhizosecretion of root-synthesized phytochemicals.

Key words: Arabidopsis, biosynthesis, exudate, glucosinolates (GLS), glucosinolates catabolites, GTR, import, rhizosphere, rhizosecretion, stele-synthesized phytochemicals, trans-cellular transport.

Introduction

The rhizosphere, the zone closely surrounding the roots where the exchange of micronutrients and organic matter occurs, is pivotal for both symbiotic and pathogenic interactions between the plant and soil organisms. There is an increased awareness of the importance of root exudation for the establishment of the microbial community in the rhizosphere (Bais et al., 2006; Badri et al., 2009b; Baetz and Martinoia, 2014; Carvalhais et al., 2015) and in the response to environmental...
signals (Hoehenwarter et al., 2016; Ziegler et al., 2016). Depending on the species, the rhizosphere is apoplastically isolated from the rest of the root by the endodermis and/or exodermis with their Casparian strip-impregnated cells. Compounds synthesized outside or within the endodermis and/or exodermis can be released to the rhizosphere by passive diffusion or through specific exporters (Badri et al., 2009a; Weston et al., 2012). Compounds that are synthesized and released into the apoplast behind the apoplastic barriers (endodermis or exodermis) can follow one of two transport routes from the inside to the outside of the endodermal/exodermal cells. The first route is mediated by plasmodesmata (PD) connecting biosynthetic cells symplastically to the endodermis from which they are released by exporters to the rhizosphere. The second route is trans-cellular and involves the repeated efflux and influx through consecutively arranged and polarized carrier proteins (Andersen et al., 2015). Even though several transporters have been identified for releasing compounds from root cells, these are all involved in the efflux of metabolites from cells outside the Casparian strip (Sugiyama et al., 2007; Badri et al., 2009a; Fourcroy et al., 2014; Sasse et al., 2015). By contrast, little is known about the transport route to the rhizosphere of specialized metabolites synthesized within the stele.

Arabidopsis thaliana (hereafter Arabidopsis) with glucosinolates (GLS) as major defence compounds has become an ideal model system to study the mechanism of transport of specialized metabolites because of its powerful genetic toolbox (Jorgensen et al., 2015). Arabidopsis produces tryptophan-derived indole GLS and methionine-derived aliphatic GLS which are further divided into short-chained (with three to five methylene groups, C3–C5) and long-chained (with six to eight methylene groups, C6–C8) aliphatic GLS (Grubb and Abel, 2006), of which the latter are primarily synthesized and stored in the roots (Andersen et al., 2013). Previous studies showed that root cells synthesizing long-chained aliphatic GLS by default export these GLS to the apoplast from where the two plasma-membrane localized glucosinolate importers, GTR1 and GTR2 (Nour-Eldin et al., 2012), are suggested to import them into local storage cells (Andersen et al., 2013). Indeed, gtr1 gtr2 roots contain drastically reduced levels of long-chained, aliphatic GLS whereas rosettes over-accumulate them. This over-accumulation is caused by translocation of the exported GLS via the xylem (Andersen et al., 2013; Madsen et al., 2014). However, the total levels of GLS in gtr1 gtr2 roots and rosettes are higher compared with wild-type levels. This shows that, in the absence of GTR1 and GTR2 which are both expressed in the root, higher amounts of GLS remain within the plant. It is conceivable that these higher amounts of GLS accumulating in the gtr1 gtr2 mutant were destined for rhizosecretion. This hypothesis proposes that GTR-mediated GLS import plays an important role in the exudation pathway of stele-synthesized GLS. Noticeably, the tissue-specific localization of GLS biosynthesis has not previously been determined in Arabidopsis roots using non-invasive approaches. In addition, although exudation of the GSLs and/or their catabolites has been observed (Schreiner et al., 2011) and suggested in Brassica crops (Choesin and Boerner, 1991; McCully et al., 2008), intact GLS have not previously been identified in the root exudates of the model plant Arabidopsis, whereas biosynthetic intermediates and catabolites of GLS have been detected in different metabolomics studies (Badri et al., 2013; Strehmel et al., 2014, 2016; Monchgesang et al., 2016; Ziegler et al., 2016).

It is shown here that long-chained, aliphatic GLS are synthesized in the stele of Arabidopsis roots and exuded into the rhizosphere. A new sampling method has enabled the detection of both intact GLS and their catabolites in Arabidopsis root exudates. Therefore, we propose that elucidating the transport pathway of long-chained, aliphatic GLS provides a model system for understanding the root exudation of stele-synthesized compounds in general. As a first step towards investigating the transport route of GLS from the site of synthesis to the rhizosphere, we compared root exudates of gtr1 gtr2 with the wild type and determined the localization of GTR at the cellular level. Our data indicate that GTR1 and GTR2 import into the symplasm is a prerequisite for the translocation of stele-synthesized GLS across the apoplastic barrier of the endodermis into the rhizosphere.

Materials and methods

Plant growth and sampling root exudates

A. thaliana Col-0 and mutants with a T-DNA insertion in both GTR1 (At3g47960) and GTR2 (At5g62680; gtr1 gtr2 dKO) were grown on a non-sterile standardized plant growth substrate (Fruhstorfer Erde type P, Germany) with a pH value of 6.0 in a climate chamber under short-day conditions (8/16h light/dark, 22 °C, 40–60% humidity). After 2 weeks, single plants were transferred to sand-filled pots and watered with nutrient solution as described by Gibeaut et al. (1997). After an additional 4 weeks, the sand was carefully removed from the roots and the plants were transferred to tubes filled with distilled water. After 1 h, plants were transferred to fresh tubes (two plants per tube) filled with bi-distilled water and exudates were collected for 4h. In order to determine whether enzymatic hydrolysis of exuded GLS takes place, 100 µl of 0.1 mM 2-propenyl GLS (Carl Roth GmbH, Germany) was added to each tube. During the whole procedure, plants were kept under the cultivation conditions. Exudates were filtered through a mixed cellulose ester membrane filter of pore size 0.22 µm (Carl Roth GmbH, Germany) to remove cellular debris and external microorganisms. Exudates from approximately 20 plants were pooled to give one sample and three samples were collected for each harvest. After collection of exudates, the roots were harvested and the fresh weight was determined to relate the amount of exudates to root biomass.

Construct cloning

Derivatives of the USER™ compatible pCambia1300U and pCambia2300U plant expression vectors (Nour-Eldin et al., 2006) were used in this work. The PCR product was then introduced into the PaCl and Nt.BbvC1 digested pCambia2300U vector by USER™ cloning. The mVenus variant of the yellow fluorescent protein was amplified with the same primer pair and introduced into the opened pCambia1300U vector.

The promoter sequences including upstream genomic sequences and the 5’ UTR of the CYP83A1, CYP83B1 genes (1033 bp and 1001 bp, upstream of the start codon, respectively) were amplified from genomic DNA of Arabidopsis thaliana ecotype Columbia-0. The corresponding coding sequences were amplified from Arabidopsis thaliana ecotype Columbia-0 cDNA. The promoter
and coding sequences were seamlessly fused by USER™ fusion (Geu-Flores et al., 2007) and, at the same time, inserted into the USER™ cassette of the opened pCambia1300-mVenus (CYP83A1, CYP83B1) vectors. The proper fusion and insertion of sequences was confirmed by DNA sequencing.

**Cellular localization**

Localization experiments were done on 12–14-d-old plants using a Leica SP5-X confocal microscope. Fluorophores were excited at the respective absorption maxima (514 nm for YFP and 546 nm for mOrange). Emission was collected at 525–540 nm (YFP), and 560–580 nm (mOrange).

**GUS assay and histology**

Detection of GUS expression was performed with 5-bromo-4-chloro-3-indoly-β-D-glucuronidase (X-Glu). Tissues were vacu-umed in 50 mM sodium phosphate buffer, pH 7.0 (1% Triton X-100/1% DMSO/10 mM EDTA/0.5 mM potassium ferrocyanide/0.5 mM potassium ferrocyanide/1 mM X-Glu) and then incubated at 37 °C. The stained samples were washed twice with 70% ethanol, fixed in 4% formaldehyde, and cleared with 70% ethanol. Tissues were mounted in 50% glycerol, and microscopy was con-duted. Histological analysis followed by sections was performed as described with some modifications. Briefly, tissues were gen-tly fixed in 90% acetone, washed with water, incubated in 50 mM sodium phosphate buffer, pH 7.0 (0.2% Triton X-100/10 mM EDTA/10 mM potassium ferrocyanide/10 mM potassium ferrocyanide/1 mM X-Glu), at 37 °C. The stained tissues were fixed in 4% paraformaldehyde, dehydrated with acetone then embedded in Spurr Resin. Resin sections (5 μm) were microtomed (Leica) and were observed after staining.

**GLS analysis**

Desulfo-GLS profiles and concentrations of lyophilized plant tissue were determined as described before (by Witzel et al. (2015) with slight modifications: For the determination of GLS in exudates, the total exudate volume (500 nl) was first subjected to catabolites analy-sis (see below). Then the exudate was spiked with 20 nmol of the internal standard (IST) 4-hydroxybenzyl GLS (Carl Roth GmbH, Karlsruhe, Germany) and loaded onto a 1 ml DEAE-Sephadex A-25 ion-exchanger column (Sigma-Aldrich Chemie GmbH, Steinheim, Germany), further treated and analysed with the UHPLC-system as reported previously (Witzel et al., 2015).

**Analysis of GLS breakdown products**

For the determination of enzymatically formed breakdown prod-ucts of the GLS in fresh plant tissues, benzonitrile (≥99.9%, Sigma-Aldrich Chemie GmbH) was added as an IST and catabo-lites were extracted with dichloromethane (Carl Roth GmbH; GC Ultra Grade) as described previously (Witzel et al., 2015). For the analysis of glucosinolate breakdown products in exudates, the total volume of each exudate (500 nl) was extracted with 50 ml of dichloromethane in a separating funnel. The separated dichloromethane layer was dried using anhydrous NaSO₄ (VWR International GmbH, Darmstadt, Germany; ≥99%) and the aqueous phase was re-extracted twice using 25 ml of dichloromethane. The combined dichloromethane extract was dried under nitrogen to 300 μl and sub-jected to GC-MS analysis as described above.

**Statistical data analysis**

To represent the changes in GLS levels, two-tailed Student’s t tests and one-way ANOVA followed by Tukey HSD Calculator multi-ple comparison post-hoc analysis (P <0.05) were performed using Origin Pro 2015.

**In silico analysis**

Cell-type-specific expression was derived from microarray studies of RNA bound to ribosomes which were immunoprecipitated by use of epitope-tagged ribosomal protein from seedlings (Mustroph et al., 2009; http://efp.ucr.edu/).

**Accession numbers**

Sequence data from this article can be found in the Arabidopsis Genome Initiative or EMBL/GenBank data libraries under accession numbers: AT3g47960 (GTR1); AT5g62680 (GTR2); AT4G13770 (CYP83A1); AT4G31500 (CYP83B1).

**Results**

**Sampling of Arabidopsis root exudate**

Towards our goal of understanding the root exudation process using Arabidopsis and GLS as a model system, we developed a method to measure GLS and their catabolites in Arabidopsis root exudates under physiological conditions (see Supplementary Fig. S1A–C at JXB online). Based on previous reports, we optimized several steps in the procedure. First, we optimized root growth conditions by growing plants in sand instead of using a hydroponics system (Badri et al., 2013; Strehmel et al., 2014). Secondly, the exudation was car-ried out in de-ionized water instead of growth media. Thirdly, dichloromethane (CH₂Cl₂) was used to denature the myrosi-nase (Sah, 1999) while partitioning intact GLS to the aqueous phase and catabolites to the organic phase. Finally, we applied DEAE-Sephadex anion exchange chromatography to enrich GLS, followed by targeted HPLC and LC-MS detection methods to measure intact GLS (Brown et al., 2003; Mikkelsen et al., 2003).

We first tested the recovery efficiency of GLS in exudates by adding exogenous 2-propenyl GLS standard (a GLS not found in Arabidopsis Col-0) to the medium prior to sampling. We were able to recover 37 ± 8% of the added 2-propenyl GLS while another 8 ± 3% was recovered as the corresponding catabo-lite product allyl-ITC (AITC) (Supplementary Fig. S1D). As 2-propenyl GLS can be taken up and translocated from the root to the shoot (Francisco et al., 2016), 2-propenyl GLS was analysed in the whole plant. No 2-propenyl GLS or AITC in either shoot or root tissue was detected (Supplementary Tables S1, S2). We concluded that, in the current sampling method, approximately 50% GLS was recovered from the exudate, of which approximately 75% GLS remained intact and approximately 25% was degraded during sample preparation.

**GLS and their catabolites in root exudates of the wild type**

Implementing our method, we set out to isolate both intact GLS and their catabolites in root exudates from Arabidopsis Col-0. After 4 h, the total amount of collected GLS and their catabolites per gram of root tissue represented approximately 1% of GLS per gram of root tissue (Fig. 1A). The ratio of GLS to their catabolites was 1:4. In root exudates from the wild type, all three classes of GLS, i.e. short-chained and long-chained aliphatic GLS as well as indole GLS, were present.
(Fig. 1B; Table 1). The profile of aliphatic GLS in the root exudate was similar to the endogenous root profile, whereas the profile of indole GLS differed between the root and the exudate (Fig. 1B, D; Table 1). For several of the GLS catabolites, the profile in root exudates differed from the profile of the GLS in root exudates (Fig. 1B, C; Table 1) and from the catabolite profile of homogenized root tissue (Supplementary Table S2). The difference in profile of GLS and their catabolites between root and exudate (Fig. 1; Table 1) suggests that hydrolysis does not occur during sampling and extraction.

![Fig. 1. Profiling of GLS and their catabolites in Arabidopsis Col-0 root exudates.](image)

**Table 1.** Comparison of the profiles of GLS and their catabolites in Arabidopsis Col-0 roots and root exudates.

Quantities are derived from the mean of at least two biological repeats each with three batches of plants (each $n=20$). GLS profiles of roots and root exudates, as well as the catabolite profile in root exudates were determined. To enable comparison between the amount of GLS and catabolites, the catabolites were converted into their corresponding GLS precursors. The values were determined by one-way ANOVA followed by Tukey HSD Calculator multiple comparison post-hoc analysis ($P<0.05$). The significance of differences between the profiles of roots and exudates in the respective comparisons are indicated by letters. Errors denote standard deviation. 4MTB, 4-(methylthio)butyl; 7MTH, 7-(methylthio)heptyl; 8MTO, 8-(methylthio)octyl; 3MSP, 3-(methylsulfinyl)propyl; 4MSB, 4-(methylsulfinyl)butyl; 7MSH, 7-(methylsulfinyl)heptyl; 8MSO, 8-(methylsulfinyl)octyl; 13M, 3-indolylmethyl; 1MOI3M, 1-methoxy-3-indolylmethyl; 4MOI3M, 4-methoxy-3-indolylmethyl glucosinolate. n.d. Not detected.

<table>
<thead>
<tr>
<th>GLS precursors of catabolites (%)</th>
<th>Root</th>
<th>Exudate</th>
<th>Exudate</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>4MTB</strong></td>
<td>3.9±1.5 a</td>
<td>11.1±8.8 a</td>
<td>27.6±9.2 b</td>
</tr>
<tr>
<td><strong>4MSB</strong></td>
<td>19.4±4.6 a</td>
<td>13.1±16.3 a</td>
<td>33.1±17.1 a</td>
</tr>
<tr>
<td><strong>7MTH</strong></td>
<td>3.4±1.7 a</td>
<td>0.7±0.7 a</td>
<td>0.3±0.4 a</td>
</tr>
<tr>
<td><strong>7MSH</strong></td>
<td>1.9±1.6 a</td>
<td>5.4±2.9 b</td>
<td>n.d.</td>
</tr>
<tr>
<td><strong>8MTO</strong></td>
<td>19.9±8.9 a</td>
<td>12.9±11.5 a</td>
<td>6.4±8.4 b</td>
</tr>
<tr>
<td><strong>8MSO</strong></td>
<td>17.9±5.0 a</td>
<td>28.3±17.2 a</td>
<td>18.9±30.6 a</td>
</tr>
<tr>
<td><strong>13M</strong></td>
<td>7.0±0.9 a</td>
<td>13.3±5.4 a</td>
<td>10.2±9.6 a</td>
</tr>
<tr>
<td><strong>1MOI3M</strong></td>
<td>21.9±7.7 a</td>
<td>7.5±6.3 b</td>
<td>n.d.</td>
</tr>
<tr>
<td><strong>4MOI3M</strong></td>
<td>4.7±2.2 a</td>
<td>7.7±5.9 a</td>
<td>3.4±5.9 a</td>
</tr>
</tbody>
</table>

* Percentage of total GLS.

* Percentage of total GLS catabolites.
Consistent with this observation, in the root exudate the ratio of catabolites to intact GLS is 4:1 while the ratio of catabolites to intact GLS for the exogenous 2-propenyl GLS is 1:4.

Cellular localization of the GLS synthesis enzymes CYP83A1 and CYP83B1 in the root

In order to define the rhizosecretion route of GLS from the site of synthesis to the rhizosphere, we first analysed the root cell type-specific localization of the fluorophore-tagged biosynthetic enzymes CYP83A1 and CYP83B1 representative of aliphatic and indolic GLS biosynthesis, respectively (Naur et al., 2003). We monitored fluorescence throughout a z-stack panel from the outer towards the inner layers of the root tissue. The CYP83A1-YFP signal was detected exclusively in the stele along the main root with relatively higher expression in the vasculature at the lateral root branch site (Fig. 2A–C). By contrast, the CYP83B1 signal was detected in both the outer and inner layers of root cells at both the main root and the lateral root branch sites (Fig. 2D–F). The stele-specific localization of aliphatic biosynthetic enzymes, combined with the presence of GLS in the rhizosphere, indicate that the transport route of aliphatic GLS from the site-of-synthesis to the rhizosphere requires a coupled trans-cellular transport pathway and/or symplastic pathway to pass the Casparian strip. By contrast, a trans-cellular independent, but symplastic transport-dependent pathway may exist for the rhizosecretion of indole GLS.

Role of GTR importers in the rhizosecretion of GLS

To explore the root exudation process of GLS, we investigated whether GTR1 and GTR2 play a role in this process. Previously, GTR1 and GTR2 have been shown to play a key role in the loading of GLS into seeds. In that study, there was no significant effect on GLS in seeds of the gtr1 single mutants and only about a 50% reduction in the gtr2 single mutants, whereas the gtr1 gtr2 double mutants had zero GLS left in seeds (Nour-Eldin et al., 2012). Due to redundancy in GTR1 and GTR2 function we have, in the present study, chosen to investigate gtr1 gtr2 double mutants to maximize any observed effect on root exudation. We first analysed the GLS distribution pattern in the rosette and roots of 6-week-old wild type and gtr1 gtr2 mutants. In gtr1 gtr2, we observed a dramatic reduction of aliphatic GLS in roots and an over-accumulation in the rosette (Supplementary Fig. S2; Table S1), in agreement with previous observations for 3-week-old hydroponically grown plants (Andersen et al., 2013). This over-accumulation of root-synthesized GLS in the rosette suggested that, in the wild type, root storage cells and/or the rhizosphere (and not the rosette) were the major sinks for those GLS.

When we measured the exudates from 6-week-old wild type and gtr1 gtr2, we found that exudates of gtr1 gtr2 root contained strongly reduced levels of particularly long-chained aliphatic GLS (Fig. 3; Table 2). The levels of 4MSB—the major short-chained aliphatic GLS—varied substantially between the different biological repeats, but was not significantly changed in gtr1 gtr2 compared with the wild type, as was also observed for indole GLS (Table 2). This showed that exudation of stele-synthesized, long-chained aliphatic GLS requires a GTR-mediated import process of GLS from the sites of synthesis to a symplastic domain for exudation.

Role of GTR importers in rhizosecretion of GLS catabolites

Catabolites of both short-chained and long-chained aliphatic GLS in root exudates in the gtr1 gtr2 mutants constituted 10% of the concentration in wild-type exudates (Fig. 4; Table 3).

Fig. 2. Propidium iodide-stained roots of 12-d-old Arabidopsis seedlings expressing ProCYP83A1:CYP83A1-mVenus (A–C) or ProCYP83B1:CYP83B1-mVenus (D–F). (A, D) Bright field channel. (B, E) Focused on the cortex layer of a lateral root branch point. (C, F) Focused on the stele tissue of a lateral root branch point. Scale bar: 100 μm.
Although GLS levels in the roots of gtr1 gtr2 themselves were also decreased, the decrease was more pronounced for the catabolites than for intact GLS which suggests that GTR-mediated import into, for example, myrosin cells is a critical step prior to hydrolysis and exudation. Puzzlingly, a major part of total aliphatic GLS catabolites in root exudates of gtr1 gtr2 mutants was from short-chained aliphatic GLS (Table 3) that was not detectable in the roots of gtr1 gtr2 (Supplementary Table S1) proposing a complete hydrolysis of short-chained GLS.
aliphatic GLS in the mutant. Catabolites of indole GLS were not significantly reduced in root exudates of gtr1 gtr2 mutants compared with the wild type (Fig. 4; Table 2).

**Cellular expression of GTR2 in the root**

In order to investigate the localization of GTR-mediated import for stele-synthesized aliphatic GLS, we analysed the cell type-specific expression of GTR2. In agreement with microarray and translome databases (Mustroph et al., 2009), a previous study indicated that GTR1 and GTR2 have similar tissue expression patterns in the root (Andersen et al., 2013). Thus, we used GTR2 reporter lines for further analysis. We first analysed the cellular expression patterns by promoter GUS fusion in Arabidopsis root (Nour-Eldin et al., 2012). Promoter activity was detected at the pericycle and inner cell layers in the stele of the elongation and maturation regions (Fig. 5A, B, arrows). Cross-sections of the root within the same region confirmed the stele accumulation and also indicated weak GTR2 expression in the cortical and epidermal cells (Fig. 5B, arrow heads). In the lateral root initiation region, GTR2 was expressed in lateral root founder cells (Fig. 5C, arrow heads), which are differentiated from specific pericycle cells from the xylem pole. At a later developmental stage of the lateral roots, the signal spread to the whole root primordia (stage III–V) and accumulated at later root branch sites (Fig. 5D). In the mature lateral root it was restricted to the vasculature and no signal was detected in the endodermal cells and root tip of the mature roots (Fig. 5A–E). Next, we analysed the cellular distribution of GTR2 protein using fluorescent protein fusion (Andersen et al., 2013). In agreement with the GUS signal, in the main root of Arabidopsis seedlings GTR2-mOrange signals were detected in the stele (Fig. 5F, G–I), of which cells next to pro-xylem cells (Fig. 5F, H) showed the strongest expression. The expression of the GTRs inside the stele (including the pericycle cells), and sporadically in the cortex, shows where the import of GLS occurs prior to exudation.

**Discussion**

**Rhizosecretion of GLS and their catabolites in Arabidopsis**

One prerequisite for using GLS in Arabidopsis as a model system for studying root exudation is that GLS are exuded and detectable. Accordingly, we developed a method that enables the detection of intact GLS and their catabolites simultaneously in root exudates from Arabidopsis. Previous studies indicated that other plants within the Brassicaceae family can exude both intact GLS and their catabolites (Schreiner et al., 2011; Ngala et al., 2015). Hitherto, in Arabidopsis, however, only GLS catabolites and GLS biosynthetic intermediates were identified in root exudates (Badri et al., 2013; Strehmel et al., 2014, 2016; Monchgesang et al., 2016). In our method, we collected root exudates from sand-grown plants in de-ionized water instead of nutrient medium which was previously used for hydroponically cultured plants (Badri et al., 2013; Strehmel et al., 2014). By replacing the artificial hydroponic growth conditions with sand, we reduced the hypoxia stress often seen in insufficiently aerated hydroponics. Furthermore, we collected the exudate from plants in de-ionized water

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**Fig. 5.** Cellular localization of GTR2 in the root. Transgenic Arabidopsis plants expressing a GTR2-mOrange fusion and GFP-GUS fusion under the control of a 2-kb promoter of GTR2 were analysed by confocal microscopy or GUS staining. (A) GUS signal shown in pericycle cells (white arrowheads) and the inner cell layers. (B) Cross-section of a root region similar to the one shown in (A). GUS signals in the stele (white arrowheads) and sporadically distributed in the cortex and epidermal cells (black arrowheads). (C) GUS signals in a lateral root initiation site (arrowheads showed lateral root founder cells). (D) GUS signals in a lateral root mature stage. (E) GUS signal in the main root tip. (F–I) mOrange signal from longitudinal optical sections along the x-y axis of intact roots of GTR2-mOrange. The mOrange signals are in green. (F) Z-stack of optical sections of intact roots of GTR2-mOrange. (G–I) Serial optical sections along the z-axis from the top to the bottom of the same region of root as in (F). The GTR2-mOrange was predominantly expressed in cells next to pro-xylem cells (F, H, white arrowheads) but not in pro-xylem cells (I, black arrowheads). Bars in (A, C–E and F–I)=25 μm. Bars in (B)=100 μm.
interactions.

is a tool for dissecting the role of GLS in plant–rhizosphere
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the defensive roles of GLS, the physiological significance of
(Hiruma et al., 2016). Despite substantial knowledge about
the defensive roles of GLS, the physiological significance of
their role in the rhizosphere is largely unknown. The measure-
ment of GLS in root exudates of the model plant Arabidopsis
is a tool for dissecting the role of GLS in plant–rhizosphere
interactions.

Import-dependent rhizosecretion of stele-synthesized metabolites

A key finding in this study is that GTR1- and GTR2-mediated import is critical for the rhizosecretion of stele-synthesized GLS. Intuitively, one would have anticipated that the rhizo-
secreted phytochemicals were produced in the outer cell layers of the root rather than in the central stele. The forma-
tion of GLS in the stele might benefit from the carbon supply from leaf-active primary metabolism via the phloem. Concurrently, the vascular-associated synthesis followed by export to the apoplast enables the efficient mobilization of defence compounds via the ascending xylem, in addition to loading on to the endodermis symplastically for exudation. This suggests that the GLS importers GTR1 and GTR2 are essential in orchestrating the flow of stele-synthesized GLS to balance above- and below-ground defence in response to environmental challenge. Many other defence compounds like nicotine (Morita et al., 2009), tropane alkaloids (Ziegler and Pacchini, 2008), and pyrrolizidine alkaloids (Hartmann and Ober, 2000) are synthesized in roots and stored in leaves. Many of these compounds are also involved in rhizosphere communication (Zhuang et al., 2013; Tsednee et al., 2014). This suggests that modulating specific transport activities (in our case, plasma-membrane importers) are essential for shaping the distribution pattern of the defence compounds at the organismal level, for example, between shoot, root, and rhizosphere.

In contrast to the stele-specific synthesis of long-chained aliphatic GLS, indole GLS are synthesized in both the stele
and cortex and their rhizosecretion occurs in a GTR1- and GTR2-independent manner (Figs 2, 3). As GTR1 and GTR2 show broad substrate specificity towards GLS (Nour-Eldin et al., 2012), the presence of indole GLS at similar levels in root exudates of gtr1 gtr2 and the wild type suggests that an additional and indole-specific GLS importer(s) exist which functions redundantly with GTR1 and GTR2.

The similar level of exuded 4MSB from gtr1 gtr2 mutant and wild-type roots (Fig. 4; Table 2) suggests that exudation of this glucosinolate is not dependent on GTR transport activity. Interestingly, 4MSB was detected inside cortical cells in a cell type-specific metabolic profiling of root cells (Moussaieff et al., 2013) and expression of 4MSB-specific biosynthesis genes, for example, CYP79F2 and FMOGS-OX1, includes cortical cells (Supplementary Fig. S3B) (Chen et al., 2003; Hruz et al., 2008; Li et al., 2011). Therefore, we suggest that root-synthesized 4MSB is continuously exported to the rhizosphere from cells outside the Casparian strip which agreed with the GLS distribution in canola roots (Brassica napus) by cell-specific sulphur analyses (McCully et al., 2008).

Exudation of catabolites of aliphatic GLS relies on GTR1 and GTR2 activity. A possible scenario is that the GTRs import aliphatic GLS to myrosin cells for hydrolysis prior to exudation. The expression pattern of the major root myrosinases TGG4 and TGG5 (Supplementary Fig. S3C, D) overlaps with that of GTR1 and GTR2 in the cortex (data not shown and Fig. 5B). This supports the loading of GLS into myrosin cells by GTRs.

In summary, our data indicate that two import processes are critical for rhizosecretion: (i) the import of stele-synthesized long-chained aliphatic GLS into an endodermis-connected symplastic domain, and (ii) the import of short-chained aliphatic GLS from the apoplast in the cortex into myrosin cells (Fig. 6). This is an important first step in understanding the root exudation process. Future studies will reveal how intact GLS and their catabolites are exported from root cells.

Supplementary data

Supplementary data can be found at JXB online.

Figure S1. Sampling and profiling of GLS and their catabolites in Arabidopsis root exudates.

Figure S2. GLS concentration of rosettes and roots of 6-week-old sand-grown Arabidopsis wild type (Col-0) and gtr1 gtr2.

Figure S3. Cell type-specific expression of MAMJ, FMOGS-OX1, TGG4, and TGG5 in Arabidopsis roots.

Table S1. GLS in the leaf and root of Arabidopsis Col-0 and the gtr1 gtr2 mutant.

Table S2. GLS catabolites in the leaf and root of Arabidopsis.

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References


